

Chapter 8

References

References

1. Khanam, H., A. Ali, and M.J.E.j.o.m.c. Asif, *Neurodegenerative diseases linked to misfolded proteins and their therapeutic approaches: a review*. 2016. **124**: p. 1121-1141.
2. Elahi, F.M. and B.L.J.N.R.N. Miller, *A clinicopathological approach to the diagnosis of dementia*. 2017. **13**(8): p. 457-476.
3. Gan, L., et al., *Converging pathways in neurodegeneration, from genetics to mechanisms*. 2018. **21**(10): p. 1300-1309.
4. Schapira, A.H. and P. Jenner, *Etiology and pathogenesis of Parkinson's disease*. *Movement disorders*, 2011. **26**(6): p. 1049-1055.
5. Gordon, P.H., *Amyotrophic lateral sclerosis*. *CNS drugs*, 2011. **25**(1): p. 1-15.
6. Gordon, P.H., *Amyotrophic lateral sclerosis: an update for 2013 clinical features, pathophysiology, management and therapeutic trials*. *Aging and disease*, 2013. **4**(5): p. 295.
7. Nana, A.L., et al., *Widespread heterogeneous neuronal loss across the cerebral cortex in Huntington's disease*. *Journal of Huntington's disease*, 2014. **3**(1): p. 45-64.
8. Lunn, M.R. and C.H. Wang, *Spinal muscular atrophy*. *The Lancet*, 2008. **371**(9630): p. 2120-2133.
9. Butterfield, D.A., et al., *Evidence of oxidative damage in Alzheimer's disease brain: central role for amyloid β -peptide*. *Trends in molecular medicine*, 2001. **7**(12): p. 548-554.
10. Lisa Prostack, E.B., Pnina Yaish and Dorit Zharhar, *β Secretase (BACE1) Activity Assay Kit*:

A FRET Based Assay Designed for BACE1 Inhibitor Screening.

11. Sun, X., W.D. Chen, and Y.D. Wang, *beta-Amyloid: the key peptide in the pathogenesis of Alzheimer's disease*. Front Pharmacol, 2015. **6**: p. 221.
12. Mohandas, E., V. Rajmohan, and B. Raghunath, *Neurobiology of Alzheimer's disease*. Indian J Psychiatry, 2009. **51**(1): p. 55-61.
13. Terry, A.V., Jr. and J.J. Buccafusco, *The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development*. J Pharmacol Exp Ther, 2003. **306**(3): p. 821-7.
14. Hampel, H., et al., *The cholinergic system in the pathophysiology and treatment of Alzheimer's disease*. Brain, 2018. **141**(7): p. 1917-1933.
15. Chen, X.Q. and W.C. Mobley, *Exploring the Pathogenesis of Alzheimer Disease in Basal Forebrain Cholinergic Neurons: Converging Insights From Alternative Hypotheses*. Front Neurosci, 2019. **13**: p. 446.
16. Francis, P.T., et al., *The cholinergic hypothesis of Alzheimer's disease: a review of progress*. J Neurol Neurosurg Psychiatry, 1999. **66**(2): p. 137-47.
17. Contestabile, A., *The history of the cholinergic hypothesis*. Behav Brain Res, 2011. **221**(2): p. 334-40.
18. Kocahan, S. and Z. Dogan, *Mechanisms of Alzheimer's Disease Pathogenesis and Prevention: The Brain, Neural Pathology, N-methyl-D-aspartate Receptors, Tau Protein and Other Risk Factors*. Clin Psychopharmacol Neurosci, 2017. **15**(1): p. 1-8.
19. Olney, J.W., D.F. Wozniak, and N.B. Farber, *Excitotoxic neurodegeneration in Alzheimer disease. New hypothesis and new therapeutic strategies*. Arch Neurol, 1997. **54**(10): p. 1234-40.
20. Huang, W.J., X. Zhang, and W.W. Chen, *Role of oxidative stress in Alzheimer's disease*. Biomed Rep, 2016. **4**(5): p. 519-522.

21. Pratico, D., *Oxidative stress hypothesis in Alzheimer's disease: a reappraisal*. Trends Pharmacol Sci, 2008. **29**(12): p. 609-15.
22. Christen, Y., *Oxidative stress and Alzheimer disease*. Am J Clin Nutr, 2000. **71**(2): p. 621S-629S.
23. Strittmatter, W.J. and A.D. Roses, *Apolipoprotein E and Alzheimer disease*. Proc Natl Acad Sci U S A, 1995. **92**(11): p. 4725-7.
24. Han, S.D. and M.W. Bondi, *Revision of the apolipoprotein E compensatory mechanism recruitment hypothesis*. Alzheimers Dement, 2008. **4**(4): p. 251-4.
25. Potter, H. and T. Wisniewski, *Apolipoprotein e: essential catalyst of the Alzheimer amyloid cascade*. Int J Alzheimers Dis, 2012. **2012**: p. 489428.
26. Tuminello, E.R. and S.D. Han, *The apolipoprotein e antagonistic pleiotropy hypothesis: review and recommendations*. Int J Alzheimers Dis, 2011. **2011**: p. 726197.
27. Toral-Rios, D., et al., *GSK3beta and Tau Protein in Alzheimer's Disease and Epilepsy*. Front Cell Neurosci, 2020. **14**: p. 19.
28. Hooper, C., R. Killick, and S. Lovestone, *The GSK3 hypothesis of Alzheimer's disease*. J Neurochem, 2008. **104**(6): p. 1433-9.
29. Marucci, G., et al., *Efficacy of acetylcholinesterase inhibitors in Alzheimer's disease*. 2021. **190**: p. 108352.
30. English, C.J.M.H.C., *Donepezil 23 mg: Is it more advantageous compared to the original?* 2012. **1**(11): p. 272-273.
31. Bajda, M., et al., *Structure-based search for new inhibitors of cholinesterases*. 2013. **14**(3): p. 5608-5632.
32. Colletier, J.P., et al., *Structural insights into substrate traffic and inhibition in acetylcholinesterase*. 2006. **25**(12): p. 2746-2756.

33. Colovic, M.B., et al., *Acetylcholinesterase inhibitors: pharmacology and toxicology*. 2013. **11**(3): p. 315-335.
34. Hendrix, S., et al., *Post hoc evidence for an additive effect of memantine and donepezil: consistent findings from DOMINO-AD study and memantine clinical trial program*. 2015. **2**(3): p. 165-171.
35. Cui, N., et al., *Biochemical and biological attributes of matrix metalloproteinases*. 2017. **147**: p. 1-73.
36. Cabral-Pacheco, G.A., et al., *The roles of matrix metalloproteinases and their inhibitors in human diseases*. 2020. **21**(24): p. 9739.
37. Swetha, R., et al., *Biomolecular basis of matrix metallo proteinase-9 activity*. 2018. **10**(9): p. 1093-1112.
38. Vandooren, J., et al., *Biochemistry and molecular biology of gelatinase B or matrix metalloproteinase-9 (MMP-9): the next decade*. 2013. **48**(3): p. 222-272.
39. Fabre, B., A. Ramos, and B.J.J.o.m.c. de Pascual-Teresa, *Targeting matrix metalloproteinases: exploring the dynamics of the S1' pocket in the design of selective, small molecule inhibitors: miniperspective*. 2014. **57**(24): p. 10205-10219.
40. Adhikari, N., et al., *Robust design of some selective matrix metalloproteinase-2 inhibitors over matrix metalloproteinase-9 through in silico/fragment-based lead identification and de novo lead modification: Syntheses and biological assays*. 2016. **24**(18): p. 4291-4309.
41. Nicolotti, O., et al., *Design, synthesis and biological evaluation of 5-hydroxy, 5-substituted-pyrimidine-2, 4, 6-triones as potent inhibitors of gelatinases MMP-2 and MMP-9*. 2012. **58**: p. 368-376.

-
42. Nuti, E., et al., *N-O-Isopropyl sulfonamido-based hydroxamates as matrix metalloproteinase inhibitors: Hit selection and in vivo antiangiogenic activity*. 2015. **58**(18): p. 7224-7240.
 43. Shimizu, H., et al., *Crystal structure of an active form of BACE1, an enzyme responsible for amyloid β protein production*. 2008. **28**(11): p. 3663-3671.
 44. Butini, S., et al., *The structural evolution of β -secretase inhibitors: a focus on the development of small-molecule inhibitors*. 2013. **13**(15): p. 1787-1807.
 45. Costanzo, P., et al., *Design, synthesis, and evaluation of donepezil-like compounds as AChE and BACE-1 inhibitors*. 2016. **7**(5): p. 470-475.
 46. Keravis, T. and C.J.B.j.o.p. Lugnier, *Cyclic nucleotide phosphodiesterase (PDE) isozymes as targets of the intracellular signalling network: benefits of PDE inhibitors in various diseases and perspectives for future therapeutic developments*. 2012. **165**(5): p. 1288-1305.
 47. Heckman, P., et al., *Phosphodiesterase inhibition and modulation of corticostriatal and hippocampal circuits: clinical overview and translational considerations*. 2018. **87**: p. 233-254.
 48. Swetha, R., et al., *Combined ligand-based and structure-based design of PDE 9A inhibitors against Alzheimer's disease*. 2022. **26**(5): p. 2877-2892.
 49. Wu, Y., et al., *Novel phosphodiesterase inhibitors for cognitive improvement in Alzheimer's Disease: Miniperspective*. 2018. **61**(13): p. 5467-5483.
 50. Hutson, P., et al., *The selective phosphodiesterase 9 (PDE9) inhibitor PF-04447943 (6-[(3S, 4S)-4-methyl-1-(pyrimidin-2-ylmethyl) pyrrolidin-3-yl]-1-(tetrahydro-2H-pyran-4-yl)-1, 5-dihydro-4H-pyrazolo [3, 4-d] pyrimidin-4-one) enhances synaptic plasticity and cognitive function in rodents*. 2011. **61**(4): p. 665-676.

-
51. Morphy, R., C. Kay, and Z.J.D.d.t. Rankovic, *From magic bullets to designed multiple ligands*. 2004. **9**(15): p. 641-651.
 52. Melchiorre, C., et al., *Acetylcholinesterase noncovalent inhibitors based on a polyamine backbone for potential use against Alzheimer's disease*. 1998. **41**(22): p. 4186-4189.
 53. Benek, O., J. Korabecny, and O.J.T.i.P.S. Soukup, *A perspective on multi-target drugs for Alzheimer's disease*. 2020. **41**(7): p. 434-445.
 54. Zhou, J., et al., *Rational design of multitarget-directed ligands: strategies and emerging paradigms*. 2019. **62**(20): p. 8881-8914.
 55. Kondej, M., P. Stepnicki, and A.A.J.I.j.o.m.s. Kaczor, *Multi-target approach for drug discovery against schizophrenia*. 2018. **19**(10): p. 3105.
 56. Prati, F., A. Cavalli, and M.L.J.M. Bolognesi, *Navigating the chemical space of multitarget-directed ligands: from hybrids to fragments in Alzheimer's disease*. 2016. **21**(4): p. 466.
 57. Oliveira Pedrosa, M.d., et al., *Hybrid compounds as direct multitarget ligands: a review*. 2017. **17**(9): p. 1044-1079.
 58. Bajda, M., et al., *Multi-target-directed ligands in Alzheimer's disease treatment*. 2011. **18**(32): p. 4949-4975.
 59. Knowles, T.P., M. Vendruscolo, and C.M. Dobson, *The amyloid state and its association with protein misfolding diseases*. *Nature reviews Molecular cell biology*, 2014. **15**(6): p. 384.
 60. Savelieff, M.G., et al., *Untangling amyloid- β , tau, and metals in Alzheimer's disease*. *ACS chemical biology*, 2013. **8**(5): p. 856-865.

61. Rajasekhar, K., K. Mehta, and T. Govindaraju, *Hybrid Multifunctional Modulators Inhibit Multifaceted A β Toxicity and Prevent Mitochondrial Damage*. ACS chemical neuroscience, 2018. **9**(6): p. 1432-1440.
62. Yong, V.W., et al., *Matrix metalloproteinases and diseases of the CNS*. Trends in neurosciences, 1998. **21**(2): p. 75-80.
63. Rosenberg, G.A., *Matrix metalloproteinases and their multiple roles in neurodegenerative diseases*. The Lancet Neurology, 2009. **8**(2): p. 205-216.
64. Lorenzl, S., et al., *Tissue inhibitors of matrix metalloproteinases are elevated in cerebrospinal fluid of neurodegenerative diseases*. Journal of the neurological sciences, 2003. **207**(1): p. 71-76.
65. Brkic, M., et al., *Amyloid β Oligomers Disrupt Blood–CSF Barrier Integrity by Activating Matrix Metalloproteinases*. The Journal of Neuroscience, 2015. **35**(37): p. 12766-12778.
66. Swetha, R., et al., *Biomolecular basis of matrix metallo proteinase-9 activity*. Future medicinal chemistry, 2018. **10**(9): p. 1093-1112.
67. Saravanan, C. and S.K. Singh, *Status of research on MMPs in India*. Expert opinion on therapeutic targets, 2011. **15**(6): p. 715-728.
68. Kumar, D., et al., *Curcumin: a potential candidate for matrix metalloproteinase inhibitors*. Expert opinion on therapeutic targets, 2012. **16**(10): p. 959-972.
69. Nordberg, A., et al., *A review of butyrylcholinesterase as a therapeutic target in the treatment of Alzheimer's disease*. 2013. **15**(2).
70. Unzeta, M., et al., *Multi-target directed donepezil-like ligands for Alzheimer's disease*. 2016. **10**: p. 205.
71. Huggins, D.J., W. Sherman, and B.J.J.o.m.c. Tidor, *Rational approaches to improving selectivity in drug design*. 2012. **55**(4): p. 1424-1444.

-
72. Wei, D., et al., *Discovery of multitarget inhibitors by combining molecular docking with common pharmacophore matching*. 2008. **51**(24): p. 7882-7888.
73. Prati, F., A. Cavalli, and M.J.M. Bolognesi, *Navigating the chemical space of multitarget-directed ligands: From hybrids to fragments in Alzheimer's disease*. 2016. **21**(4): p. 466.
74. Cavalli, A., et al., *Multi-target-directed ligands to combat neurodegenerative diseases*. 2008. **51**(3): p. 347-372.
75. Zemek, F., et al., *Outcomes of Alzheimer's disease therapy with acetylcholinesterase inhibitors and memantine*. 2014. **13**(6): p. 759-774.
76. Panek, D., et al., *Design, synthesis, and biological evaluation of 1-benzylamino-2-hydroxyalkyl derivatives as new potential disease-modifying multifunctional anti-Alzheimer's agents*. 2018. **9**(5): p. 1074-1094.
77. Prati, F., et al., *Novel 8-Hydroxyquinoline Derivatives as Multitarget Compounds for the Treatment of Alzheimer's Disease*. 2016. **11**(12): p. 1284-1295.
78. Więckowska, A., et al., *Novel multitarget-directed ligands aiming at symptoms and causes of Alzheimer's disease*. 2018. **9**(5): p. 1195-1214.
79. Kumar, D., et al., *Development of Piperazinediones as dual inhibitor for treatment of Alzheimer's disease*. *European Journal of Medicinal Chemistry*, 2018. **150**: p. 87-101.
80. Kumar, D., et al., *Biological profiling of piperazinediones for the management of anxiety*. *Pharmacology Biochemistry and Behavior*, 2019. **176**: p. 63-71.
81. Friesner, R.A., et al., *Glide: a new approach for rapid, accurate docking and scoring. I. Method and assessment of docking accuracy*. *Journal of medicinal chemistry*, 2004. **47**(7): p. 1739-1749.
82. Rajasekhar, K., C. Madhu, and T. Govindaraju, *Natural tripeptide-based inhibitor of multifaceted amyloid β toxicity*. *ACS chemical neuroscience*, 2016. **7**(9): p. 1300-1310.

-
83. Jan, A., D.M. Hartley, and H.A. Lashuel, *Preparation and characterization of toxic A β aggregates for structural and functional studies in Alzheimer's disease research*. Nature protocols, 2010. **5**(6): p. 1186.
84. Vyas, N.A., et al., *Acetylcholinesterase and A β Aggregation Inhibition by Heterometallic Ruthenium (II)–Platinum (II) Polypyridyl Complexes*. Inorganic chemistry, 2018.
85. Satheeshkumar, S., et al., *Design, Synthesis and Biological Evaluation of Carbazole Derivatives as Antitubercular and Antibacterial Agents*. Current Bioactive Compounds, 2018. **14**: p. 1-15.
86. Revilla, S., et al., *Lenti-GDNF gene therapy protects against Alzheimer's disease-like neuropathology in 3xTg-AD mice and MC65 cells*. CNS neuroscience & therapeutics, 2014. **20**(11): p. 961-972.
87. Modi, G., et al., *Structural Modifications of Neuroprotective Anti-Parkinsonian (-)-N 6-(2-(4-(Biphenyl-4-yl) piperazin-1-yl)-ethyl)-N 6-propyl-4, 5, 6, 7-tetrahydrobenzo [d] thiazole-2, 6-diamine (D-264): an Effort toward the Improvement of in Vivo Efficacy of the Parent Molecule*. Journal of medicinal chemistry, 2014. **57**(4): p. 1557-1572.
88. Shidore, M., et al., *Benzylpiperidine-Linked Diarylthiazoles as Potential Anti-Alzheimer's Agents: Synthesis and Biological Evaluation*. Journal of medicinal chemistry, 2016. **59**(12): p. 5823-5846.
89. Ganeshpurkar, A., D. Kumar, and S.K. Singh, *Design, synthesis and collagenase inhibitory activity of some novel phenylglycine derivatives as metalloproteinase inhibitors*. International journal of biological macromolecules, 2018. **107**: p. 1491-1500.

-
90. Umre, R., et al., *In vitro, in vivo and in silico antiulcer activity of ferulic acid*. Future Journal of Pharmaceutical Sciences, 2018. **4**(2): p. 248-253.
 91. Lee, S., et al., *Untangling Amyloid-beta, Tau, and Metals in Alzheimer's Disease*. 2013.
 92. Reybier, K., et al., *Free Superoxide is an Intermediate in the Production of H₂O₂ by Copper (I)-A β Peptide and O₂*. Angewandte Chemie International Edition, 2016. **55**(3): p. 1085-1089.
 93. Lim, P., A. Huss Jr, and C. Eckert, *Oxidation of aqueous sulfur dioxide. 3. The effects of chelating agents and phenolic antioxidants*. The Journal of Physical Chemistry, 1982. **86**(21): p. 4233-4237.
 94. Atmaca, G., *Antioxidant effects of sulfur-containing amino acids*. Yonsei medical journal, 2004. **45**: p. 776-788.
 95. Mensch, J., et al., *In vivo, in vitro and in silico methods for small molecule transfer across the BBB*. 2009. **98**(12): p. 4429-4468.
 96. Fiorito, J., et al., *Identification of a novel 1, 2, 3, 4-tetrahydrobenzo [b][1, 6] naphthyridine analogue as a potent phosphodiesterase 5 inhibitor with improved aqueous solubility for the treatment of Alzheimer's disease*. 2017. **60**(21): p. 8858-8875.
 97. Prickaerts, J., P.R.A. Heckman, and A. Blokland, *Investigational phosphodiesterase inhibitors in phase I and phase II clinical trials for Alzheimer's disease*. Expert Opin Investig Drugs, 2017. **26**(9): p. 1033-1048.
 98. Nabavi, S.M., et al., *Phosphodiesterase inhibitors say NO to Alzheimer's disease*. Food Chem Toxicol, 2019. **134**: p. 110822.
 99. Liu, S., et al., *Structural basis for the catalytic mechanism of human phosphodiesterase 9*. Proc Natl Acad Sci U S A, 2008. **105**(36): p. 13309-14.

-
100. Feneck, R., *Phosphodiesterase inhibitors and the cardiovascular system*. Continuing Education in Anaesthesia Critical Care & Pain, 2008. **8**(2).
 101. Salter, E.A. and A. Wierzbicki, *The mechanism of cyclic nucleotide hydrolysis in the phosphodiesterase catalytic site*. J Phys Chem B, 2007. **111**(17): p. 4547-52.
 102. Miller, M., *Phosphodiesterase inhibition in the treatment of autoimmune and inflammatory diseases: current status and potential*. Journal of Receptor, Ligand and Channel Research, 2014. **volume 8**: p. 19-30.
 103. Heckman, P.R., C. Wouters, and J. Prickaerts, *Phosphodiesterase inhibitors as a target for cognition enhancement in aging and Alzheimer's disease: a translational overview*. Curr Pharm Des, 2015. **21**(3): p. 317-31.
 104. Garcia-Osta, A., et al., *Phosphodiesterases as therapeutic targets for Alzheimer's disease*. ACS Chem Neurosci, 2012. **3**(11): p. 832-44.
 105. Kleiman, R.J., et al., *Phosphodiesterase 9A regulates central cGMP and modulates responses to cholinergic and monoaminergic perturbation in vivo*. Journal of Pharmacology and Experimental Therapeutics, 2012. **341**(2): p. 396-409.
 106. Voet, A., et al., *Pharmacophore modeling: advances, limitations, and current utility in drug discovery*. Journal of Receptor, Ligand and Channel Research, 2014.
 107. Daisy, P., et al., *A database for the predicted pharmacophoric features of medicinal compounds*. Bioinformation, 2011. **6**(4): p. 167-8.
 108. Carpenter, T.S., et al., *A method to predict blood-brain barrier permeability of drug-like compounds using molecular dynamics simulations*. Biophys J, 2014. **107**(3): p. 630-641.
 109. Jasial, S., Y. Hu, and J. Bajorath, *How Frequently Are Pan-Assay Interference Compounds Active? Large-Scale Analysis of Screening Data Reveals Diverse Activity*

-
- Profiles, Low Global Hit Frequency, and Many Consistently Inactive Compounds.* J Med Chem, 2017. **60**(9): p. 3879-3886.
110. Baell, J.B. and G.A. Holloway, *New substructure filters for removal of pan assay interference compounds (PAINS) from screening libraries and for their exclusion in bioassays.* J Med Chem, 2010. **53**(7): p. 2719-40.
111. Baell, J.B. and J.W.M. Nissink, *Seven Year Itch: Pan-Assay Interference Compounds (PAINS) in 2017-Utility and Limitations.* ACS Chem Biol, 2018. **13**(1): p. 36-44.
112. Hollingsworth, S.A. and R.O. Dror, *Molecular Dynamics Simulation for All.* Neuron, 2018. **99**(6): p. 1129-1143.
113. Ivanova, L., et al., *Molecular Dynamics Simulations of the Interactions between Glial Cell Line-Derived Neurotrophic Factor Family Receptor GFRalpha1 and Small-Molecule Ligands.* ACS Omega, 2018. **3**(9): p. 11407-11414.
114. Singh, R., et al., *Identifying potential GluN2B subunit containing N-Methyl-D-aspartate receptor inhibitors: An integrative in silico and molecular modeling approach.* Journal of Biomolecular Structure and Dynamics, 2020. **38**(9): p. 2533-2545.
115. Ganeshpurkar, A., et al., *Structure-based screening and molecular dynamics simulation studies for the identification of potential acetylcholinesterase inhibitors.* Molecular Simulation, 2019. **46**(3): p. 169-185.
116. Daneman, R. and A. Prat, *The blood-brain barrier.* Cold Spring Harb Perspect Biol, 2015. **7**(1): p. a020412.
117. Zhang, M.Q. and B. Wilkinson, *Drug discovery beyond the 'rule-of-five'.* Curr Opin Biotechnol, 2007. **18**(6): p. 478-88.
118. Morris, G.M., R. Huey, and A.J. Olson, *Using AutoDock for ligand-receptor docking.* Curr Protoc Bioinformatics, 2008. **Chapter 8**: p. Unit 8 14.

-
119. El-Hachem, N., et al., *AutoDock and AutoDockTools for Protein-Ligand Docking: Beta-Site Amyloid Precursor Protein Cleaving Enzyme 1(BACE1) as a Case Study*. *Methods Mol Biol*, 2017. **1598**: p. 391-403.
120. Vora, J., et al., *Molecular docking, QSAR and ADMET based mining of natural compounds against prime targets of HIV*. *J Biomol Struct Dyn*, 2019. **37**(1): p. 131-146.
121. Zhang, P., et al., *Design, synthesis and evaluation of pyrazolopyrimidinone derivatives as novel PDE9A inhibitors for treatment of Alzheimer's disease*. *Bioorg Med Chem Lett*, 2020. **30**(14): p. 127254.
122. Opo, F., et al., *Structure based pharmacophore modeling, virtual screening, molecular docking and ADMET approaches for identification of natural anti-cancer agents targeting XIAP protein*. *Sci Rep*, 2021. **11**(1): p. 4049.
123. Sandeep, G., et al., *AUDocker LE: A GUI for virtual screening with AUTODOCK Vina*. *BMC Res Notes*, 2011. **4**: p. 445.
124. Jaghoori, M.M., B. Bleijlevens, and S.D. Olabarriaga, *1001 Ways to run AutoDock Vina for virtual screening*. *J Comput Aided Mol Des*, 2016. **30**(3): p. 237-49.
125. Quiroga, R. and M.A. Villarreal, *Vinardo: A Scoring Function Based on Autodock Vina Improves Scoring, Docking, and Virtual Screening*. *PLoS One*, 2016. **11**(5): p. e0155183.
126. Trott, O. and A.J. Olson, *AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading*. *J Comput Chem*, 2010. **31**(2): p. 455-61.
127. Guan, L., et al., *ADMET-score - a comprehensive scoring function for evaluation of chemical drug-likeness*. *Medchemcomm*, 2019. **10**(1): p. 148-157.

-
128. Ya'u Ibrahim, Z., et al., *Molecular docking studies, drug-likeness and in-silico ADMET prediction of some novel β -Amino alcohol grafted 1,4,5-trisubstituted 1,2,3-triazoles derivatives as elevators of p53 protein levels*. Scientific African, 2020. **10**.
129. Viana Nunes, A.M., et al., *preADMET analysis and clinical aspects of dogs treated with the Organotellurium compound RF07: A possible control for canine visceral leishmaniasis?* Environ Toxicol Pharmacol, 2020. **80**: p. 103470.
130. Yan, A., Z. Wang, and Z. Cai, *Prediction of human intestinal absorption by GA feature selection and support vector machine regression*. Int J Mol Sci, 2008. **9**(10): p. 1961-76.
131. Singh, R., et al., *Identifying potential GluN2B subunit containing N-Methyl-D-aspartate receptor inhibitors: an integrative in silico and molecular modeling approach*. J Biomol Struct Dyn, 2020. **38**(9): p. 2533-2545.
132. Hospital, A., et al., *Molecular dynamics simulations: advances and applications*. Adv Appl Bioinform Chem, 2015. **8**: p. 37-47.
133. Ai, R., M. Qaiser Fatmi, and C.E. Chang, *T-Analyst: a program for efficient analysis of protein conformational changes by torsion angles*. J Comput Aided Mol Des, 2010. **24**(10): p. 819-27.
134. Chen, Z. and C.J.P.i.n. Zhong, *Decoding Alzheimer's disease from perturbed cerebral glucose metabolism: implications for diagnostic and therapeutic strategies*. 2013. **108**: p. 21-43.
135. Coimbra, J.R., et al., *Highlights in BACE1 inhibitors for Alzheimer's disease treatment*. 2018. **6**: p. 178.
136. Das, B. and R.J.C.d. Yan, *A close look at BACE1 inhibitors for Alzheimer's disease treatment*. 2019. **33**(3): p. 251-263.

-
137. Swetha, R., et al., *Multifunctional hybrid sulfonamides as novel therapeutic agents for Alzheimer's disease*. 2019. **11**(24): p. 3161-3178.
138. Ganeshpurkar, A., et al., *Identification of sulfonamide based butyrylcholinesterase inhibitors through scaffold hopping approach*. 2022. **203**: p. 195-211.
139. Di, L., et al., *High throughput artificial membrane permeability assay for blood–brain barrier*. *European journal of medicinal chemistry*, 2003. **38**(3): p. 223-232.
140. Gutti, G., et al., *Discovery of novel series of 2-substituted benzo [d] oxazol-5-amine derivatives as multi-target directed ligands for the treatment of Alzheimer's disease*. 2019. **182**: p. 111613.
141. Saxena, G., et al., *Gugulipid, an extract of Commiphora whightii with lipid-lowering properties, has protective effects against streptozotocin-induced memory deficits in mice*. *Pharmacology Biochemistry and Behavior*, 2007. **86**(4): p. 797-805.
142. Ellman, G.L., et al., *A new and rapid colorimetric determination of acetylcholinesterase activity*. *Biochemical pharmacology*, 1961. **7**(2): p. 88IN191-9095.
143. Lee, M.-R., et al., *Anti-amnesic effect of Chong–Myung–Tang on scopolamine-induced memory impairments in mice*. *Journal of ethnopharmacology*, 2010. **132**(1): p. 70-74.
144. Ganeshpurkar, A., et al., *Effect of sulfonamide derivatives of phenylglycine on scopolamine-induced amnesia in rats*. 2023. **9**(1): p. 13-31.
145. Baldissera, M.D., et al., *Nerolidol-loaded nanospheres prevent behavioral impairment via ameliorating Na⁺, K⁺-ATPase and AChE activities as well as reducing oxidative stress in the brain of Trypanosoma evansi-infected mice*. 2017. **390**: p. 139-148.

Appendix

LD₅₀ of *N*-benzyl piperazinyl sulfonamide derivatives**LD₅₀ determination of compound 72****Table A0.1** LD₅₀ determination protocol for compound 72

| | |
|---|----------------------------|
| Test substance | |
| 1. Physical nature | Solid |
| 2. Code | 72 |
| Vehicle | 0.5% CMC solution in water |
| Test animals | Rat |
| 1. Sex | Female |
| 2. Number | 3 |
| Test conditions | |
| 1. Dose | 300 mg/kg |
| 2. Rationale for the selection of the starting dose | |
| 3. Dosing volumes | 0.7 ml |
| 4. Time & date of dosing | 12:30 pm 14/01/2023 |

Table T3 Effect of compound 72 on the body wt. of the animals at the dose of 300 mg/kg.

| Group | Body wt. (gm) on 14/01/2023 at 12:20 pm | Body wt. (gm) on 15/01/2023 at 12:20 pm | Body wt. (gm) on 16/01/2023 at 12:20 pm | Body wt. (gm) on 17/01/2023 at 12:20 pm | Body wt. (gm) on 18/01/2023 at 12:20 pm |
|-------|---|---|---|---|---|
| 1 | 215 | 214 | 207 | 217 | 228 |
| 2 | 228 | 223 | 230 | 239 | 244 |
| 3 | 235 | 234 | 225 | 238 | 247 |

Table T4 The onset of toxicity with compound 72 in the period of 72h.

| Group | Body wt. Changes (gm) | | | | | Onset of toxicity | Reversibility | Date & time of death |
|-------|-----------------------|---------|---------|----------|----------|-------------------|---------------|----------------------|
| | 14/1/23 | 15/1/23 | 16/1/23 | 17/01/23 | 18/01/23 | | | |
| 1 | 00 | 02 | 01 | 4 | 5 | - | - | - |
| 2 | 00 | 01 | 01 | 4 | 6 | - | - | - |
| 3 | 00 | 02 | 03 | 5 | 10 | - | - | - |

Discussion and interpretation of results: Animals were dosed as per the OECD guideline 423 at 300 mg/kg doses. All animals were alive at 300mg/kg dose within 72 hrs and next 14 days.

Conclusions: No toxicity was observed at **300mg/kg**.

Table A0.2 LD₅₀ determination protocol for the compound **72**.

| | |
|---|----------------------------|
| Test substance | |
| 1. Physical nature | Solid |
| 2. Code | 72 |
| Vehicle | 0.5% CMC solution in water |
| Test animals | Rat |
| 1. Sex | Female |
| 2. Number | 3 |
| Test conditions | |
| 1. Dose | 2000 mg/kg |
| 2. Rationale for the selection of the starting dose | No death at 300mg/kg |
| 3. Dosing volumes | 0.7 ml |
| 4. Time & date of dosing | 11:15 pm 19/01/2023 |

Table A0.3 Effect of compound **72** on the body wt. of the animals at the dose of 2000 mg/kg.

| Group | Body wt. (gm) on 19/01/2023 at 11 am | Body wt. (gm) on 20/01/2023 at 11 am | Body wt. (gm) on 21/01/2023 at 11 am |
|-------|---|---|---|
| 1 | 220 | - | - |
| 2 | 211 | - | - |
| 3 | 236 | - | - |

Table A0.4 The onset of toxicity with compound **72** in the period of 72h.

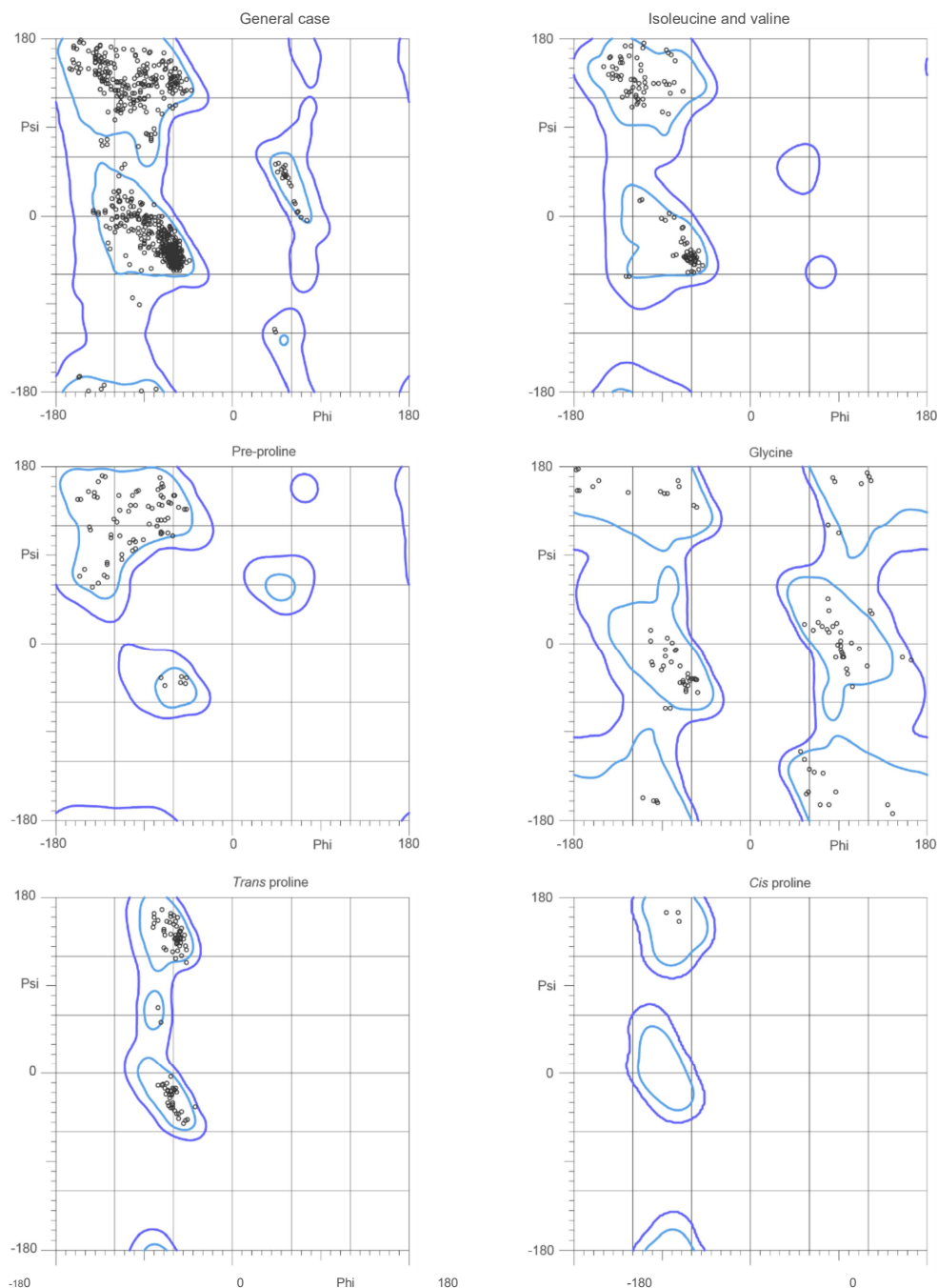
| Group | Body wt. Changes (gm) | | | Onset of toxicity | Reversibility | Date & time of death |
|-------|-----------------------|----------|----------|-------------------|---------------|----------------------|
| | 19/01/23 | 20/01/23 | 21/01/23 | | | |
| 1 | 0 | 0 | 0 | 19/01/23, 3:30 pm | No | 19/01/23, 7:43 pm |
| 2 | 0 | 0 | 0 | 19/01/23, 3:50 pm | No | 19/01/23, 6 pm |
| 3 | 0 | 0 | 0 | - | No | 20/01/23, 8:30 am |

Discussion and interpretation of results: Animals were dosed as per the OECD guideline 423 at 300 mg/kg and 2000 mg/kg doses. All animals died at 2000 mg/kg dose within 72 hrs.

Conclusions: As per OECD guideline $LD_{50} = 1000 \text{ mg/kg}$.

MolProbity Ramachandran analysis

4ey7.pdb, model 1



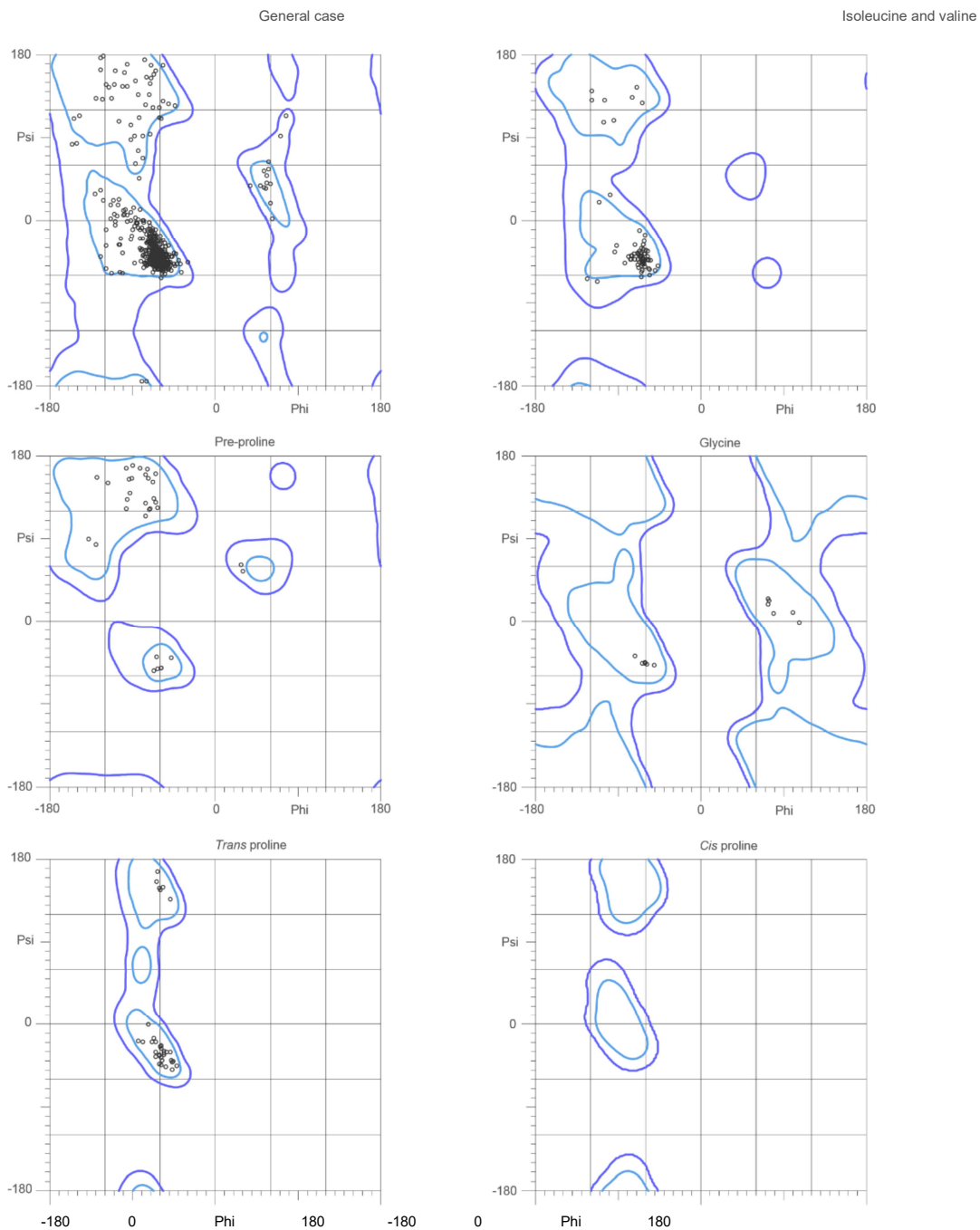
97.1% (1040/1071) of all residues were in favored (98%) regions.

100.0% (1071/1071) of all residues were in allowed (>99.8%) regions.

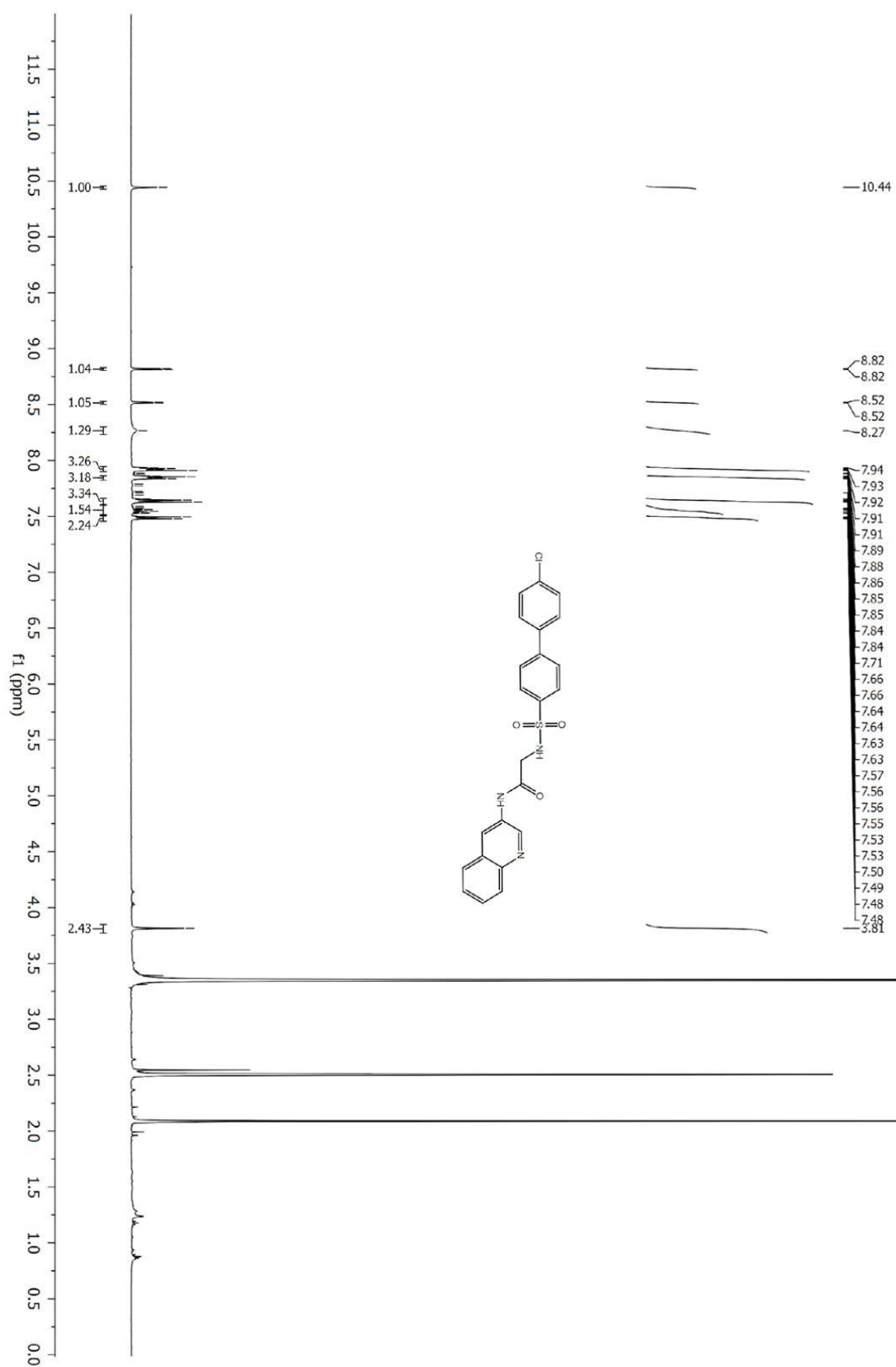
There were no outliers

Lovell, Davis, et al. Proteins 50:437 (2003)

MolProbity Ramachandran analysis 6a3n.pdb, model 1



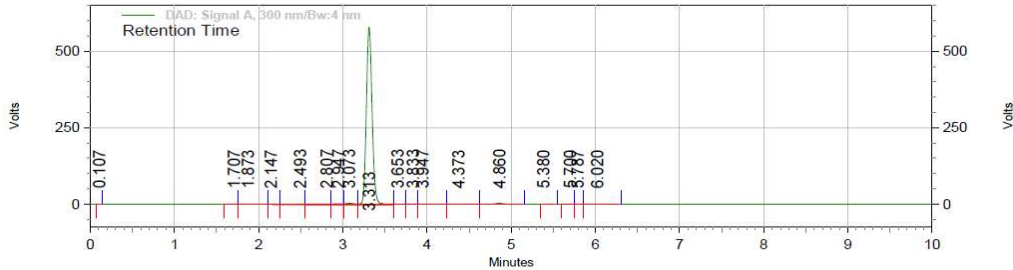
6.6% (618/640) of all residues were in favored (98%) regions.
100.0% (640/640) of all residues were in allowed (>99.8%) regions.
There were no outliers.
Lovell, Davis, et al. Proteins 50:437 (2003)

¹H NMR of Compound 41

HPLC data of Compound 41

Area % Report

Data File: C:\DATA\SKSINGH\RAYALA\COMPOUND-41 152018-11-03 12-02-33 (GMT +05-30).rslt\COMPOUND-41-125.dat
 Method: C:\Enterprise\Projects\Method\untitled.met
 Acquired: 03-11-2018 12:15:47 (GMT +05:30)
 Printed: 03-11-2018 12:52:10 (GMT +05:30)



DAD: Signal A, 300 nm/Bw:4 nm Results

| Retention Time | Area | Area % | Height | Height % |
|----------------|---------|--------|---------|----------|
| 0.107 | 48 | 0.00 | 20 | 0.00 |
| 1.707 | 2078 | 0.03 | 288 | 0.02 |
| 1.873 | 17681 | 0.29 | 780 | 0.06 |
| 2.147 | 10536 | 0.17 | 1196 | 0.10 |
| 2.493 | 33574 | 0.56 | 3592 | 0.29 |
| 2.807 | 33708 | 0.56 | 2311 | 0.19 |
| 2.947 | 28803 | 0.48 | 4516 | 0.36 |
| 3.073 | 39684 | 0.66 | 5587 | 0.45 |
| 3.313 | 5743115 | 95.39 | 1215865 | 97.56 |
| 3.653 | 15574 | 0.26 | 2111 | 0.17 |
| 3.833 | 15512 | 0.26 | 2162 | 0.17 |
| 3.947 | 27921 | 0.46 | 2355 | 0.19 |
| 4.373 | 15979 | 0.27 | 770 | 0.06 |
| 4.860 | 35263 | 0.59 | 4570 | 0.37 |
| 5.380 | 153 | 0.00 | 14 | 0.00 |
| 5.700 | 156 | 0.00 | 29 | 0.00 |
| 5.787 | 106 | 0.00 | 25 | 0.00 |
| 6.020 | 1053 | 0.02 | 101 | 0.01 |

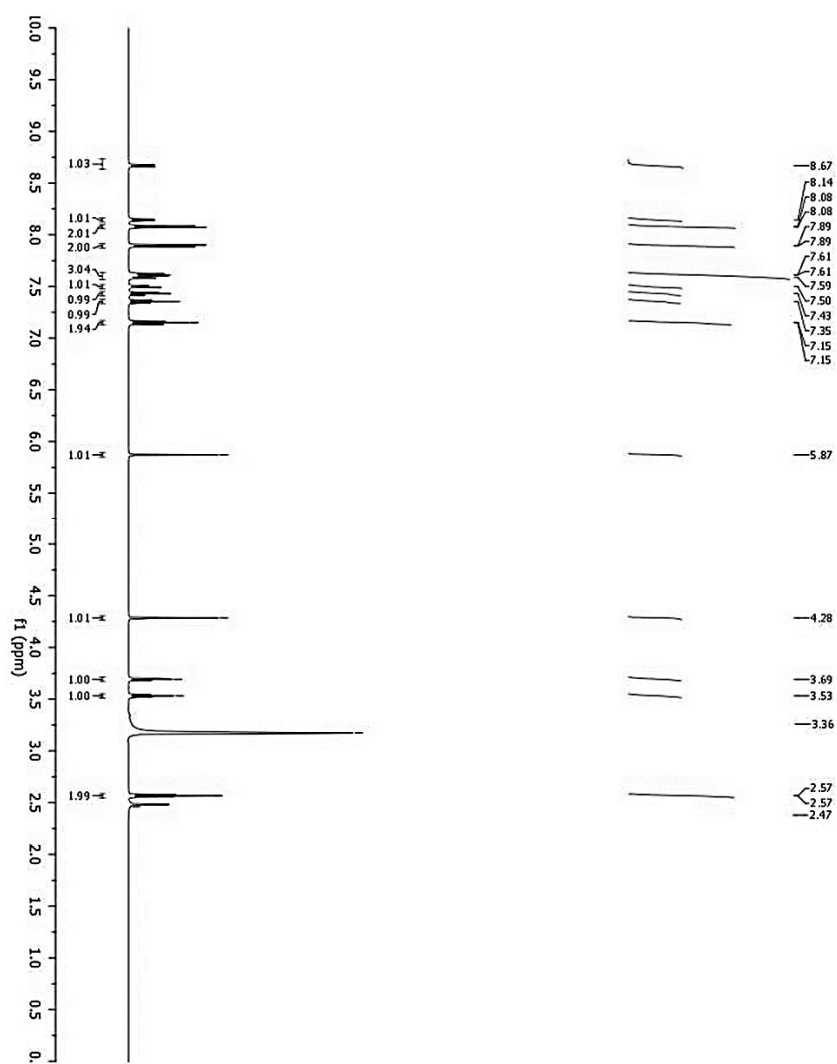
| | | | | |
|--------|---------|--------|---------|--------|
| Totals | 6020944 | 100.00 | 1246292 | 100.00 |
|--------|---------|--------|---------|--------|

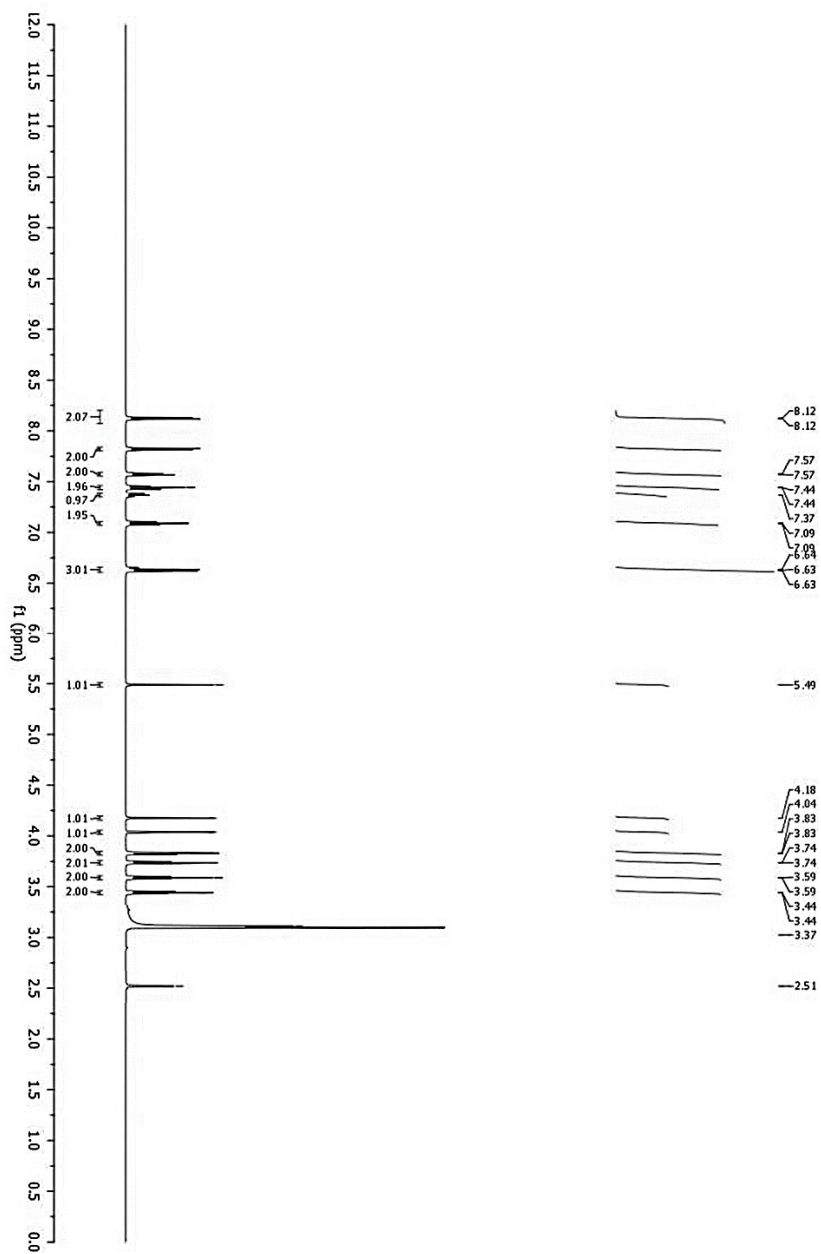
| Retention Time | Area | Area % | Height | Height % |
|----------------|---------|--------|---------|----------|
| 3.313 | 6087249 | 99.01 | 1227953 | 99.60 |
| 4.860 | 54922 | 0.89 | 4916 | 0.40 |
| 6.900 | 6164 | 0.10 | 63 | 0.01 |

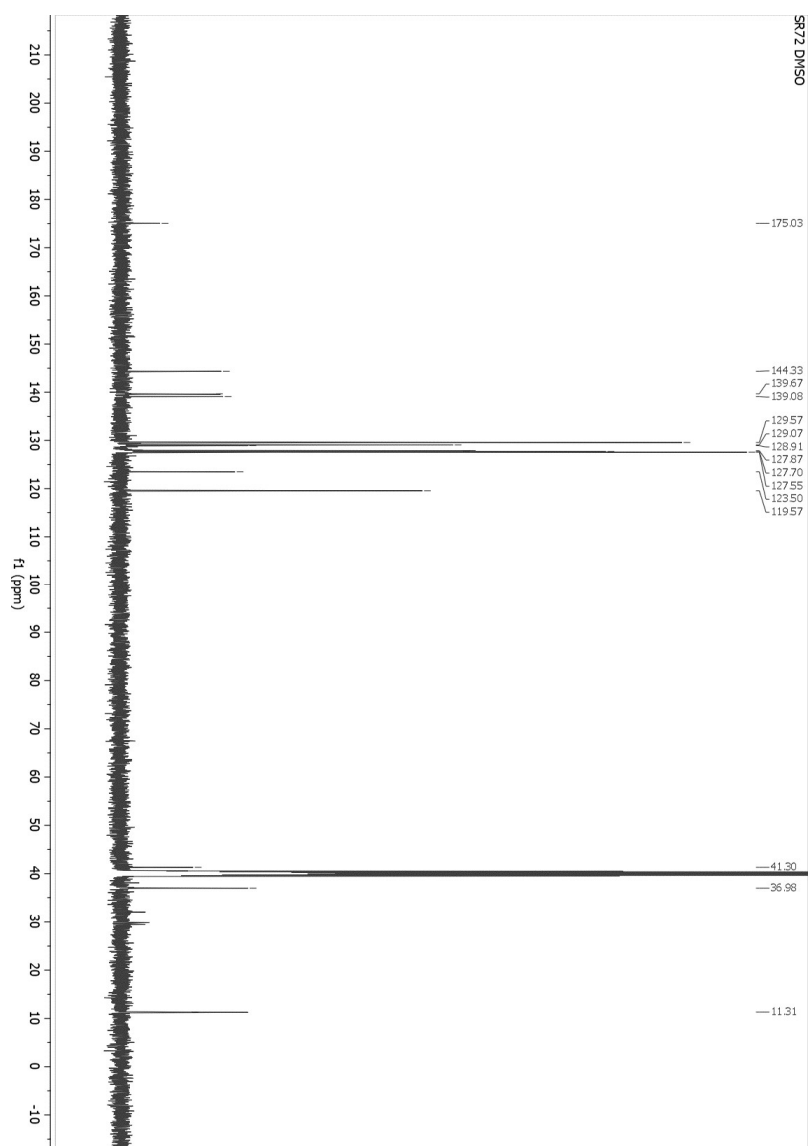
| | | | | |
|--------|---------|--------|---------|--------|
| Totals | 6148335 | 100.00 | 1232932 | 100.00 |
|--------|---------|--------|---------|--------|

215.0 nm Results

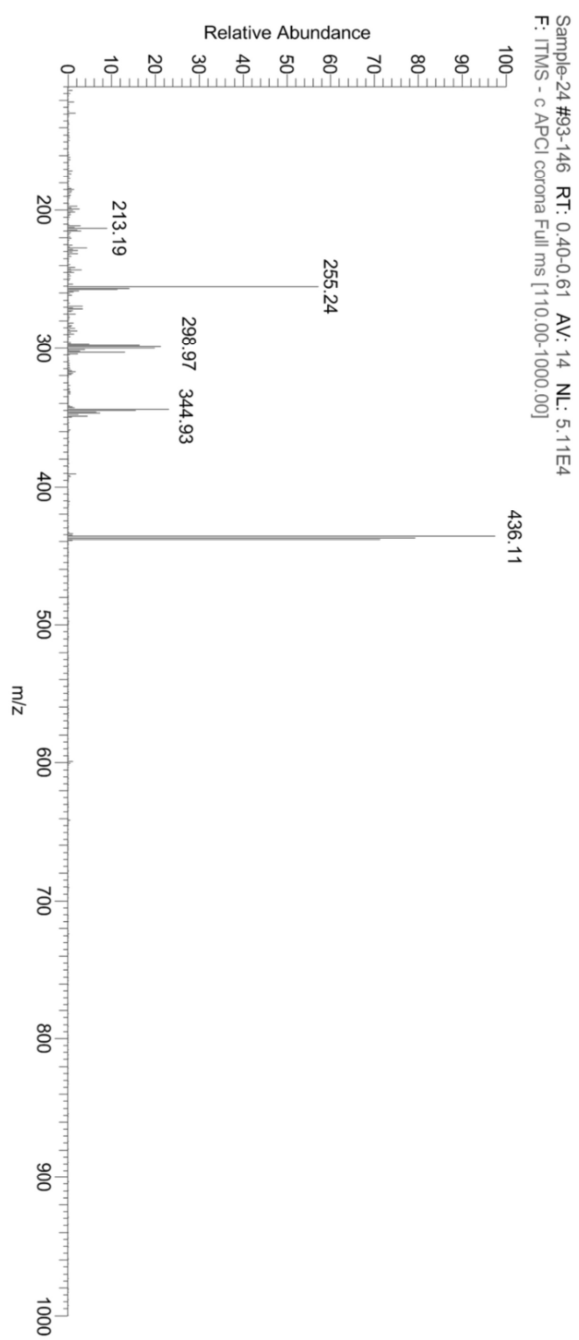
| Retention Time | Area | Area % | Height | Height % |
|----------------|------|--------|--------|----------|
|----------------|------|--------|--------|----------|

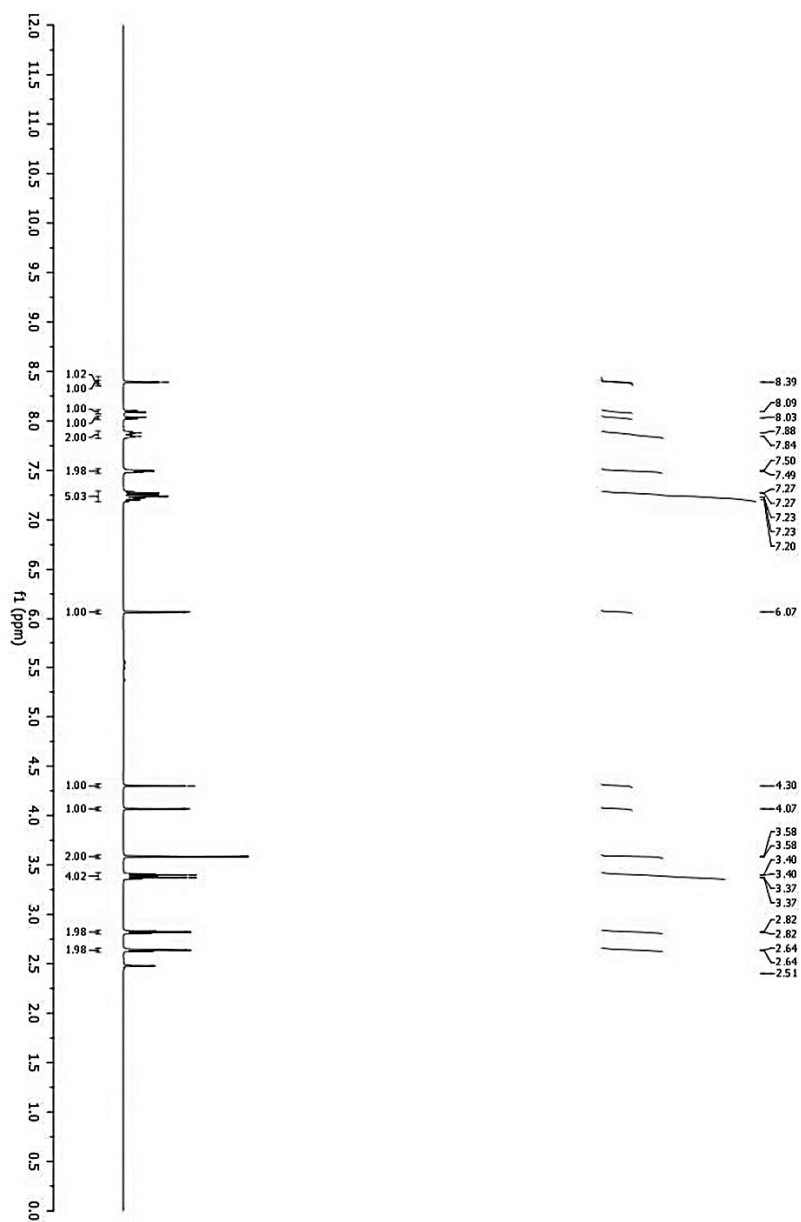
^1H NMR of Compound 42

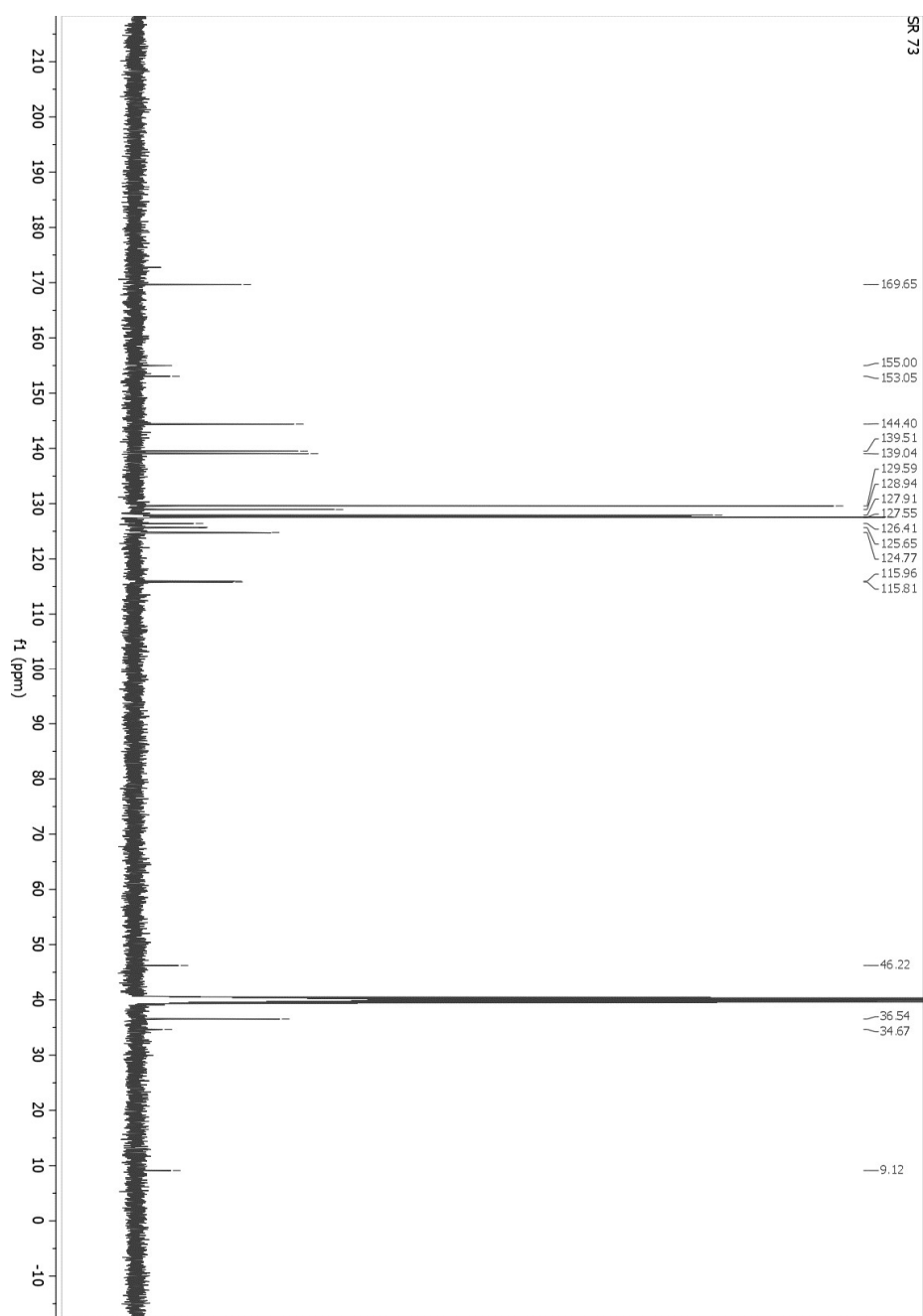
¹H NMR of Compound 72

^{13}C NMR of Compound 72

LCMS of Compound 72



¹H NMR of Compound 73

^{13}C NMR of Compound 73

List of Publications

1. Rayala Swetha, Anjali Sharma, Ravi Singh, Ankit Ganeshpurkar, Devendra Kumar, Ashok Kumar, and Sushil K. Singh. "Combined ligand-based and structure-based design of PDE 9A inhibitors against Alzheimer's disease." *Molecular Diversity* 26, no. 5 (2022): 2877-2892.
2. Rayala Swetha, Chandrim Gayen, Devendra Kumar, Tryambak Deo Singh, Gyan Modi, and Sushil Kumar Singh. "Biomolecular basis of matrix metallo proteinase-9 activity." *Future medicinal chemistry* 10, no. 9 (2018): 1093-1112.
3. Rayala Swetha, Devendra Kumar, Sukesh K. Gupta, Ankit Ganeshpurkar, Ravi Singh, Gopichand Gutti, Dileep Kumar, Srabanti Jana, Sairam Krishnamurthy, and Sushil K. Singh. "Multifunctional hybrid sulfonamides as novel therapeutic agents for Alzheimer's disease." *Future Medicinal Chemistry* 11, no. 24 (2019): 3161-3178.
4. Ghosh, Powsali, Ravi Singh, Ankit Ganeshpurkar, Rayala Swetha, Devendra Kumar, Sushil Kumar Singh, and Ashok Kumar. "Identification of potential death-associated protein kinase-1 (DAPK1) inhibitors by an integrated ligand-based and structure-based computational drug design approach." *Journal of Biomolecular Structure and Dynamics* (2022): 1-13.
5. Singh, Ravi, Ankit Vyankatrao Pokle, Powsali Ghosh, Ankit Ganeshpurkar, Rayala Swetha, Sushil Kumar Singh, and Ashok Kumar. "Pharmacophore-based virtual screening, molecular docking and molecular dynamics simulations study for the identification of LIM kinase-1 inhibitors." *Journal of Biomolecular Structure and Dynamics* (2022): 1-15.
6. Bajad, Nilesh Gajanan, Rayala Swetha, Ravi Singh, Ankit Ganeshpurkar, Gopichand Gutti, Ravi Bhushan Singh, Ashok Kumar, and Sushil Kumar Singh. "Combined

-
- structure and ligand-based design of dual BACE-1/GSK-3 β inhibitors for Alzheimer's disease." *Chemical Papers* (2022): 1-18.
7. Ganeshpurkar, Ankit, Rayala Swetha, Devendra Kumar, Gore P. Gangaram, Ravi Singh, Gopichand Gutti, Srabanti Jana, Dileep Kumar, Ashok Kumar, and Sushil K. Singh. "Protein-protein interactions and aggregation inhibitors in Alzheimer's disease." *Current topics in medicinal chemistry* 19, no. 7 (2019): 501-533.
 8. Sharma, Anjali, Rayala Swetha, Nilesh Gajanan Bajad, Ankit Ganeshpurkar, Ravi Singh, Ashok Kumar, and Sushil Kumar Singh. "Cathepsin B-A Neuronal Death Mediator in Alzheimer's Disease Leads to Neurodegeneration." *Mini Reviews in Medicinal Chemistry* (2022).
 9. Ganeshpurkar, Ankit, Ravi Singh, Devendra Kumar, Pravin Gore, Shalini Shivhare, Divya Sardana, Rayala Swetha, Ashok Kumar, and Sushil Kumar Singh. "Identification of sulfonamide based butyrylcholinesterase inhibitors through scaffold hopping approach." *International Journal of Biological Macromolecules* 203 (2022): 195-211.
 10. Bajad, Nilesh Gajanan, Rayala Swetha, Gopichand Gutti, Anjali Sharma, Meenakshi Singh, Ashok Kumar, and Sushil Kumar Singh. "Systematic review on role of structure based drug design (SBDD) in the identification of anti-viral leads against SARS-Cov-2." *Current Research in Pharmacology and Drug Discovery* 2 (2021): 100026.
 11. Bajad, Nilesh Gajanan, Rayala Swetha, Gopichand Gutti, Meenakshi Singh, Ashok Kumar, and Sushil Kumar Singh. "A systematic review of carbohydrate-based bioactive molecules for Alzheimer's disease." *Future Medicinal Chemistry* 13, no. 19 (2021): 1695-1711.
 12. Makar, Subhajit, Tanmay Saha, Rayala Swetha, Gopichand Gutti, Ashok Kumar, and Sushil K. Singh. "Rational approaches of drug design for the development of selective

estrogen receptor modulators (SERMs), implicated in breast cancer." *Bioorganic Chemistry* 94 (2020): 103380.

13. Saha, Tanmay, Subhajit Makar, Rayala Swetha, Gopichand Gutti, and Sushil K. Singh. "Estrogen signaling: An emanating therapeutic target for breast cancer treatment." *European Journal of Medicinal Chemistry* 177 (2019): 116-143.

More papers are under preparation and/or review.