

## Chapter 2

### Literature Review

*In this chapter, a number of recent studies have been critically analyzed to demonstrate that how the surface polarization or electric field stimulation together with functional properties of piezoelectric biomaterials can synergistically modulate cell functionality or tissue regeneration, in vitro / in vivo. The challenging problem of processing the compositionally tailored bioceramics is emphasized in particular reference to (Na, K)NbO<sub>3</sub>, an implantable biomaterial with the most attractive combination of piezoresponsive properties. In addition, an overview of bacterial infection in implants and various stimuli to inhibit bacterial infection such as, external electrical stimulation, magnetic field stimulation and surface charge potential has also been discussed.*

#### 2.1 Introduction

Bioelectrical cues in natural living bone, such as, piezoelectricity, pyroelectricity and ferroelectricity, etc. have been reported as some of the key factors in regulating metabolic activities like growth, structural remodeling as well as fracture healing [1, 2, 3, 4, 5]. The non-centrosymmetry of the collagen molecule has been suggested as the primary reason for the bioelectricity in living bone [1, 2, 5, 6, 7, 8]. The endogenous electric field of living bone also helps in governing the cell metabolism, such as, growth and proliferation, differentiation, motility, etc [9]. For example, human tibia generates a piezoelectric potential of about 300  $\mu\text{V}$  during walking [10]. The application of physiological compressive loads increases the negative charges due to piezoelectric potential in bone [6, 11]. These negative charges promote the functionality of osteoblast cells and consequently, enhance bone regeneration [6]. However, low mechanical pressure results in reduced neo-bone formation

as compared to higher pressure [12, 13]. The fractured region of living bone becomes more electronegative than the other regions [12, 14]. As mentioned above, the generation of these negative charges promotes the activity of osteoblast cells, which help in matrix mineralization at the fractured site [6, 11]. It can, therefore, be suggested that piezoelectric stimulation plays an important role in controlled bone regeneration. The piezoelectric collagen in bone also develops streaming potential under stress, which results in the reduction of hydraulic permeability as well as an increase in stiffness [15]. Apart from piezoelectricity, the biological response of hard tissues can also be improved by direct electrical stimulation [16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26].

The stress induced potential in piezo-bioceramics and piezo-biopolymers augments bone metabolism [27, 28, 29, 30, 31, 32]. Piezoelectric bioceramics have already been demonstrated as sources of *in vivo* energy for biosensors and pacemakers [33, 34]. Piezo-bioceramics/ biopolymers develop surface electrical charges under the application of external stress, similar to bone [Fig. 2.1 (A) and (B)] [35, 36, 37]. The polarized piezoelectric surfaces are reported to improve the osteogenic performance of natural bone [38, 39, 40, 41, 42]. Piezoelectric implants can potentially promote bone fracture repair [40, 42]. The implantation of piezoelectric scaffolds at the damaged site of the bone defect, develop stress generated electrical stimuli through physiological loads [Fig. 2.1 (C)] [43]. Such hyperpolarization activates voltage-gated calcium channels of the cell membrane [40]. The intracellular  $\text{Ca}^{2+}$  ions play an important role in cell proliferation [44]. It can, therefore, be inferred that piezoelectric biocompatible scaffolds result in an early repair of damaged tissue as compared to their non-piezoelectric counterparts [40]. For example, Liu et al. [45] compared the osseointegration of positively polarized  $\text{BiFeO}_3$  (+BFO) membrane – strontium

titanate (STO), negatively polarized BFO membrane – STO implant and non-polarized STO implant by the fixation in the rat femur. A built in or integral electric field [Fig. 2.1 (D)] develops at the interface of oppositely charged +BFO-STO (+75 mV) and electronegative bone defect surface (-52 to 87 mV) [45, 46], which consequently promote more osseointegration as compared to -BFO-STO and non-polarized STO surfaces [Fig. 2.1 (E)].

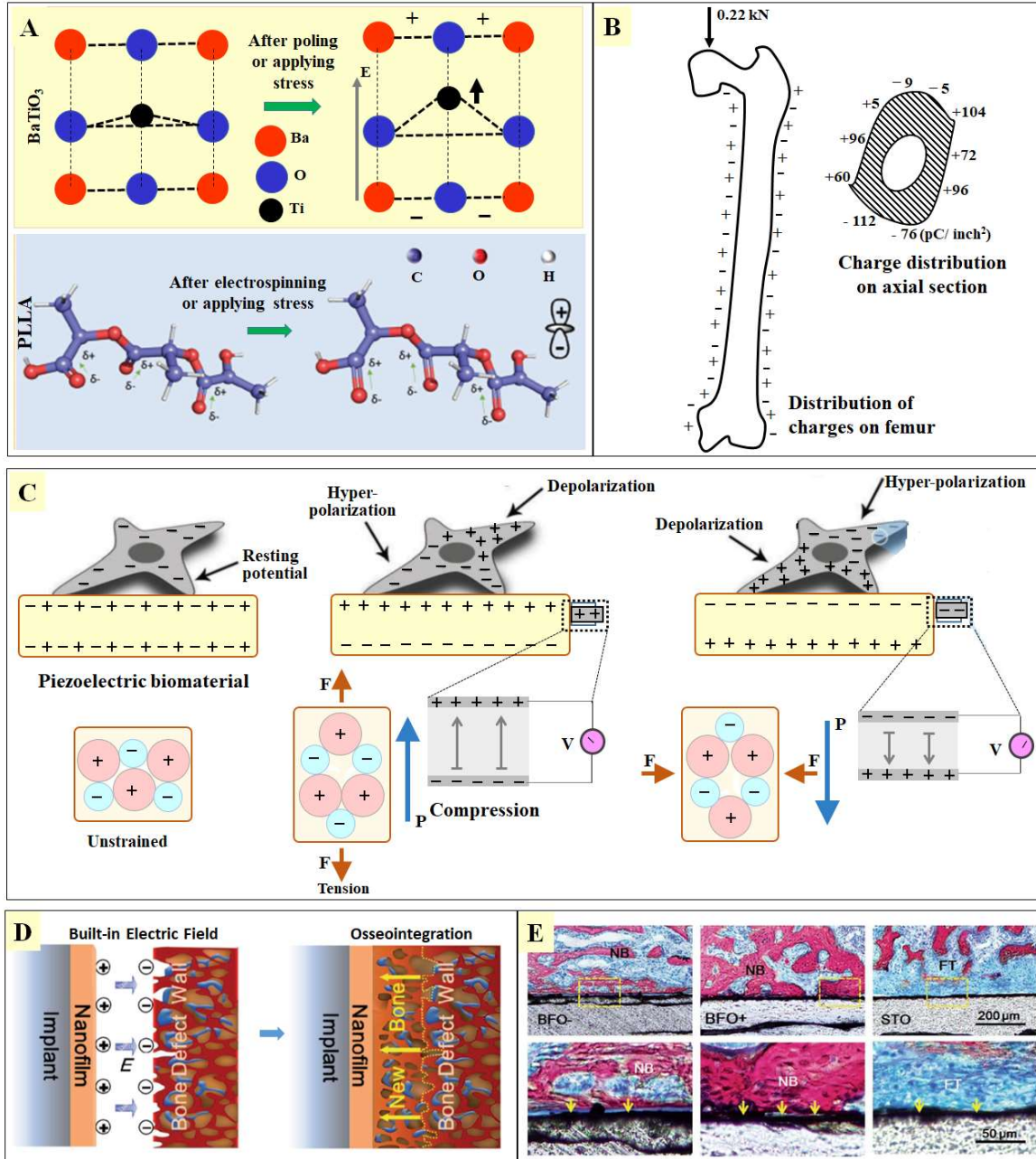


Fig. 2.1. Stress generated electrical charges in natural bone and piezoelectric biomaterials induced endogenous cellular potential and bone formation. (A) Schematic demonstration of the generation of aligned dipoles or surface charge on piezoelectric ceramic ( $BaTiO_3$ ) and polymer (PLLA) with the application of mechanical stress, similar to human bone (Reproduced with the permission from publisher, Ref. [35]). (B) Stress generated surface

*electrical charge on the circumference of human femur (Reproduced with the permission from publisher, Ref. [37]). (C) Schematic illustration of the generation of surface electrical charge on strained piezoelectric material and resulting hyper-polarization/ depolarization of cellular membrane (Reproduced with the permission from publisher, Ref. [43]). (D) Schematic representation of built-in electric field between positively charged nanocomposite and negatively charged bone wall, which enhances the osseointegration (Reproduced with the permission from publisher, Ref. [45]). (E) Histological evidences demonstrating significant neobone formation on the positively charged implant, when compared to negatively charged and uncharged implant (Yellow dotted block: implant-tissue interface, NB: nascent bone, FT: fibrous tissue, BFO: BiFeO<sub>3</sub>, STO: SrTiO<sub>3</sub>) (Reproduced with the permission from publisher, Ref. [45]).*

Figs. 2.2 (A), (B) and (C) illustrate the piezoelectric voltage generation from human bone during our routine activity [47, 48, 49]. For this purpose, piezoelectric sensors were embedded with skin, which was then subjected to stress during bend-release of fingers and elbow, during running etc. Positive and negative voltages were produced during bending and releasing of fingers, elbow and knee. Wang et al. [50] experimentally evaluated stress generated electrical potential from the implanted piezoelectric P(VDF-TrFE) scaffold during daily activity of Sprague Dawley rat [Fig. 2.2 (D)]. At the pulling condition of rat's leg, the peak output current and voltage from the strained implanted piezoelectric scaffold were recorded as 6 nA and 6 mV, respectively. Therefore, it can be suggested that the piezoelectric scaffold, implanted in the wounded site, generates electrical potential under physiological loads. This further stimulates osteoblastic activity and consequently, promotes bone regeneration in the wounded region.

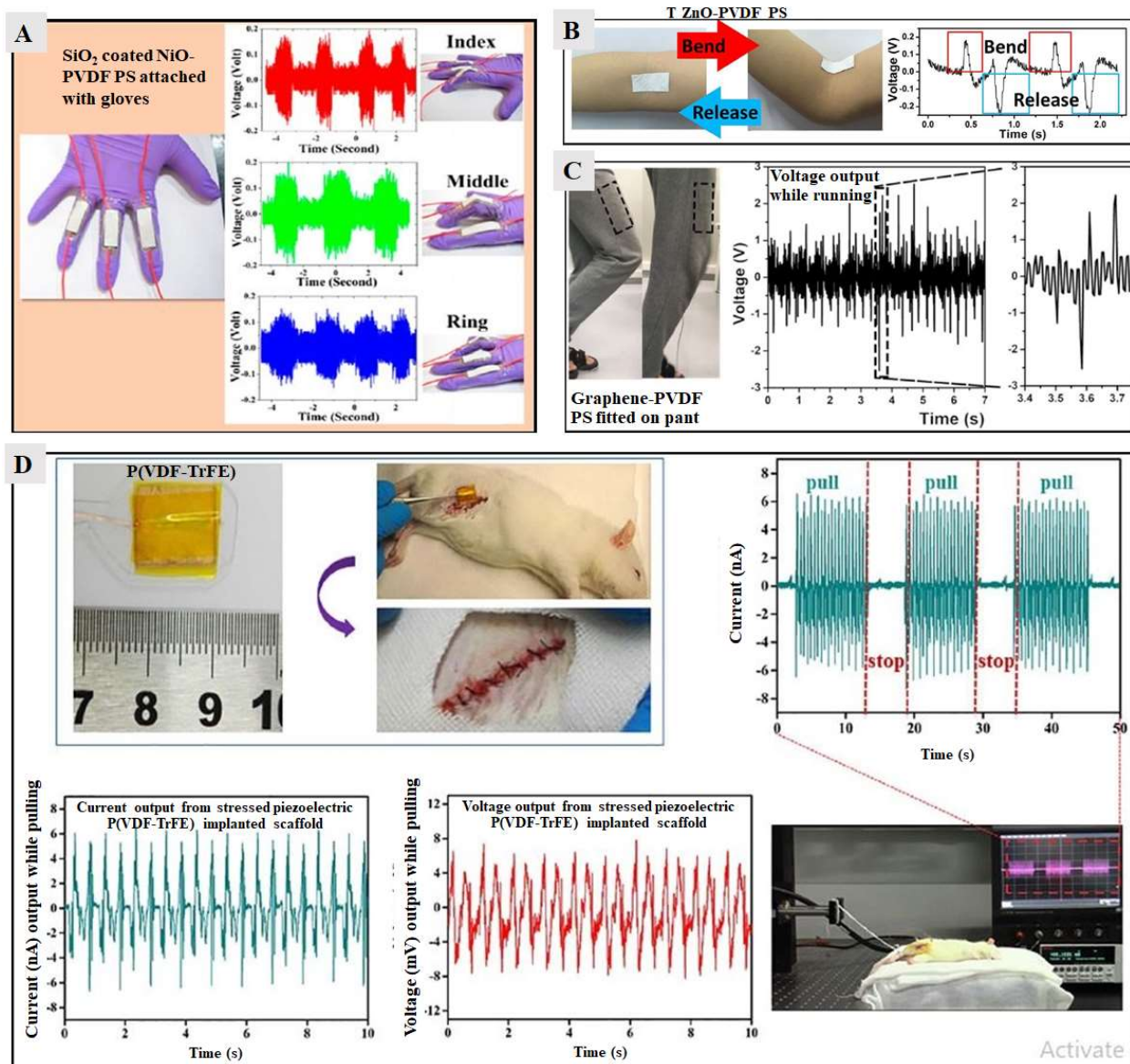


Fig. 2.2. Piezoelectric potential generated from natural bone during regular physiological activity. [(A), (B) and (C)] PVDF based piezoelectric sensors (PS) as detectors of human activity in terms of piezoelectricity induced voltage output. [(D)] P(VDF-TrFE) based piezopolymers (nanogenerator) implanted in the thigh of Sprague Dawley rat, mimicking piezoelectric potential generation during the regular activity. (A) PS nanocomposite, fabricated with PVDF/  $\text{SiO}_2$  coated NiO assembled with gloves to determine the piezoelectric voltage output variation during stretching and bending movement of index, middle and ring

*fingers (Reproduced with the permission from publisher, Ref. [47]). (B) A PS nanocomposite with tetrapod ZnO/ PVDF/ polyester fabric, attached with elbow, to evaluate the amount of voltage output during bending and release (Reproduced with the permission from publisher, Ref. [48]). (C) A PS fabricated with PVDF/ graphene in polyester fabric attached with marked region to determine the voltage output variation during running (Reproduced with the permission from publisher, Ref. [49]). (D) Electrospun P(VDF-TrFE) nanoscaffold implanted in the thigh of Sprague Dawley rat. The leg of rat was pulled up and released through a motor (bottom-right) and resulting variation in current output are shown (top-right) during pulling and releasing. The current and voltage output are also shown in bottom-left and bottom-middle graph, respectively in the pulling condition of rat's leg (Reproduced with the permission from publisher, Ref. [50]).*

Against the above backdrop, the potentiality of various piezoelectric bioceramics and biopolymers in mimicking electro-mechanical response of living bone has been revealed. Towards this end, the processing-related challenges to meet the desired piezoresponsive property combination have been discussed along with the possible combinations of remedies. In addition, the advantageous effects of polarized piezoelectric substrates in augmenting the biocompatibility have been discussed objectively.

NKN and BaTiO<sub>3</sub> possess ferroelectricity at a temperature lower than the Curie temperature, as represented in Figs. 2.3 (a) and (b), respectively. In Na<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub> ceramics, temperature dependent ferroelectric phase transformations occur from cubic to tetragonal (at ~ 420 °C) to monoclinic / orthorhombic (at ~ 210 °C) to rhombohedral ( $\leq - 160$  °C) phases [51, 52, 53]. However, in BaTiO<sub>3</sub> ceramics, ferroelectric phase transformations take place from tetragonal

(130 to 5 °C) to orthorhombic (5 to - 90 °C) to rhombohedral ( $\leq$  - 90 °C) phases. Overall, NKN and BaTiO<sub>3</sub> exhibit ferroelectric phase at room temperature [54].

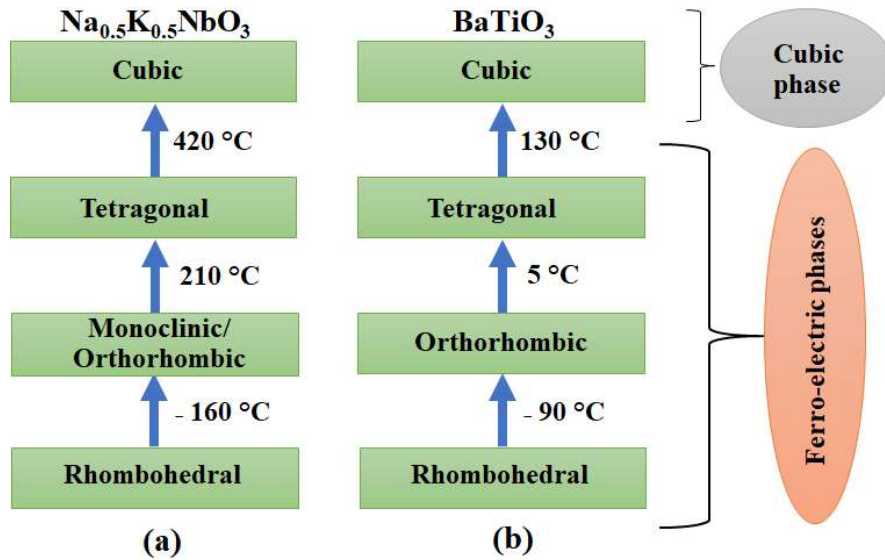


Fig. 2.3. Schematic illustrating the temperature dependent phase transformations (a) In Na<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub> [51, 52, 53] and (b) In BaTiO<sub>3</sub> ceramics [54].

## 2.2. Piezoelectric biomaterials

Among various biomaterials, piezoelectric bioceramics, such as, sodium potassium niobate (Na<sub>x</sub>K<sub>y</sub>NbO<sub>3</sub>, where, 0 ≤ x ≤ 0.8; 0.2 ≤ y ≤ 1; NKN) [55] or NKN-based ceramics [56, 57], BaTiO<sub>3</sub> [5, 38, 58, 59, 60], lithium sodium potassium niobate [61, 62, 63], LiNbO<sub>3</sub> [64, 65], MgSiO<sub>3</sub> [66, 67, 68], KNbO<sub>3</sub> [69], ZnO [44], boron nitride [70], bismuth ferrite (BiFeO<sub>3</sub>) [71] are reported to demonstrate reasonable potentiality to be prospective electroactive prosthetic implants. Similar examples from the class of piezoelectric biopolymers include polyvinylidene fluoride (PVDF) [42], poly-L-lactic acid (PLLA) [72], polyhydroxybutyrate (PHB) [73], polyamides (Nylons, peptides, etc.) [74], polysaccharides (cellulose, chitosan, etc.) [75, 76, 77], collagen [78], etc. The dielectric, piezoelectric, ferroelectric, pyroelectric and electromechanical properties of the above mentioned piezoelectric bioceramics/

biopolymers are summarized in Table 2.1 [7, 8, 29, 63, 65, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 102, 103, 104, 105, 106, 107, 108, 109, 110, 111, 112, 113, 114, 115, 116, 117, 118, 119, 120, 121, 122, 123, 124, 125, 126, 127, 128].

**Table 2.1. A comparison of dielectric constants ( $\epsilon_r$ ), piezoelectric strain coefficients ( $d_{33}$ ), mechanical quality factors ( $Q_m$ ), remnant polarization ( $P_r$ ), electromechanical coupling coefficients ( $k_p$ ) and pyroelectric coefficients ( $p$ ) of various piezoelectric biomaterials with those of the natural bone.**

<b>Piezoelectric Biomaterials</b>	<b><math>\epsilon_r</math> (at 1-100 kHz)</b>	<b><math>d_{33}</math> (pC/N)</b>	<b><math>Q_m</math></b>	<b><math>P_r</math> (<math>\mu\text{C}/\text{cm}^2</math>)</b>	<b><math>k_p</math> (%)</b>	<b>(<math>p</math>) (<math>\mu\text{C}/\text{m}^2\text{K}</math>)</b>	<b>Ref.</b>
$\text{Na}_{0.5}\text{K}_{0.5}\text{NbO}_3$	657	160	240	31.4	44	-	[83, 84]
Barium titanate ( $\text{BaTiO}_3$ )	1135	191	32	12.6	35	200	[85, 86, 87, 88, 89]
$\text{Li}_{0.06}(\text{Na}_{0.5}\text{K}_{0.5})_{0.94}\text{NbO}_3$	722.4	124	-	16.8	30.63	-	[63, 90]
Lithium niobate ( $\text{LiNbO}_3$ )	62	23	-	-	-	103.9	[65, 91, 92]
Magnesium silicate ( $\text{MgSiO}_3$ )	-	-1.74 ( $d_{31}$ )	-	-	-	-	[93]
Potassium niobate ( $\text{KNbO}_3$ )	394	91.7	325	-	28	93	[94, 95]
Zinc oxide ( $\text{ZnO}$ )	12.27	12.4	-	16.31	48 ( $k_{33}$ )	9.4	[89, 97, 98, 99]
Boron nitride (BN)	2 - 4	31.2 ( $d_{31}$ )	-	-	-	-	[100, 101]
Polyvinylidene fluoride (PVDF)	6 - 12	34	17.2	13	20 ( $k_{33}$ )	41	[102, 103, 104, 105, 106, 107, 108]
P(VDF-TrFE)	18	38	-	9.9	29 ( $k_{33}$ )	50	[103, 109, 110, 111, 112, 113]

Poly-L-lactic acid (PLLA)	3 - 4	9.82 (d <sub>14</sub> )	30.3	-	-	-	[29, 114, 115]
Polyhydroxybutyrate (PHB)	2 - 3.5	1.6-2 (d <sub>14</sub> )	-	-	-	-	[29, 116, 117]
Polyamide	4 - 5	4 (d <sub>31</sub> )	8.5	5.6 - 8.6	11	5	[29, 113, 118, 119, 120, 121]
Collagen	2.6	0.2 - 2 (d <sub>14</sub> )	-	0.29	-	0.12 - 0.37 @ 40 °C	[122, 123, 124, 125]
Chitosan	3.94	0.2 - 1.5 (d <sub>14</sub> )	-	0.178	-	7	[122, 123, 126, 127, 128]
Human bone	9.2	7.50 - 9	-	0.00068	-	0.0036 ± 0.0021	[7, 8, 79, 80, 81, 82, 96]

Among various piezoelectric ceramics, Na<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub> and BaTiO<sub>3</sub> exhibit comparatively higher piezoelectric and electrical properties. PVDF shows the best piezoelectric and electrical properties among mentioned piezoelectric polymers. Therefore, such piezoelectric ceramic and polymer can be suggested as potential candidate for orthopedic applications.

The piezoelectricity of the above materials [Table 2.1] is associated with their non-centrosymmetric structures. On the basis of the degree of symmetry or orientation, crystals can be classified into 32 crystal classes [Fig. 2.4] [129, 130]. Among 32 crystal classes, 21 classes possess non-centrosymmetric structures. Of these, one belongs to combined cubic symmetry and does not possess piezoelectricity, whereas, remaining 20 crystal classes exhibit piezoelectric phenomena. 10 crystal classes, among these 20 non-centrosymmetric classes, develop polarization only by the application of mechanical stress and the remaining 10 crystal classes possess spontaneous polarization i.e., they possess piezoelectricity as well

as pyroelectricity. A subgroup of these crystal classes demonstrates reversible spontaneous polarization which defines the ferroelectric behavior [129, 130].

As far as piezoelectric properties are concerned, NKN-based ceramics have been demonstrated among the best candidates, because of their reasonable piezoelectric strain coefficient ( $d_{33} = 260 \text{ pC/N}$ ), mechanical quality factor (240), high electromechanical coupling coefficient ( $k_{33} = 53 \%$ ,  $k_p = 48 \%$ ), and high Curie temperature ( $420^\circ\text{C}$ ) as well as relatively lower density ( $4.51 \text{ g/cm}^3$ ) as compared to other piezoelectric biocompatible materials [83, 131, 132, 133]. NKN has been patented as a potential biocompatible implant material, as it supports the growth of human monocytes [55].  $\text{BaTiO}_3$  is another piezoelectric candidate with proven biocompatibility, *in vivo*. It has been demonstrated that  $\text{BaTiO}_3$  promotes osteogenesis in dog femur bone [38, 58]. Below its Curie temperature ( $T_c = 130^\circ\text{C}$ ),  $\text{BaTiO}_3$  possesses a polar asymmetric structure that imparts piezoelectricity [39, 134].  $\text{MgSiO}_3$ , with an asymmetric tetragonal structure, has been reported to be a good biocompatible piezoelectric substitute for bone, as testified by various *in vitro* and *in vivo* studies [66, 68, 135, 136].

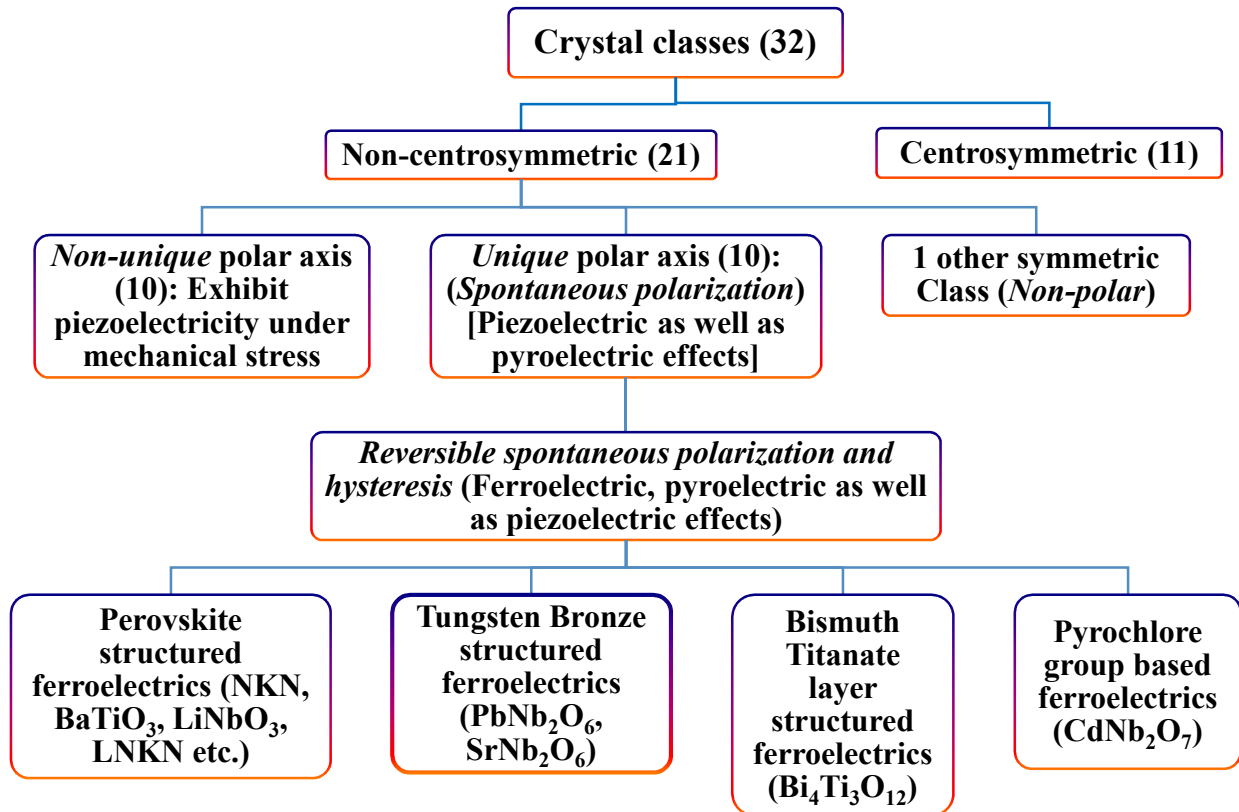


Fig. 2.4. Classification of various crystal classes on the basis of the center of symmetry with reference to piezoelectricity, pyroelectricity and ferroelectricity (adapted from refs. [129] and [130]).

LiNbO<sub>3</sub> is another piezoelectric material, which has been suggested as a potential alternative for bone prosthesis [64, 65, 137]. LiNbO<sub>3</sub> possesses admirable ferroelectric (spontaneous polarization;  $p_s = 78 \mu\text{C}/\text{cm}^2$ ), pyroelectric ( $P = 103.9 \mu\text{C}/\text{m}^2\text{K}$ ) and piezoelectric ( $d_{33} = 23 \text{ pC}/\text{N}$ ) properties [65, 91, 137]. Piezoelectric KNbO<sub>3</sub> is currently being used as a bio-probe for disease diagnosis and could be another alternative as an electroactive bone substitute [138, 139]. ZnO is another ceramic with reasonable piezoelectric ( $d_{33} = 12.4 \text{ pC}/\text{N}$ ) [98], pyroelectric ( $9.4 \mu\text{C}/\text{m}^2\text{K}$ ) [89] and dielectric ( $\epsilon = 12.27$ ) [98] potentials. It also has the ability to generate piezoelectric potential, which results in enhanced cellular functionality and

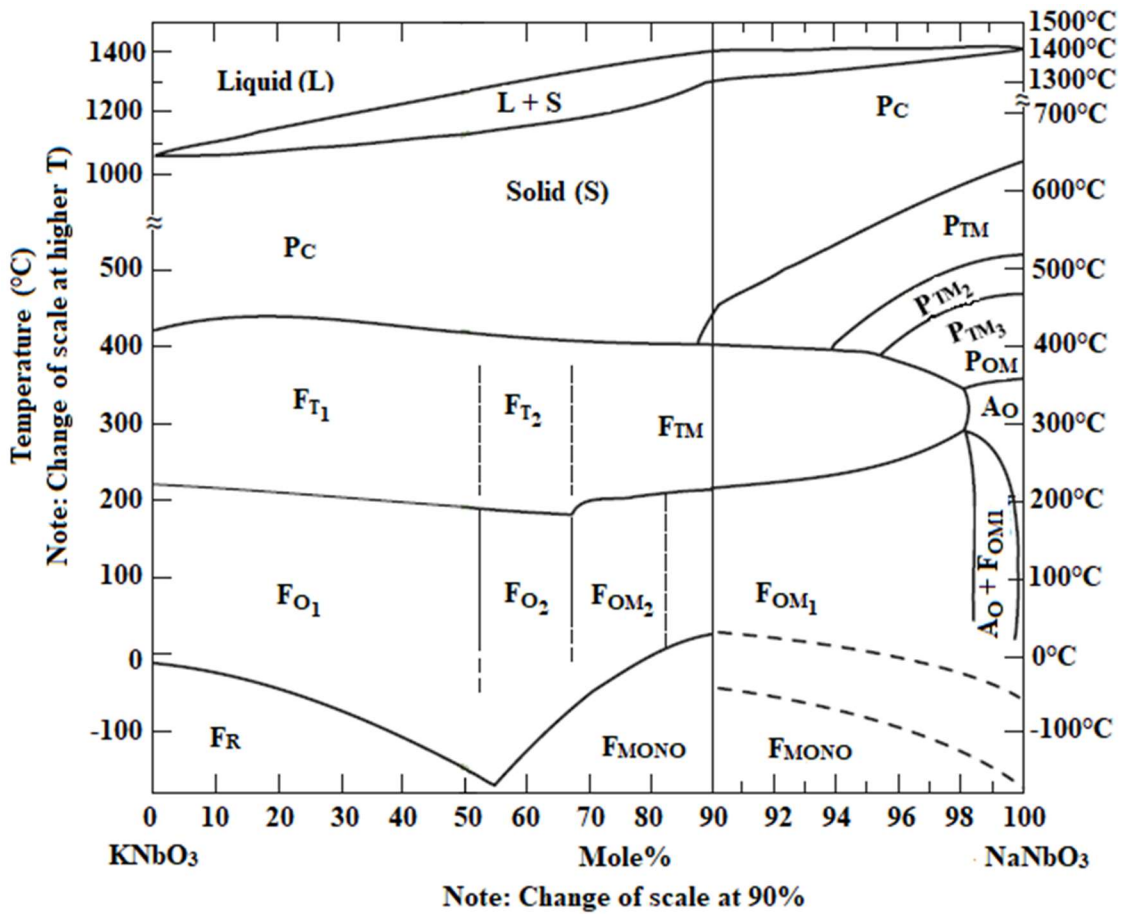
subsequently, promotes osteogenesis [44]. The piezoelectric boron nitride (BN) nanotubes, in its pristine as well as composite forms, have been reported to promote the proliferation and differentiation of mesenchymal stem cells [70]. Piezoelectric natural (proteins, polysaccharides, etc.) / synthetic (PVDF, PLLA, PHB, etc.) biopolymers are another class of emerging materials, that can be utilized for the regeneration of soft as well as hard tissues [27, 29, 75, 78].

Piezo-bioceramics and piezo-biopolymers are emerging biomaterials as next generation orthopedic implants. However, a number of processing concerns is to be addressed in synthesizing the consolidated stoichiometric composition of the above mentioned piezoelectric ceramics. For example, the volatilization of alkali elements (Na, K) restricts the high temperature processing and consequently, the densification of NKN-based ceramics by conventional processing techniques [131, 140, 141, 142, 143]. To address such processing concerns, a number of approaches, such as use of sintering aids (to reduce the sintering temperature) [144, 145, 146, 147], reduced atmosphere sintering [148, 149], advanced sintering routes, viz. cold isostatic pressing [62], hot pressing [150], hot isostatic pressing [151], spark plasma sintering [152], additive engineering [153, 154, 155, 156] are being investigated.

### **2.3. Processing-related challenges for piezoelectric NKN-based ceramics system**

$\text{Na}_x\text{K}_{(1-x)}\text{NbO}_3$  is a solid solution of ferroelectric  $\text{KNbO}_3$  and anti-ferroelectric  $\text{NaNbO}_3$  [Fig. 2.5 (a)] [51, 52]. As illustrated in Fig. 2.5 (a), three morphotropic phase boundaries (MPB) occur at  $\text{NaNbO}_3$  proportions of 52.5, 67.5 and 82.5 mol. %, respectively [157]. The highest value of the piezoelectric strain coefficient obtained at MPB, which corresponds to the composition,  $x = 0.5$  [52, 157, 158, 159]. Below its Curie temperature (420 °C), three

ferroelectric polymorphs, namely tetragonal, monoclinic / orthorhombic and rhombohedral exist. The polymorphic phase transitions (PPT) from rhombohedral to monoclinic / orthorhombic to tetragonal occur at temperatures of - 160 °C and 200 °C, respectively [Fig. 2.5 (a)] [51, 52, 53]. The PPT in NKN depends on compositions of the precursors as well as temperature [Figs. 2.5 (a) and (b)] [160, 161, 162, 163]. The piezoelectric properties of NKN can be improved by stabilizing the high temperature (200 °C) tetragonal phase at room temperature. The room temperature tetragonal phase in NKN can be established with the doping of Li (7 mol. %) [164], LiTaO<sub>3</sub> (5 - 6 mol. %) [153, 160], LiNbO<sub>3</sub> (5 - 7 mol. %) [154], LiSbO<sub>3</sub> (5.2 mol. %) [161] etc. in NKN matrix. Figs. 2.5 (b) and (c) distinguish the physical significance of PPT and MPB in NKN. As the tetragonal to monoclinic / orthorhombic phase transition temperature can be shifted to room temperature by compositional modifications in NKN, PPT can be suggested to be dependent on composition, as well. However, MPB is dependent on the composition only [165]. PPT is less preferable as compared to MPB due to temperature-dependent fluctuations in dielectric and piezoelectric properties [166, 167]. At room temperature, NKN unit cell is described as the orthorhombic phase, while its primary perovskite cell (ABO<sub>3</sub>) shows monoclinic symmetry [157].



(a)

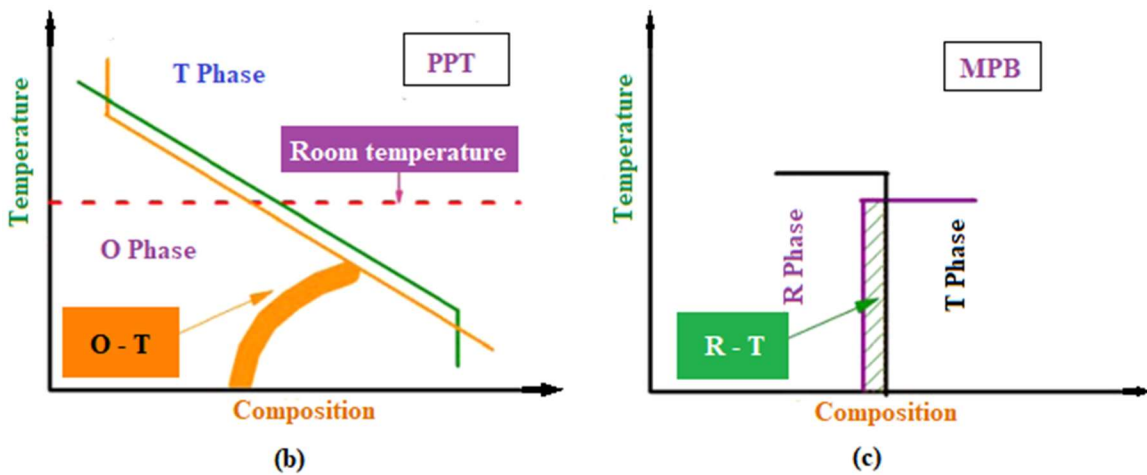
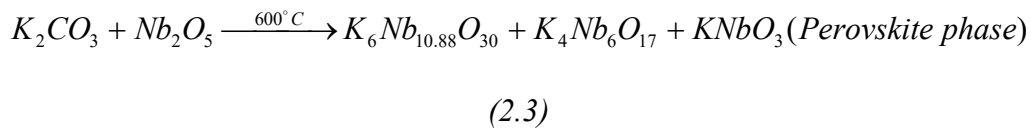
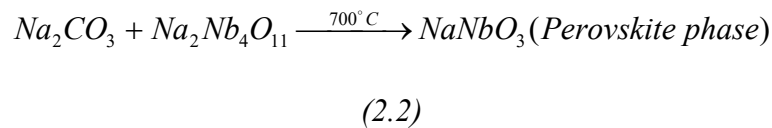
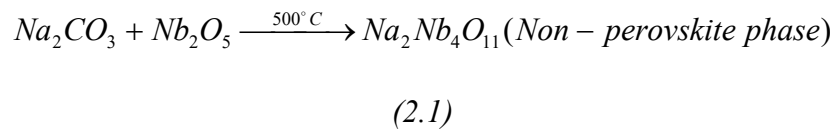
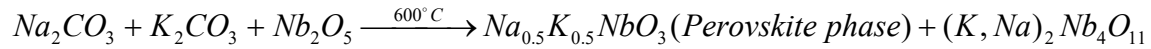


Fig. 2.5 (a). Phase diagram of  $\text{KNbO}_3\text{-NaNbO}_3$  solid solution, where P and F denote the paraelectric and ferroelectric phases, respectively. R, O, T and C indicate rhombohedral, monoclinic / orthorhombic, tetragonal and cubic phases, respectively. In  $\text{Na}_x\text{K}_{1-x}\text{NbO}_3$ , the

*morphotropic phase boundary (MPB) at  $x = 0.5$  distinguishes two orthorhombic and two tetragonal phases. Below Curie temperature, polymorphs like tetragonal - monoclinic / orthorhombic and monoclinic / orthorhombic - rhombohedral coexist at about 200 °C and -160 °C, respectively, (Reproduced with the permission from publisher, Ref. [52]). (b) Polymorphic phase transformation (PPT): tetragonal to monoclinic / orthorhombic (polymorphs) phase transformation temperature of NKN can be shifted to room temperature by chemical modification, indicating the dependency of PPT on composition as well as temperature, (c) Morphotropic phase boundary (MPB): Coexistence of rhombohedral and tetragonal phases at a certain composition (Reproduced with the permission from the publisher, Ref. [165]).*

Malice et al. [168] suggested that the number of intermediate reactions takes place between the constituent phases ( $\text{Na}_2\text{CO}_3$ ,  $\text{Nb}_2\text{O}_5$ , and  $\text{K}_2\text{CO}_3$ ) during the formation of  $\text{Na}_{0.5}\text{K}_{0.5}\text{NbO}_3$  ceramics via solid-state synthesis route. During calcination, two raw binary ( $\text{Na}_2\text{CO}_3/\text{Nb}_2\text{O}_5$ ,  $\text{K}_2\text{CO}_3/\text{Nb}_2\text{O}_5$ ) phases and a tertiary raw ( $\text{Na}_2\text{CO}_3/\text{K}_2\text{CO}_3/\text{Nb}_2\text{O}_5$ ) phase reacts, as follows [168],

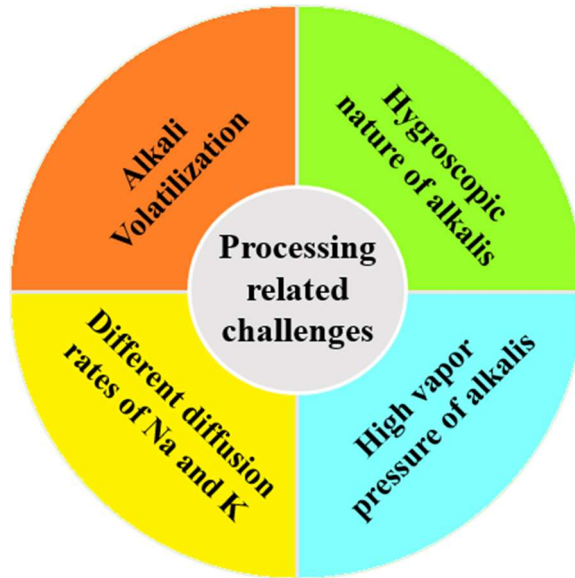




(2.4)

The calcination temperature for NKN has been suggested to be in the range of 750 °C to 950 °C [83, 131, 132, 168, 169, 170, 171, 172, 173].

The alkali volatilization at the higher temperatures (> 1000 °C) and hygroscopic nature of alkaline carbonate precursors, especially K<sub>2</sub>CO<sub>3</sub> are the primary concerns associated with the processing of stoichiometric NKN-based ceramics [140, 174]. The melting temperature of NaNbO<sub>3</sub> and KNbO<sub>3</sub> are about 1420 °C and 1040 °C, respectively [142, 175]. The consolidation, as well as microstructural control during high temperature sintering of NKN-based ceramics is difficult due to their close sintering and solidus temperatures [131, 141, 142, 176]. The solidus and liquidus temperatures for Na<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub> are about 1140 °C and 1280 °C, respectively [142]. In addition, a difference in diffusion rates of Na and K ions, with K<sup>+</sup> being the slowest diffusing species, retards the formation of stoichiometric NKN [143]. High vapor pressure of alkalis at higher temperatures (~ 990 °C) results in alkali phase reduction in NKN. The difference in equilibrium vapor pressures of K (P<sub>K</sub> = 8×10<sup>-3</sup> Pa) and Na (P<sub>Na</sub> = 3×10<sup>-3</sup> Pa) has also been suggested as one of the issues in the processing of stoichiometric NKN-based ceramics [177]. Fig. 2.6 represents the key processing concerns in achieving phase pure and dense NKN.



*Fig. 2.6. Schematic representing key issues, associated with the processing of stoichiometric NKN [131, 141, 142, 143, 176, 177].*

#### **2.4. Addressing the processing concerns for piezoelectric NKN-based ceramics**

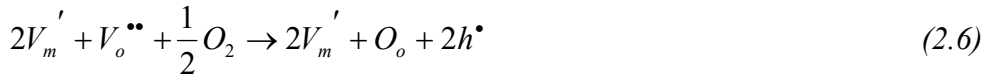
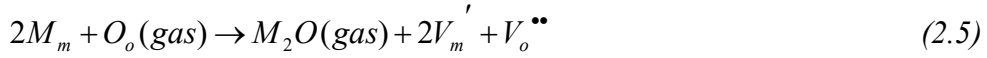
The alkali volatilization at higher temperatures results in the formation of secondary phases, such as,  $K_6Nb_{10.88}O_{30}$ ,  $K_2Nb_4O_{11}$ , etc. [178]. The sintering of NKN, above 1000 °C, causes the formation of liquid phase due to the evaporation of  $Na_2O$ , which results in higher densification [141]. However,  $Na_2O$  evaporation results in the reduction of piezoelectric properties of NKN ceramics [141]. In order to control the evaporation of  $Na_2O$ , various sintering aids (low melting point oxides) such as, ZnO doped - CuO (sintering temperature,  $T_s = 900$  °C) [144],  $V_2O_5$  ( $T_s = 920$  °C) [145], (K, Na) - Germanate ( $T_s = 1000$  °C) [146],  $Li_2O$  ( $T_s = 1000$  °C) [147] etc. have been incorporated to reduce the sintering temperature of NKN-based ceramics. The addition of 5 mol. %  $BaTiO_3$  to the NKN specimen ( $0.95 Na_{0.5}K_{0.5}NbO_3 - 0.05 BaTiO_3$ ) has also been suggested as one of the ways to control  $Na_2O$  evaporation [141]. Such modification results in excellent piezoelectric properties ( $k_p = 36$  %,  $d_{33} = 225$  pC/N) at relatively lower sintering temperature of 1060 °C (2 h) [141].

The consolidation of NKN can also be improved by liquid phase sintering (LPS) [179], where sintering aids with the lower melting point are incorporated in NKN matrix phase to reduce the sintering temperature. It prevents alkali phase reduction, which consequently, enhances the densification of NKN-based ceramics. Therefore, the piezoelectric as well as electrical properties of NKN can be improved either by reducing the sintering temperature below 1000 °C or by controlling the evaporation of Na<sub>2</sub>O at higher temperatures (> 1000 °C). The important LPS aids for NKN includes, Cu, Zn, borate and germinate-based compounds [146, 180, 181, 182, 183]. Cu-based sintering aids are K<sub>4</sub>CuNb<sub>8</sub>O<sub>23</sub> (0.5 mol. %), K<sub>5.4</sub>Cu<sub>1.3</sub>Ta<sub>10</sub>O<sub>29</sub> (0.38 mol. %), etc., which enhance the densification (> 97.6 % ρ<sub>th</sub>) and piezoelectric properties (d<sub>33</sub> = 180 - 190 pC/N, k<sub>p</sub> = 39 - 42 %, Q<sub>m</sub> = 1200 - 1400) of NKN-based ceramics, at relatively lower sintering temperatures (1095 - 1120 °C) [180]. The addition of ZnO (1 mol. %) and K<sub>1.94</sub>Zn<sub>1.06</sub>Ta<sub>5.19</sub>O<sub>15</sub> (0.5 mol. %) has also been reported to provide improved densification (upto 96.3 % ρ<sub>th</sub>) and piezoelectric properties (d<sub>33</sub> = 123 - 126 pC/N, k<sub>p</sub> = 40 - 42 %, Q<sub>m</sub> = 60 - 140) of NKN-based ceramics [181, 182]. Similarly, borate-based sintering aids such as, 0.45 wt. % of borax or Na<sub>2</sub>B<sub>4</sub>O<sub>7</sub> · 10 H<sub>2</sub>O (ρ<sub>r</sub> = 95.5 % ρ<sub>th</sub>, d<sub>33</sub> = 131.6 pC/N, k<sub>p</sub> = 34.8 %, Q<sub>m</sub> = 109) [183] and 1 wt. % of germinate-based K, Na-germinate sintering aids (ρ<sub>r</sub> = 95.6 % ρ<sub>th</sub>, d<sub>33</sub> = 120 pC/N, k<sub>p</sub> = 40 %, Q<sub>m</sub> = 77) also contribute to the densification and piezoelectric properties of NKN-based ceramics [146].

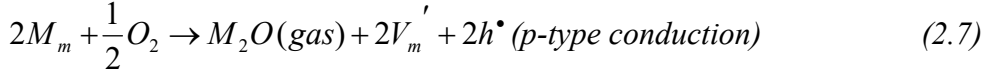
Kobayashi et al. [148, 149] suggested that the sintering in reduced oxygen can effectively diminish the possibility of the volatilization of alkalis during the processing of NKN ceramics. Such processing results in relatively high dielectric constant (415.6) and resistivity (2.70 × 10<sup>9</sup> Ω·m) in NKN. In reduced atmosphere sintering, the energy for the formation of oxygen vacancies decreases and that of alkali vacancies increases. Consequently, oxygen

vacancy increases and alkali (Na, K) vacancy decreases, which indicates a decrease in alkali volatilization. Alkali volatilization during sintering in air as well as reduced oxygen atmosphere can be represented as [148, 149],

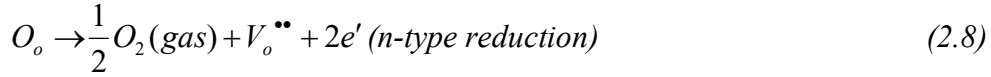
(a) In air atmosphere (p-type conduction),



The combination of Eqns. (2.5) and (2.6) yields,



(b) In reduced oxygen atmosphere (n-type conduction),



where, M,  $V_m$ , and  $V_o$  indicate alkali metal, vacancies of alkali metal and oxygen, respectively.

#### 2.4.1. Conventional processing routes

A number of efforts have been made to obtain reasonable densification in NKN-based ceramics through an inexpensive conventional sintering route. Egerton et al. [169] fabricated NKN ceramics via solid-state route at various sintering temperatures, ranging from 1050 - 1325 °C, which led to the densification of 94 - 97 %  $\rho_{th}$ . Similarly, a relative density of 93.4 - 95.3 %  $\rho_{th}$  has been reported for the sintering temperatures, ranging from 1100 - 1115 °C (2 h) [172, 184, 185]. However, densification of 97 - 97.6 %  $\rho_{th}$  has been demonstrated for the sintering temperature of 1120 - 1125 °C of NKN, which subsequently, exhibits good piezoelectric response ( $d_{33} = 120$  pC/N) [131, 173]. Zuo et al. [132] reported maximum

densification of 98.4 %  $\rho_{th}$  in NKN on reducing the particle size in the nano range (70 nm), prior to conventional sintering (1100 °C, 4h). Similarly, Du et al. [173] suggested that apart from sintering temperature (1120 °C, 2h), heating rate (optimized at 5 °C/ min) is one of the important parameters to provide reasonable densification (97.56 %  $\rho_{th}$ ) and piezoelectric properties ( $d_{33} = 120$  pC/N and  $k_p = 40$  %). LiNbO<sub>3</sub> (7 mol. %) modified NKN ceramic i.e., 0.93 (Na<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub>) - 0.07 (LiNbO<sub>3</sub>), possesses excellent piezoelectric properties ( $d_{33} = 240$  pC/N and  $k_p = 35$  %), which was consolidated at 1030 °C, followed by annealing at 1050 °C (i.e., two-step sintering) [186]. However, it has been reported that modification with LiNbO<sub>3</sub> as 0.942 (Na<sub>0.535</sub>K<sub>0.480</sub>) NbO<sub>3</sub> - 0.058 LiNbO<sub>3</sub> results in further improvement in piezoelectric properties ( $d_{33} = 314$  pC/N,  $k_p = 41.2$  %), when sintered at approximately similar temperature (1060 °C) [187]. Doping of Li and Ta at alkali and niobium side in NKN i.e., (Li<sub>0.04</sub>K<sub>0.44</sub> Na<sub>0.52</sub>) (Nb<sub>0.85</sub>Ta<sub>0.15</sub>) O<sub>3</sub> and subsequent sintering at 1090 °C, reveals increased densification up to 98 % with good piezoelectric property ( $d_{33} \sim 206$  pC/N) [188]. NKN doped with Ta, Sb, and Li i.e., (Na<sub>0.52</sub> K<sub>0.48-x</sub>) (Nb<sub>1-x-y</sub>Sb<sub>y</sub>) O<sub>3</sub> - LiTaO<sub>3</sub> results in excellent piezoelectric properties ( $d_{33} = 242 - 400$  pC/N,  $k_p = 36 - 54$  %), when conventionally sintered in the temperature range of 1060 - 1120 °C for 3 h [189]. BaTiO<sub>3</sub> (5 mol. %) modified NKN exhibits high piezoelectric strain coefficient (225 pC/N), while sintered at 1060 °C for 2 h [190]. With the conventional sintering technique, it is difficult to achieve the desirable densification for NKN-based ceramics, which is required for the optimal dielectric and piezoelectric properties. Therefore, advanced processing routes have been adapted.

#### **2.4.2. Advanced processing routes**

A number of advanced processing routes have been demonstrated to provide better consolidation and piezoelectric properties in NKN-based ceramics. Hot pressing (1100 °C at

38.61 MPa for 20 min.) was the first non-conventional processing route, wherein comparatively refined microstructure and improved piezoelectric ( $d_{33} = 160$  pC/N) response were observed with highly dense ( $\sim 99\%$   $\rho_{th}$ ) NKN-based ceramic [79]. Similarly, Li modified and hot-pressed [850 – 1150 °C at  $\sim 7 - 69$  MPa for 1 h] (Na, K, Li) NbO<sub>3</sub> has been developed with excellent electromechanical coupling coefficient ( $k_p = 45\%$ ) along with a very high densification of 98.8 - 99.8 % of  $\rho_{th}$  [150]. The issues, associated with the consolidation of NKN-based ceramics have also been suggested to address via cold isostatic pressing (CIP) where, high pressure of about 100 to 200 MPa is applied to obtain highly dense ceramics with improved piezoelectric properties [62, 153, 154, 191]. Wang et al. [62] fabricated 50 % porous Li<sub>6</sub>Na<sub>5</sub>K<sub>44</sub>NbO<sub>3</sub> ceramic using CIP at a pressure of 100 MPa, followed by sintering at 1075 °C for 2h. Despite of significant porosity, the CIPed material has been demonstrated to possess reasonable piezoelectric properties ( $d_{33} = 117 \pm 6$  pC/N,  $k_p = 24 \pm 3\%$ ). CIPed (200 MPa) NKN-based ceramics, modified with BaTiO<sub>3</sub> (2 mol. %,  $T_s = 1100 - 1250$  °C), LiNbO<sub>3</sub> (5 to 7 mol. %,  $T_s = 1040 - 1100$  °C), LiTaO<sub>3</sub> (5 to 6 mol. %,  $T_s = 1110$  °C) or SrTiO<sub>3</sub> (0.5 to 2 %,  $T_s = 1100 - 1250$  °C) have been demonstrated to provide profound densification (97 - 98.4 %  $\rho_{th}$ ) with reasonable piezoelectric properties ( $d_{33} = 96 - 235$  pC/N,  $k_p = 29 - 44\%$ ) [Table 2.2] [153, 154, 155, 156]. It has been suggested that such kind of improvement in density occurs due to the increase in grain size. In addition, piezoelectric properties are enhanced due to the formation of MPB at the given composition of additives. Maiwa et al. [151] used hot isostatic pressing at temperature and pressure of 800 °C and 200 MPa (for 1 h, in argon atmosphere), respectively, to obtain NKN-based ceramic with a reasonable dielectric ( $\epsilon_r = 149$ ) and piezoelectric ( $d_{33} = 40$  pC/N) properties. Wang et al. [133] synthesized LiTaO<sub>3</sub> as well as Sb-modified NKN powder i.e., (Na<sub>0.52</sub>K<sub>0.4425</sub>)

$(\text{Nb}_{0.8825}\text{Sb}_{0.08})\text{O}_3 - 0.0375 \text{LiTaO}_3$  or (NKNS-3.75LT) with chemisorption sol-gel technique, where, perovskite structure of NKN was formed at a comparatively lower calcination temperature (at 500 – 600 °C) than the conventional fabrication methods (~ 800 °C). Sol-gel prepared  $(\text{Na}_{0.52}\text{K}_{0.4425})(\text{Nb}_{0.8825}\text{Sb}_{0.08})\text{O}_3 - 0.0375 \text{LiTaO}_3$  exhibits favorable piezoelectric properties ( $d_{33} \sim 418 \text{ pC/N}$ ,  $T_c \sim 257 \text{ °C}$  and  $k_p \sim 54 \%$ ) by conventional sintering at 1090 °C. Spark plasma sintering (SPS) is one of the most emerging non-conventional routes of sintering to obtain highly dense (~ 96 to 99.8 %  $\rho_{\text{th}}$ ) NKN-based ceramics, with excellent dielectric ( $\epsilon_r = 243 - 550$ ) as well as piezoelectric properties ( $d_{33} \sim 148 - 280 \text{ pC/N}$ ,  $k_p \sim 26 - 48 \%$ ) [152, 158, 171, 192, 193, 194]. These materials were consolidated under the combined action of high pressure (~ 0.6 GPa), rapid heating (~ 150 °C /min.) and at a comparatively lower sintering temperature (920 – 1100 °C). The reduced sintering temperature suppresses the alkali volatilization in NKN, which has very important consequences as far as the dielectric and piezoelectric properties are concerned. Chemically modified NKN with (8 mol. %)  $\text{LiTaO}_3$  as well as with Li/Ta/Nb i.e.,  $(\text{Li}_{0.04}\text{K}_{0.44}\text{Na}_{0.52})(\text{Nb}_{0.85}\text{Ta}_{0.15})$ , consolidated by SPS, exhibit adequate piezoelectric properties ( $d_{33} = 185 - 243 \text{ pC/N}$ ,  $k_p = 44 - 50 \%$ ) [Table 2.2] [195, 196, 197, 198]. Table 2.2 summarizes the influence of various sintering techniques on densification, piezoelectric strain coefficient ( $d_{33}$ ) and electromechanical coupling coefficient ( $k_p$ ) of NKN/ NKN-based ceramics.

**Table 2.2. Effect of various processing techniques on densification ( $\rho_{th}$ ), piezoelectric strain coefficient ( $d_{33}$ ) and electromechanical coupling coefficient ( $k_p$ ) of the NKN/ NKN-based ceramics.**

S.N.	Monolithic/ modified	Sintering temperature ( $T_s$ , °C)/ processing	% $\rho_{th}$	$d_{33}$ (pC/N)	$k_p$ (%)	Ref.
1.	Un-doped	1050 - 1325 (Conventional)	93.4 - 98.5	80 - 110	31 - 40	[132, 169, 172, 184, 185]
2.	Mg (1 mol. %) doped	1125 (Conv.)	97.1	-	45	[131]
3.	Modified with Li (4 to 7 mol. %)	1020 - 1100 (Conv.)	90.6 - 95.0	152 - 240	35 - 38	[186, 188]
4.	Modified with Li (4 mol. %) and Ta (15 mol. %)	1090 (Conv.)	98.0	206	38 - 40	[188]
5.	Modified with LiNbO <sub>3</sub> (5.8 mol. %)	1020 - 1080 (Conv.)	-	314	41.2	[187]
6.	Modified with LiTaO <sub>3</sub> (0 - 16 mol. %) and Sb (7 mol. %)	1060 - 1120 (Conv.)	-	242 - 400	36 - 54	[189]
7.	Modified with BaTiO <sub>3</sub> (5 mol. %)	1060 (Conv.)	90.0	225	36	[190]
8.	Modified with LiF (5 mol. %)	1020 (Conv.)	91.0	152	38	[148, 149]
9.	Un-doped	1080 - 1100 (Hot pressing)	99.0	160	48	[83]
10.	Modified with Li (5 mol. %)	850 - 1150 (Hot pressing)	98.8 - 99.8	-	-	[150]
11.	Un-doped	1120 (Cold isostatic pressing)	93.4	-	-	[191]
12.	Modified with Li (6 mol. %)	1075 (CIP)	94.34	123	27	[62]
13.	Modified with BaTiO <sub>3</sub> (2 mol. %)	1100 - 1250 (CIP)	98.4	104	29	[155]

14.	Modified with LiNbO <sub>3</sub> (5 to 6 mol. %)	1040 - 1100 (CIP)	-	235	44	[154]
15.	Modified with SrTiO <sub>3</sub> (5 mol. %)	1100 - 1250 (CIP)	98.4	96	32.5	[156]
16.	Modified with LiTaO <sub>3</sub> (5 mol. %)	1110 (CIP)	97.0	200	36	[153]
17.	Un-doped	1040 - 1100 (Spark plasma sintering)	98.0	-	38	[158]
18.	Un-doped	920 (SPS)	96.0 - 99.0	148	39 - 48	[152, 192, 193]
19.	Modified with LiTaO <sub>3</sub> (8 mol. %)	980 - 1000 (SPS)	98.0	185	50	[195]
20.	Modified with Li (5 mol. %) & Ta (1 mol. %)	850 - 1000 (SPS)	99.2	225 - 243	44 - 50	[196, 197, 198]
21.	Modified with Li (4 mol. %) & Ag (6 mol. %)	980 (SPS)	97.4	127	-	[194]
22.	Un-doped	800 (Hot isostatic pressing)	94	40	-	[151]
23.	Modified with LiF (5 mol. %)	1020 (Reduced atm. conv. sintering)	95.0	138	36	[148, 149]
24.	-	1090 (Sol gel - Conv. sintering)	102.66	418	54	[133]

### 2.4.3. Chemical modification

Apart from non-conventional sintering routes, property enhancement with chemical modification is a multifaceted and economical technique for the improvement in densification as well as piezoelectric and electrical properties of NKN-based ceramics. Malice et al. [199] investigated the effect of 0.5 atom. % doping of alkaline earth metals at A-site of NKN as  $(\text{Na}_{0.5}\text{K}_{0.5})_{1-2y} - \text{AE}_y\text{NbO}_3$  ( $y = .005$ ), (AE = Ca, Sr, Ba, Mg) which was

sintered at 1115 °C for 2 h. It has been found that Ca<sup>2+</sup> and Sr<sup>2+</sup> doping results in the improved piezoelectric property ( $d_{33}$  up to 95 pC/N) as compared to undoped NKN ( $d_{33} = 80$  pC/N), which is attributed to the modification in phase transition temperature [Table 2.3]. However, increased densification was only observed with Sr<sup>2+</sup> from 94.4 %  $\rho_{th}$  (undoped NKN) to 96.0 %  $\rho_{th}$ . Chang et al. [200] investigated the effect of the addition of 0.5 mol. % AETiO<sub>3</sub> (AE = Ca, Sr, Mg, Ba) with N<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub> after sintering at 1115 °C for 2 h. In perovskite structure of NKN, doping of Ca<sup>2+</sup> (ionic radii = 0.135 nm), Ba<sup>2+</sup> (0.160 nm), Sr<sup>2+</sup> (0.144 nm) on A-site ions such as, Na<sup>+</sup> (0.139 nm) and K<sup>+</sup> (0.164 nm) while, an addition of Ti<sup>4+</sup> (0.061 nm) on B-site Nb<sup>5+</sup> (0.064 nm) ion, results in an increase in unit cell parameters of NKN. While doping of Mg<sup>2+</sup> (0.072 nm) on A-site along with the addition of Ti<sup>4+</sup> on B-site results in the shrinkage of its unit cell parameters and causes poor and inhomogeneous microstructure. Therefore, MgTiO<sub>3</sub> (0.5 mol. %) addition to NKN, shows poor densification (90.9 %  $\rho_{th}$ ) and piezoelectric properties ( $d_{33} = 51$  pC/N,  $k_p = 15$  %,  $Q_m = 70$ ) as compared to undoped NKN ( $\rho_r = 96.4$  %  $\rho_{th}$ ,  $d_{33} = 122$  pC/N and  $k_p = 38$  %,  $Q_m = 124$ ). The increase in unit cell parameters contributes to the piezoelectric as well as electrical properties of NKN [Table 2.3]. Therefore, 0.5 mol. % of CaTiO<sub>3</sub>, SrTiO<sub>3</sub> and BaTiO<sub>3</sub> addition to NKN exhibit good densification (95.8 - 97.8 %  $\rho_{th}$ ) and relevant electrical properties ( $d_{33} = 85 - 116$  pC/N and  $k_p = 26 - 39$  %,  $Q_m = 156 - 180$ ) [200]. A similar experiment has been performed to analyze the effect of Ca<sup>2+</sup> and Ba<sup>2+</sup> dopants in N<sub>0.5</sub>K<sub>0.5</sub>NbO<sub>3</sub> upto 2 mol. % after sintering at the temperature of 1125 °C for 2 h [201]. The maximum densification and piezoelectric properties for both Ca<sup>2+</sup> or Ba<sup>2+</sup> dopants were observed at 0.5 mol. % addition, above which densification reduces. It has been suggested that Ca<sup>2+</sup> and Ba<sup>2+</sup> ions preferably replaces to Na<sup>+</sup> and K<sup>+</sup>, respectively, with almost similar ionic radii. Therefore, the doping of Ca<sup>2+</sup> ions,

above 0.5 mol. % reduces the densification, preferably due to the reduction of sodium phase [201]. However, the addition of  $\text{Ba}^{2+}$  at higher concentrations ( $> 0.5$  mol. %) reduces the densification and piezoelectric properties due to the formation of a potassium-based secondary phase [201]. Zuo et al. [132] also reported that the abnormal grain growth during sintering results in reduced consolidation and piezoelectric properties. The authors suggested that the addition of CdO (1 mol. %) to NKN-based ceramics shows abnormal grain growth behavior during sintering and exhibits lower densification ( $\rho_r = 95.3 \% \rho_{th}$ ,  $T_s = 1100 \text{ }^\circ\text{C} / 4$  h). However, similar densification was observed with ZnO-doped (1 mol. %) NKN-based ceramics at the lower sintering temperature ( $1000 \text{ }^\circ\text{C} / 4$  h), which undergo controlled grain growth. [132]. Further, it has been explored that the doping of similar valence ions on A and B sites such as,  $\text{Li}^+$  on A-site and  $\text{Ta}^{5+}$  and  $\text{Sb}^{5+}$  on B-site of  $\text{ABO}_3$  i.e.,  $(\text{Na}_{0.52}\text{K}_{0.44}\text{Li}_{0.04})(\text{Nb}_{0.86}\text{Ta}_{0.10}\text{Sb}_{0.04})\text{O}_3$  textured NKN-based ceramics results in better piezoelectric properties ( $d_{33} = 416 \text{ pC/N}$ ,  $k_p = 61 \%$ ) [202]. It has been suggested that the improvement in piezoelectric properties is observed due to the modification in phase boundaries after shifting of different phase transition temperatures, as discussed in pervious section [202]. It has also been investigated that the doping of  $\text{Zr}^{4+}$  or  $\text{Ti}^{4+}$  ions (0.5 mol. %) on the Nb site of  $\text{N}_{0.5}\text{K}_{0.5}\text{NbO}_3$  also results in enhancement in piezoelectric properties ( $d_{33} = 124 - 134 \text{ pC/N}$  and  $k_p = 33 - 35 \%$ ) [203]. It happens due to the formation of oxygen vacancy defects, which restricts the domain wall motion and results in stability in the piezoelectric behavior of ceramics [203]. The influence of grain size on the piezoelectric and electrical properties of ceramics can be understood in a way that grain boundaries are amorphous in nature, which reduces overall crystallinity and consequently, the ferroelectricity. As grain size decreases, the grain boundaries increases, which weakens the ferroelectricity of the material [204].

As previously discussed in section 2.3, that the formation of MPB in the processing of NKN-based ceramics results in improvement in the piezoelectric properties. Several additives have been reported that enhances the piezoelectric properties of  $N_{0.5}K_{0.5}NbO_3$  ceramics such as, 2 mol. %  $BaTiO_3$  ( $d_{33} = 104$  pC/N and  $k_p = 29$  %), 6 mol. %  $LiNbO_3$  ( $d_{33} = 235$  pC/N and  $k_p = 42$  %), 5 mol. %  $LiTaO_3$  ( $d_{33} = 200$  pC/N and  $k_p = 36$  %) and 2 mol. %  $SrTiO_3$  ( $d_{33} = 92$  pC/N and  $k_p = 26.6$  %) [153, 154, 155, 156]. It was demonstrated that the improved piezoelectric properties were attributed due to the formation MPB at the mentioned composition of additives. The effect of various dopants on the densification, piezoelectric properties, and dielectric constant are summarized in Table 2.3.

**Table 2.3. Effect of doping on densification, piezoelectric ( $d_{33}$ ) and dielectric ( $\epsilon_r$ ) properties of NKN.**

<b>Secondary phase</b>	<b>Increase/ decrease in densification (% <math>\rho_{th}</math>)</b>	<b><math>d_{33}</math> (pC/N)</b>	<b>Electromech. Coup. Coeff. (<math>k_p</math>)</b>	<b><math>\epsilon_r</math></b>	<b>Ref.</b>
$Ca^{++}$ (0.5 mol. %)	94.4 to 93.8 %	95	-	495	[199]
$Mg^{++}$ (0.5 mol. %)	94.4 to 91.2 %	-	-	-	..do..
$Sr^{++}$ (0.5 mol. %)	94.4 to 96.0 %	95	-	500	..do..
$Ba^{++}$ (0.5 mol. %)	94.4 to 94.1 %	-	-	-	..do..
$Ca^{++}$ (0.5 mol. %)	96.4 to 97.8 %	116	39 %	411	[200]
$Mg^{++}$ (0.5 mol. %)	96.4 to 90.9 %	51	15 %	480	..do..
$Sr^{++}$ (0.5 mol. %)	96.4 to 97.1 %	110	35 %	448	..do..
$Ba^{++}$ (0.5 mol. %)	96.4 to 95.8 %	85	26 %	393	..do..
ZnO (1 mol. %)	96.0 to 97.0 %	117	44 %	652	[132]
$SnO_2$ (1 mol. %)	96.0 to 98.0 %	108	39 %	627	..do..

Sc <sub>2</sub> O <sub>3</sub> (1 mol. %)	96.0 to 94.2 %	100	26 %	578	..do..
CdO (1 mol. %)	96.0 to 95.3 %	107	42 %	493	..do..
BaTiO <sub>3</sub> (2 mol. %)	96.0 to 98.4 %	104	29 %	1003	[155]
LiNbO <sub>3</sub> (6 mol. %)	96.2 to 96.4 %	235	42 %	-	[154]
LiTaO <sub>3</sub> (5 mol. %)	96.0 to 97.0 %	200	36 %	570	[153]
SrTiO <sub>3</sub> (2 mol. %)	96.2 to 98.0 %	92	26.6 %	873	[156]

## 2.5. Piezoelectricity and biocompatibility

Several studies suggest that the polarized piezoelectric bioceramics and biopolymers improve the osteogenesis through various ways [5, 38, 55, 56, 59, 63, 64, 65]. The electrical potential, induced in piezoelectric ceramics can potentially enhance the bioactivity and cellular response for hard tissue regeneration [Fig. 2.7] [17, 39, 42, 205, 206, 207].

### 2.5.1. Origin of biomineralization and cytocompatibility

*In vitro* biomineralization study on polarized piezoelectric BaTiO<sub>3</sub> ceramic demonstrated the deposition of calcium phosphate (CaP) layer on the negatively polarized surface, while no CaP layer was observed on the positively charged surface [Fig. 2.7 (A)] [39]. Due to electrostatic interaction between Ca<sup>2+</sup> ions, present in simulated body fluid (SBF) and negatively charged surface, deposition of Ca<sup>2+</sup> ions takes place on the polarized surfaces [39]. The anions (HPO<sub>4</sub><sup>-</sup> and OH<sup>-</sup>), in turn, interact with these deposited Ca<sup>2+</sup> ions and, form CaP layer on the negatively polarized surfaces [Fig. 2.7 (A)]. However, Cl<sup>-</sup> ions, present in SBF are attracted towards positively charged surfaces, which do not support the formation of the CaP layer [39] [Fig. 2.7 (A)]. Similarly, it has been demonstrated that the negatively polarized surface of HA- Li modified (6 mol.%) NKN (LNKN)-HA functionally graded

material almost doubled the rate of apatite formation [right part of Fig. 2.7 (A)] as compared to non-polarized monolithic HA (briefly explained in section 2.5.4.1.) [207]. Zhau et al. [206] elucidated enhanced cell proliferation on polarized Ti - 10 vol. % polyvinylidene fluoride (PVDF) polymer [Fig. 2.7 (B)]. The cations ( $\text{Ca}^{2+}$ ,  $\text{H}^+$  ions) from the biological fluid, adhere on the negatively charged surface. These cations further attract negatively charged proteins (like integrin, fibronectin) and cytomembrane. Consequently, the cell adhesion and proliferation on the negatively charged PVDF-Ti increase.  $\text{Ca}^{2+}$  ions, present in the apatite layer, activate cell adhesion factors [205] [Fig. 2.7 (C)].  $\text{Ca}^{2+}$  ions, adhered on the apatite rich polarized surface, enter into the cells and move towards the nearest cell [205]. Such kind of transportation of  $\text{Ca}^{2+}$  ions between the cells promotes the cell-cell adhesion and anchoring of cells on the CaP layer, formed on the negatively polarized ceramic surfaces [205]. Similarly, negatively polarized piezoelectric ceramic enhance the cell growth as compared to their positively polarized surfaces [Fig. 2.7 (D)] [17].

As discussed above, the negatively polarized surface attracts cations like,  $\text{Ca}^{2+}$  ions present in the biological fluid, a phenomenon which serves as a stimulus for the adsorption of proteins and subsequently, promotes cellular adhesion [17, 205]. However, positively charged surface attracts anions like  $\text{Cl}^-$ ,  $\text{OH}^-$ , etc, which restricts the adhesion of proteins as well as cells [17]. Apart from above mentioned *in vitro* response, *in vivo* assessment of unpolarized and polarized  $\beta$ -PVDF polymer samples was made by the implantation in rat's femur [42]. It has been observed that neo trabecular bone regeneration takes place on the polarized polymer samples after 4 weeks of implantation, which was almost absent in case of unpolarized polymers of identical macromolecular chemistry, for the similar period (4 weeks) of implantation [Fig. 2.7 (E)]. The mechanical stimulus, induced by physiological loadings in

rat's femur generates a voltage gradient on the polarized implant surfaces, which is responsible for the increased bone growth.

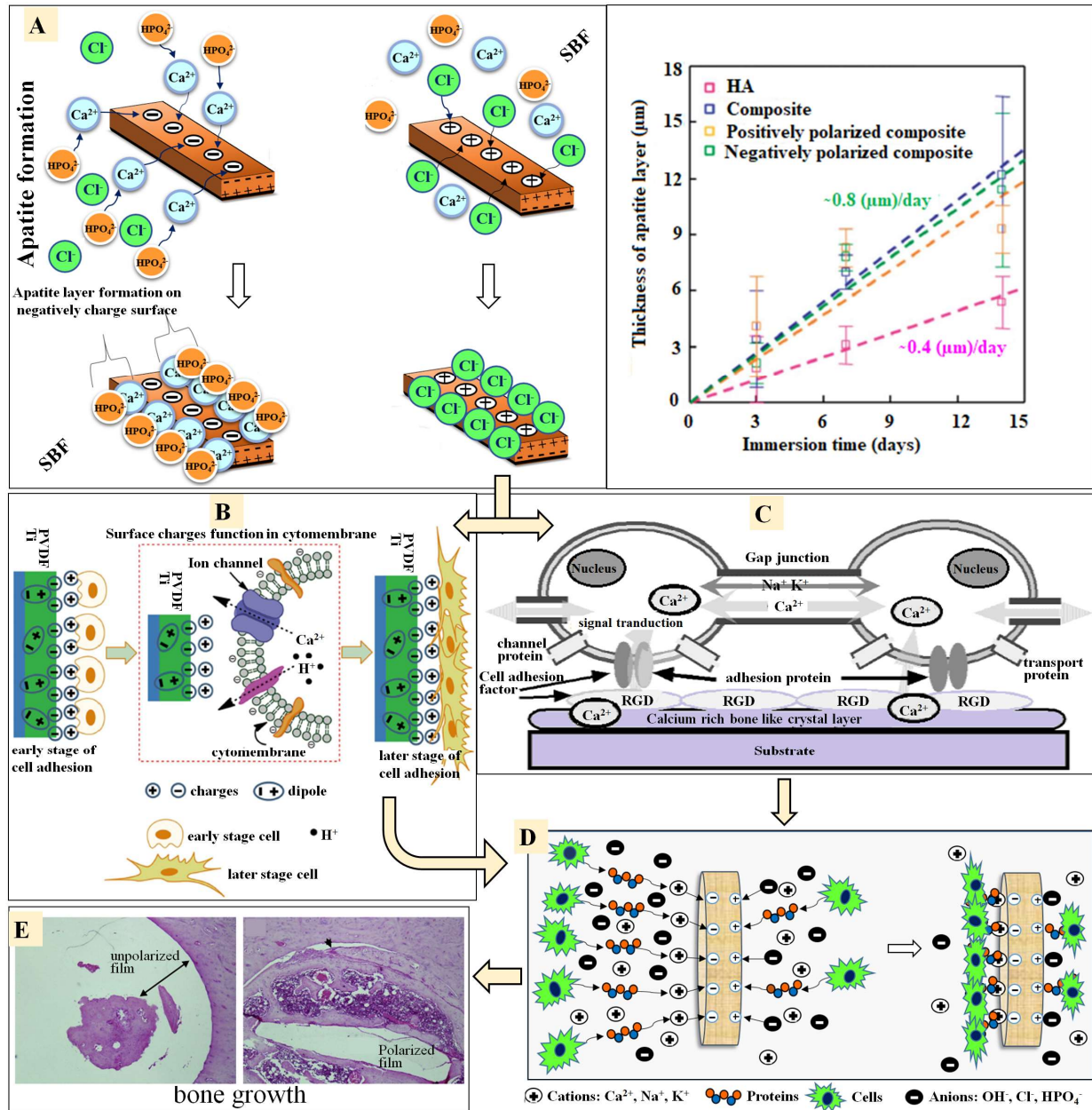


Fig. 2.7. Surface polarization induced bioactivity and cellular response, in vitro and in vitro.

Polarized surfaces of piezoelectric materials promote the adsorption of apatite ions ( $\text{Ca}^{2+}$ ,  $\text{HPO}_4^{2-}$ ) or proteins and cell adhesion, which consequently influence the hard tissue regeneration. (A) Schematic illustrating the mechanism of accelerated apatite layer

formation on negatively charged surface as compared to the positively charged surface [39]. The polarization induced enhanced rate of apatite formation on HA-LNKN-HA functionally graded material (FGM) as compared to unpolarized FGM and HA (Reproduced with the permission from publisher, Ref. [207]). (B) Schematic representing the polarization induced enhanced osteogenesis. Cations (like  $\text{Ca}^{2+}$  ions and  $\text{H}^+$  ions) from the physiological environment are attracted towards the negatively charged PVDF-Ti surface. Consequently, these cations attract negatively charged cytomembrane (Reproduced with the permission from the publisher, Ref. [206]). (C) An illustration of neuroblastoma cell response on polarized HA surface.  $\text{Na}^+$  and  $\text{Ca}^{2+}$  work as neurotransmitters between two cells through gap junction.  $\text{Ca}^{2+}$  activates protein adhesion factors and consequently, enters into the cells through these adhesive proteins and moves towards the neighboring cell through gap junction (Reproduced with the permission from the publisher, Ref. [205]). (D) Schematic demonstration of the influence of surface polarity on the interaction between the sample surface and constituents of culture media such as cations, anions, proteins and cells [17, 205]. Negatively charged surface attracts cations, followed by proteins as well as cells. Positively charged surface attracts anions, which inhibits the protein adsorption. (E) H & E stained images, representing the healing of defects (double arrow) on the unpolarized and polarized  $\beta$ -PVDF film, implanted in rat's femur for 4 weeks. Unpolarized  $\beta$ -PVDF implanted film could not exhibit neobone formation after 4 weeks of implantation; however, the defect on the polarized  $\beta$ -PVDF implanted film was about to be completely covered by bone marrow like spongy tissue and trabecular bone tissue after 4 weeks of implantation (Reproduced with the permission from the publisher, Ref. [42]).

### 2.5.2. Origin of bone healing

The damaged tissues can be healed by the piezoelectricity induced electrical stimulation as well as direct mechanical stimulation [30, 40, 44, 208]. These mechanisms can be elaborated through two independent moduli [Fig. 2.8]. According to the first module, piezoelectric scaffolds generate electrical potentials under the influence of functional loads. Such type of electrical stimulation activates voltage-gated  $\text{Ca}^{2+}$  channels and stretch activated calcium channels pathways, which results in an increase in intracellular  $\text{Ca}^{2+}$  ions [Fig. 2.8] [40, 44]. In addition, the local electrical field can modify the configuration of membrane receptors and opens receptor channels, which allow an influx of intracellular  $\text{Ca}^{2+}$  ions from the endoplasmic reticulum [Fig. 2.8] [40, 44]. These increased  $\text{Ca}^{2+}$  ions in cells activate calcium modulated proteins as well as calcineurin (a kind of calcium dependent protein phosphatase) [30, 40]. These calcineurin proteins react with the phosphorylated nuclear factor of activated cells ( $\text{NF-AT-PO}_4$ ) and convert it into dephosphorylated nuclear factors of activated cells ( $\text{NF-AT}$ ) [30, 40]. These dephosphorylated NFs-AT translocate to the nucleus, where they combine with other proteins and result in gene transcription [Fig. 2.8]. Gene transcription promotes the synthesis of  $\text{TGF-}\beta$  and BMP-2 and consequently, regulates cell metabolism and ECM synthesis [30, 40].

According to second module, mechanical stimulation directly activates mechanoreceptors, such as integrins [30, 209]. The activated integrins translocate protein kinase C (PKC) to the cell membrane and activate MAP Kinase (MAPK) pathways for transmembrane signaling [30, 209] and consequently, transfer extracellular mechanical signals to actin [209]. RACK 1 protein (intracellular receptor) associated with integrin, binds with the PKC domain. Therefore, RACK1 mediated PKC-integrin interaction plays a key role in transmembrane

signal transduction [30, 209, 210]. Now, these signaling cascades transfer towards the nucleus and interact with mechanosensitive transcription factors and consequently, excites gene transcription [30, 209].

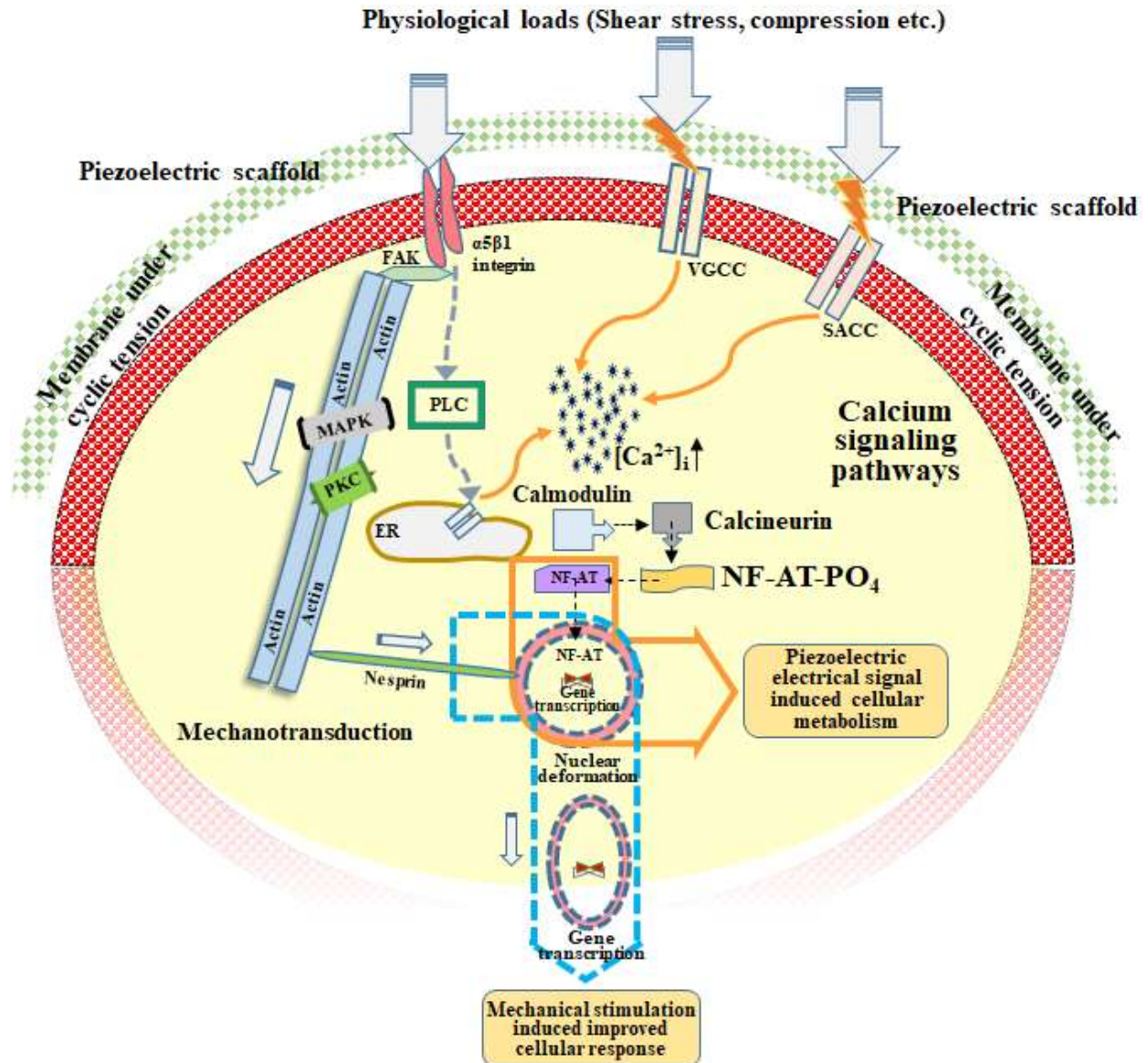


Fig. 2.8. Schematic illustration of the effect of mechanical stimulation on cell metabolism through intracellular electrical signal transduction, leading to activation of mechanoreceptors [30, 40, 44, 209]. Electromechanical signals facilitate outside-in  $Ca^{2+}$  ions influx through the opening of voltage gated  $Ca^{2+}$  channels (VGCC) and stretch activated

*calcium channels (SACC) [40, 44]. Consequently, intracellular ions increase which activate the calmodulated protein and calcineurin [30, 40]. These calcium activated calcineurin dephosphorylates the phosphorylated nuclear factor of activated cells (NF-AT) and shifts towards the nucleus, where it executes gene transcription [30, 40]. Mechanically stimulated membrane receptors open up receptor channels and permit intracellular  $Ca^{++}$  ions from endoplasmic reticulum calcium stores [40, 44]. Also, physiological loads directly activate mechanoreceptors of the cell membrane like integrin, which actuate PKC (protein kinase C) and MAPK (mitogen activated protein kinase) signaling pathways [30, 209, 210]. These signaling cascade propagates towards the nucleus, where it interacts with mechanosensitive transcription factor and results in gene transcription [30, 209, 210].*

The polarized (at 13 kV for 30 min.) piezoelectric BaTiO<sub>3</sub> - polymer [poly(vinylidene fluoride trifluoroethylene)] nanocomposite membranes demonstrated long lasting electrical charge storage capability. These biomaterials can facilitate the differentiation of rat bone marrow mesenchymal stem cells (BMSCs) into osteoblast cells [28]. The implantation of such polarized piezoelectric biocomposite in the calvarial site defect of rat bone has been demonstrated to promote early bone regeneration [28]. The mechanical deformation of bone collagen induces both, negative (under compression) as well as positive (under tension) potentials due to piezoelectricity [15]. Natural bones possess inherent negative zeta potential (- 5 mV) [15]. The negative charges, developed due to the compression of bone collagen, increase overall zeta potential as well as streaming potential in natural living bone [15]. This stress-stimulated electrical potential enhances the electroosmosis, and consequently, decreases the hydraulic permeability and increases the stiffness of bone [15].

### **2.5.3. Influence of electrical stimulation on the biological response**

The application of optimal electrical stimulation results in enhanced proliferation and differentiation of human mesenchymal stem cells on biomaterials surfaces, which subsequently, assists in bone repair and regeneration [16, 17, 21, 24, 25]. The cellular response can also be enhanced by improving the conductivity of the substrate surface in a controlled manner [25]. Apart from the improved cellular response, the application of electrical field with specific pulse rate results in electroporation, which can be utilized for cancer treatment [17, 24]. Electrical stimulation can also control the directional movement of cells [25, 27]. In absence of an electric field, a cell membrane exhibits negative potential. The application of a direct current electrical field (dcEF) hyperpolarizes the anodal side membrane of the cell [25, 27]. Consequently,  $\text{Ca}^{2+}$  ions diffuse at the anodal side membrane of the cell. This increased amount of  $\text{Ca}^{2+}$  ions on the anode side results in depolymerization of actin, which causes shrinkage in cell size at the anodal end. At the same time, due to the depolarization at the cathode side membrane of the cell, the number of  $\text{Ca}^{2+}$  ions decreases, which causes the polymerization of actin and protrusion on this side of the cell. Such kind of changes in cell morphology promotes the cell movement towards the cathode side [25, 27]. However, the depolarization of the cathode side membrane of the cell opens its voltage-gated  $\text{Ca}^{2+}$  channels and consequently, increases the amount of intracellular  $\text{Ca}^{2+}$  ions [25]. Therefore, the amount of  $\text{Ca}^{2+}$  ions on both, cathodal side (due to the influx of extracellular  $\text{Ca}^{2+}$  ions) and on the anodal side (due to diffusion), causes a net movement of cells either towards cathode side or anode side, where the number of  $\text{Ca}^{2+}$  ions are less [25]. In other words, the direct current electrical field (dcEF) can control the net movement of the cells [25, 27]. Such type of controlled cell locomotion through electrically modified endogenous

charges leads to an increased attachment of cells on the wounded site that assists in wound healing. Such a phenomenon also prevents cellular accumulation on the cancerous site which is used in cancer treatment.

The efficacy of a number of piezoelectric bioceramics and biopolymers as a prospective substitute for electrically-active bone tissue is described in subsequent sections, which reveal the potentiality of piezoelectric materials as next generation materials for orthopedic implant applications.

#### **2.5.4. Biomaterials-based perspective**

##### **2.5.4.1. NKN-based piezoelectric ceramics**

Nilson et al. [55] examined the biocompatibility of NKN using human monocytes. Jalalian et al. [57] fabricated the dense ferroelectric NKN nanofibers with the help of sol-gel electro-spinning route and reported cube on cube growth of NKN nano-crystals in the crystallographic direction [001]. Such textural evolution in nanofibers suggested good potential of the electrically polarized NKN nanofibers as a scaffold for repair and regrowth of damaged / injured tissues [57]. The space charge on the polarized surfaces of NKN-based ceramics plays an important role in modulating the biological responses, such as protein adsorption, cell proliferation, etc. [56, 62, 63].

Chen et al. [56] reported that the polarized (@ 25 kV/cm) NKN increases protein adsorption on both, positively ( $0.43 \text{ mg}\cdot\text{cm}^{-2}$ ) as well as negatively ( $0.45 \text{ mg}\cdot\text{cm}^{-2}$ ) charged surfaces as compared to unpolarized ( $0.34 \text{ mg}\cdot\text{cm}^{-2}$ ) surface. This study indicates that protein adsorption is independent of charge polarity. However, a significant increase in cell density has been reported on the negatively polarized surface than positively polarized as well as unpolarized surfaces, while cultured with MC3T3 osteoblasts cells [56]. More recently, Yao et al. [211]

investigated that polarized NKN shows significantly higher spreading and proliferation of rBMSCs cells as compared to unpolarized NKN. In another study, the addition (30 vol.%) of polarized NKN with 1393 bioglass results in comparatively higher cell proliferation than monolithic 1393 bioglass [212]. Lithium modified NKN ( $\text{Li}_{0.06}\text{Na}_{0.5}\text{K}_{0.44}\text{NbO}_3$ , LNKN) ceramics possess excellent piezoelectric properties ( $d_{33} = 222$ ) [213], alongwith superior biocompatibility as compared to unmodified NKN [61, 62, 63]. LNKN (polarized at  $\sim 22 - 24$  kV/cm) exhibits excellent chemical stability and hydrophilicity against biological fluid, which indicates its suitability for orthopedic applications [61]. Electrically treated LNKN surface results in the rapid rate of apatite formation [63]. LNKN (polarized at E-field intensity of  $22 - 24$  kV/cm) supports 33 % higher growth of osteoblasts on the polarized surface, as compared to that on the unpolarized surface [62]. The NKN has been potentially utilized to induce the piezoelectricity in HA-NKN functionally graded material (FGM) [207, 214]. In FGM development, the piezoelectric NKN layer has been introduced between HA layers [214]. Such a concept of FGM development has been reported to increase the polarizability of HA by more than three times without affecting its excellent biocompatibility. However, the incorporation of the LNKN layer between HA layers increases the polarizability up to six times [207].

As discussed earlier, polarization plays very important role in cellular functioning. The augmented polarizability of HA can potentially increase cellular growth and proliferation. The findings of above-mentioned studies are quite appealing in terms of augmented *in vitro* biocompatibility, which is attributed to the surface charge potential of the polarized piezoelectric NKN. Therefore, the clinical implications of polarized NKN based ceramics can be realized as the physiological loads can generate surface electrical charge, which can

stimulate bone regeneration [30, 40]. Apart from the applications in bone tissue engineering, NKN-based ceramics can be utilized in nerve and skin replenishment as well as drug delivery [215].

#### **2.5.4.2. Other piezoelectric ceramics**

BaTiO<sub>3</sub> exhibits phase transformation from symmetrical cubic (paraelectric or non-polar) to an asymmetric (ferroelectric or polar) structure below its Curie temperature (130 °C), which occurs due to the relative atomic displacements between the positive centroid Ti<sup>4+</sup> and face centered O<sup>2-</sup> ions [5, 39]. Hwang et al. [39] performed *in vitro* bioactivity on BaTiO<sub>3</sub> ceramics, polarized above and below its Curie temperature. The Ca/ P ratio was recorded to be 1.5 - 1.7 on negatively charged BaTiO<sub>3</sub> surface, polarized above Curie temperature (i.e., 160 °C, E<sub>p</sub> = 5 kV/cm). However, this ratio was 1.2 to 1.5 on the negatively charged surface, when polarized below Curie temperature at the same polarizing field [39]. A similar experiment has been performed by Park et al. [216] on BaTiO<sub>3</sub> ceramics, polarized above its Curie temperature (i.e., T = 160 °C, E<sub>p</sub> = 5 kV/cm). A thick layer of CaP has been observed on the negatively charged surface of the piezoelectric sample [Fig. 2.7 (A)]. Similarly, negatively charged BaTiO<sub>3</sub> thin film has also been demonstrated to enhance the cellular functionality of L929 mouse fibroblast cells than positively charged as well as uncharged films [217]. The anticipated toxicity concern of BaTiO<sub>3</sub> particulates has been addressed by intra-articular injection of HA-40 wt.% BaTiO<sub>3</sub> particles (up to 25 mg/ml ) in the knee joint of mice (Fig. 2.9) [218]. The activity and weight of mice were observed to be normal up to 7 days of post-injection. Histopathological analyses suggest that the injected particles were neither translocated to any of the major organs such as heart, kidney, liver, lung etc. nor caused any adverse effect to these organs [218]. These results confirm the absence of any

systemic toxicity of such piezo-biocomposites. It has been demonstrated that the injected particles were accumulated in the close vicinity of knee joint region without causing any inflammatory/ foreign body reaction [218].

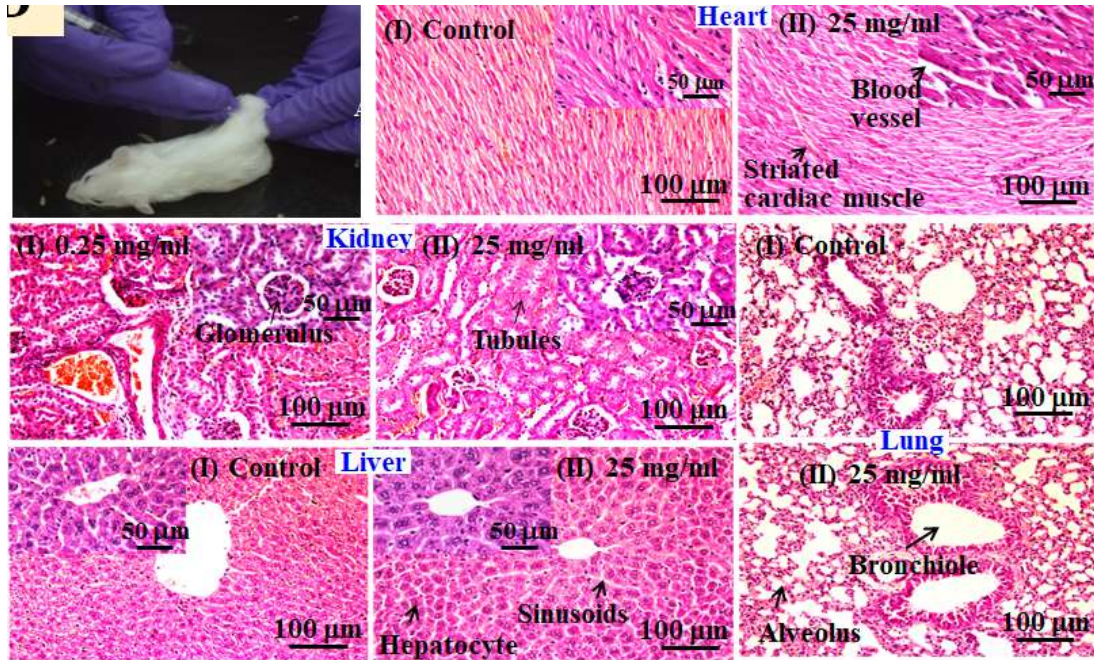


Fig. 2.9. Intra-articularly injected HA-40 wt.% BaTiO<sub>3</sub> particulate in the knee joint of mice could not cause any systematic toxicity in vital organs, such as heart, kidney, liver and lung at 7 days of post-injection, as evident from representative fluorescent images (Reproduced with the permission from the publisher, Ref. [218]).

MgSiO<sub>3</sub> (MS) possesses an asymmetric tetragonal perovskite structure, which is responsible for its piezoelectricity and spontaneous polarization [93]. The biodegradable nature of MgSiO<sub>3</sub> facilitates the piezoelectric implant to be easily replaced by newly developed bone tissue [136]. A porous scaffold consisting of HA nanowire - MgSiO<sub>3</sub> nanocomposite with Chitosan polymer i.e., HA-MS-CS [(HA-MS) : CS = 7 : 3 weight ratio] demonstrates enhanced rat BMSC [135]. Although, biodegradability facilitates cell growth and bone

regeneration with MgSiO<sub>3</sub> based scaffolds, the uncontrolled biodegradability limits their *in vivo* trials upto small animal models.

LiNbO<sub>3</sub> is a well-known ferroelectric material with excellent polarizability (spontaneous polarization,  $P_s = 78 \mu\text{C}/\text{cm}^2$ ) [137] at room temperature, which can be easily reversed by the application of the electric field. Apart from stable polarizability, LiNbO<sub>3</sub> also possesses good pyroelectric ( $P_r = 103.9 \mu\text{C}/\text{m}^2\text{K}$ ) [91] as well as piezoelectric properties ( $d_{33} = 23 \text{ pC/N}$ ) [65]. The electroactive LiNbO<sub>3</sub> surface adsorbs the organic molecules, like d-cysteine, which helps in stimulating polarized LiNbO<sub>3</sub> surface for protein adsorption [219]. Carville et al. [64] demonstrated that the charged surfaces of LiNbO<sub>3</sub> induces ionic exchange and protein interaction from cell media which consequently, results in significant enhancement in the proliferation of MC3T3 osteoblast cells, irrespective of charge polarity. In another study, it has been reported that the negatively charged surfaces facilitate significant cellular (NIH-3T3) elongation with more spreaded cell morphology as compared to the positively charged surfaces [137]. The negatively polarized LiNbO<sub>3</sub> surface promote the adhesion of cations, which further attracts electronegatively charged cellular membrane. However, positively charged surface attracts negatively charged ions, which restricts the adhesion of cells and in fact, causes cell migration [137]. In contrast, positively charged surface of LiNbO<sub>3</sub> exhibits 15% higher wound healing (cellular migration) capability than the negatively charged surface, which was almost double than that of the glass control surface [137]. On the other hand, Vanek et al. [65] elucidated a different hypothesis that the positively charged surface of polarized LiNbO<sub>3</sub> substrate preferably adsorbs electronegatively charged cell-adhesion proteins such as fibronectin and vitronectin. Therefore, the cell differentiation is promoted on

the positively charged surface (0.037 ALP /  $\mu\text{g}$  of BCA protein) as compared to the negatively charged surface (0.030 ALP /  $\mu\text{g}$  of BCA protein) of  $\text{LiNbO}_3$  [65].

$\text{KNbO}_3$  is an emerging piezoelectric perovskite material having its application in the biomedical field as a bio-probe for disease diagnosis [69, 138, 139].  $\text{KNbO}_3$  nanowires also find their application in scanning light imaging in life sciences, due to their second harmonic generation with larger non-linear stable polarizability, at room temperature. Therefore, it has the ability to be used as a tunable coherent light source [138].  $\text{KNbO}_3$  nanocrystals, coated with amino dextrin (AD), have been demonstrated as a bio-marker for the diagnosis of non-tumor cells, as AD coated  $\text{KNbO}_3$  nano-crystals suitably attach with the human lung derived BEAS-2B non-tumor cells [139]. In addition,  $\text{KNbO}_3$  nanoparticles are suggested to be highly toxic for prostate cancerous cells (DU-145), which enables the application of  $\text{KNbO}_3$  in prostate cancer treatment [69].

Zinc has been proved as an important element that stimulates the osteoblastic activities [220]. Owing to its piezoelectric nature, nanosized  $\text{ZnO}$  sheets develop a local electric field in the response of inherent mechanical force from the cells, which improves the metabolic activity of human SaOS-2 osteoblast-like cells and macrophages [44]. Due to the inherent stress from cells on the piezoelectric nanosheets and locally developed electrical charges, the membrane potential of adjacent cell changes. Consequently, voltage gated  $\text{Ca}^{2+}$  channels and stretch activated  $\text{Ca}^{2+}$  channels open, which allows the influx of extracellular  $\text{Ca}^{2+}$  ions [44]. There are number of literature reports which suggest that an optimized amount of  $\text{ZnO}$  reinforcement to a biomaterial causes a reasonable increase in the compatibility with fibroblasts, osteoblasts, stem cells, etc. [221, 222, 223].

Nakhmanson et al. [224] investigated the piezoelectricity and spontaneous polarization in boron nitride nanotubes (BNNT). BNNT supports the differentiation of hMSCs into osteoblast cells by releasing a trace amount of boron in culture medium and the stress applied from BNNT fibers on the hMSC cells [70]. Such kind of stress causes stretching of cells that activate actin filament and consequently, results in enhanced osteogenesis [70]. Another factor, which promotes the osteogenesis of BNNT is their positive affinity for cellular protein [225]. The adherence and proliferation of MG-63 and hMSCs are enhanced on Ackermine - BN composite scaffold [226]. The electrospun mats, prepared by the reinforcement of 1 to 5 wt. % boron nitride on gelatin polymer, exhibit good biomineralization and cytocompatibility with HOS human osteosarcoma cells [227].

#### **2.5.4.3. Piezoelectric polymers**

The inherent piezoelectricity and biocompatibility of a specific class of polymers make them suitable for bone tissue engineering applications. These polymers can be classified as synthetic and natural piezoelectric polymers. The synthetic piezoelectric biocompatible polymers include  $\beta$ ,  $\gamma$  or  $\delta$  phases of polyvinylidene fluoride (PVDF) and its copolymer, poly-L-lactic acid (PLLA), polyhydroxybutyrate (PHB) and polyamides (Nylons, peptides, etc.), etc. Some of the biocompatible polysaccharides such as, cellulose (wood, ramie), chitosan, amylose (starch), and proteins such as, collagen, silk, keratin possess inherent piezoelectricity, which are categorized as natural piezoelectric polymers.

The piezoelectricity has been observed in the stretched and polarized film of PVDF polymer [228].  $\beta$ ,  $\gamma$ , and  $\delta$  phases of PVDF possess permanent dipole moment and exhibit piezoelectricity [103]. The biocompatibility of piezoelectric PVDF polymeric substrates has been investigated with MC3T3-E1 osteoblast cells [29, 229, 230, 231, 232], human

mesenchymal stem cells (hMSCs) [233, 234], human adipose stem cells (hASC) [235, 236], and goat marrow stem cells [237]. Poly (L-Lactic) acid or PLLA is another synthetic polymer with reasonable piezoelectricity ( $d_{14} = -10$  pC/N) and biodegradability [123, 238]. Barroca et al. [239] demonstrated the surface polarization induced fibronectin adsorption on PLLA film, prepared by spin coating. PLLA is currently being used to make fastening devices for orthopedic application such as bone plate, screws, pins, washers etc. due to its excellent ossiointegration [240, 241]. The piezoelectric poly 3-hydroxybutyrate or PHB ( $d_{14} = 1.3$  pC/N) is a member of a polyester polyhydroxyalkanoates (PHA), which has been demonstrated to be biodegradable [242, 243, 244]. Several studies have been performed with PHB and its copolymers such as, PHBHV or poly (3-hydroxybutyrate-co-3-hydroxyvalerate) and PHBHH or poly-(hydroxybutyrate-co-hydroxyhexanoate)-based polymers to investigate their biocompatibility, *in vitro* [73, 245, 246, 247, 248, 249, 250].

Collagen is a natural protein, found in living bone and due to its non-centrosymmetric structure, bone exhibits piezoelectricity [6, 251]. Collagen has been proved as an excellent biocompatible material due to its reasonable cell binding ability and low antigenicity [78, 252]. In addition, the collagen-based scaffold has been reported to support the functionality of SaOS-2 osteoblast-like cells [253]. However, high degradation rate and low stiffness of natural polymers put the concern on their mechanical strength [252, 254]. Chitosan is another piezoelectric natural polysaccharide polymer, as reviewed by Martino et al. [75]. It has been demonstrated that chitosan-based polymer scaffolds exhibit several favorable characteristics to be used as orthopedic implant such as, osteoconductivity, porosity, easy molding ability, antibacterial response and minimum foreign body reaction [75]. Cellulose is another piezoelectric polysaccharide natural polymer, which can be a suitable candidate for bone

tissue applications due to its superior biocompatibility and good mechanical strength. Zaborowska et al. [77] observed that cellulose-based microporous scaffold results in significantly higher proliferation of MC3T3-E1 osteoblast cells as compared to the nanoporous scaffold.

## **2.6. Surface charge polarization induced antibacterial response**

### **2.6.1. Bacterial infection**

As far as the issues associated with the development of prosthetic implants are concerned, bacterial infection is among the critical issues which lead to implant failure [255, 256, 257]. The gram positive cocci (*Streptococci*, *Staphylococcus aureus*, *coagulase-negative staphylococci*) and gram negative bacteria (*Pseudomonas aeruginosa*, *Escherichia Coli* etc.) are the common bacteria associated with the infection in the prosthetic orthopedic implants [258, 259]. In general, bacteria reach the implant surface through the blood vessels route and contain a capsule of extracellular matrix (ECM) protein around them which helps in colonizing bacteria and subsequently, form a three dimensional bacterial biofilm on the implant surface [260, 261, 262]. This biofilm protects bacteria from the immune defense system and antibiotics [263, 264, 265]. Sometimes, the antibodies or immune cells, produced by the immune defense system to neutralize biofilm antigens, fail to penetrate the bacterial biofilm [266, 267, 268]. For this purpose, antibiotics have been recognized to be a suitable alternative. However, the bacteria develop resistance against antibiotics in a dose and time dependent manner [268]. Therefore, antibiotics do not remain effective for a longer duration.

### **2.6.2. Influence of surface charge and other external stimuli on bacteria**

Gram negative bacteria consists of two layers, a thin layer of peptidoglycan (~ 5 - 10 nm), surrounded by an outer layer of lipopolysaccharides. However, gram positive bacterial cell

wall contains a single thick layer of peptidoglycan (~ 20 - 80 nm) [269, 270, 271, 272]. Although, both the bacterial cells are negatively charged, the *E. coli* bacteria possess about 1.5 times higher negative charge than that of gram positive bacteria [273, 274]. Due to specific electrical surface potentials, bacteria respond against electric or magnetic field which generates reactive oxygen species (ROS) that damage bacterial cell wall and consequently, kills the bacteria [275, 276, 277, 278, 279]. It has been reported that the magnetic field (100 mT for 4 h) inhibits the growth of *E. coli* and *S. epidermidis* bacterial cells [275, 276, 277]. In addition, the exposure of external electric field (direct current ~ 100 mA or direct current electric field ~ 1 V/cm) hinders the growth of biofilm on the implant and consequently, enhances the antibacterial response [278, 279]. Bacteria interact with polarized piezoelectric substrates by means of microelectric field of charged surfaces and electrical potential of bacterial membranes [211, 280, 281, 282]. Recently, Verma et al. [212, 283, 284] reported the surface charge induced antibacterial response of polarized piezoelectric biocomposites. Tan et al. [280] and Yao et al. [211] manipulated the electrostatic interaction by polarizing (25 kV/cm for 30 min) sodium potassium niobate ( $\text{Na}_{0.5}\text{K}_{0.5}\text{NbO}_3$ ) piezoceramic surfaces against negatively charged membranes of *S. aureus* bacterial cells. Since, bacterial cell membrane exhibits electrical charge, it interacts with polarized surfaces. The microelectric field generates on the polarized surfaces, enhances the rate of electrolysis of water that causes breakage of hydrogen bond and consequently, produces ROS such as  $\text{H}_2\text{O}_2$ ,  $\text{O}_2^-$  etc [211, 281, 285, 286]. Such ROS are toxic for the bacterial cells [211, 280, 281, 287]. Swain et al. [282] demonstrated that the polarization (4.5 kV/cm) of barium titanate – hydroxyapatite piezo-biocomposite surfaces increase the zone of inhibition (> 2 times) for *S. aureus*, *E. coli* and *P. aeruginosa* bacteria as compared to non-polarized composite surfaces.

Apart from the positively polarized surfaces, negatively polarized surfaces have also been suggested to reduce bacterial adhesion due to electrostatic repulsion between mutually opposite charged surfaces of substrate and bacteria [281, 287]. In addition, the negatively polarized surface shows excellent hydrophilicity, which reduces the bacterial cell population [17, 288, 289, 290, 291]. Singh et al. [281] reported that the viability of *E. coli* and *S. aureus* bacterial cells are reduced by 53% and 40 %, respectively on negatively charged surfaces of HA - 7.5 % ZnO bioceramic composite. External electrical or magnetic stimulation needs sophisticated device installation near the patient and tool insertion in the human body to reach to the infected implant surface during treatment which may cause additional concerns. Therefore, surface electrical treatment or surface polarization can be regarded as a suitable technique to induce the antibacterial property in prosthetic implants, prior to implantation.

Recently, it has been demonstrated that the addition of the  $\text{Na}_{0.5}\text{K}_{0.5}\text{NbO}_3$  (30 vol. %) phase in HA and bioglasses (45S5 and 1393) enhances the antibacterial response of biocomposites where the inhibition of bacterial strains is observed to depend on surface charge polarity [212, 283, 284]. Apart from surface charge polarization, the presence of sodium and potassium (individual or in combination) in bacterial extracellular matrices creates an unfavorable environment for the growth of bacteria and consequently, enhances the antibacterial response [292, 293]. The antibacterial performance of any material system depends on the number of factors such as surface charge, surface chemistry, oxidative stress etc. [292, 293, 294, 295].

The piezoelectricity has been demonstrated as an intrinsic functional property to regulate the bone metabolism. In a continuous surge towards bone mimicking materials, the development of piezoelectric bioceramics and biopolymers has been discussed herein for bone tissue

engineering applications. Towards this end, number of processing related challenges, such as, densification, volatilization of alkali elements for a range of compositionally tuned piezoelectric bioceramics, in particular reference to sodium potassium niobate, have been discussed. Importantly, the piezoelectricity / electrical stimulation induced augmented bioactivity, cellular response, tissue regeneration, etc. have been highlighted with a specific emphasis on the biocompatibility of polarized piezoelectric bioceramics and biopolymers. Different ways to overcome bacterial infection on implants such as antibiotics, external stimuli (electrical and magnetic field) and surface charge polarization have been comprehensively analyzed along with their limitations. In addition, recent progress on demonstrating the efficacy of the surface charge polarization induced antibacterial response of bioceramics was also discussed.

Despite various available literatures demonstrate the potentiality of piezoelectric ceramics, however, actual mechanisms of electrical stimulation induced improved biological response of piezoelectric biomaterials are still unclear. The combined effect of electrostatic and dynamic electrical stimulation have not been explored well, as such stimuli positively affect the cellular or antibacterial response individually. Further, the effect of surface polarization on the surface characteristics such as, surface chemistry or wettability should be investigated, as such surface properties directly influence the cellular and bacterial response. The leaching behavior of biomaterials is another important factor that may affect their performance. It has been hypothesized that the external electrical stimulation influences the voltage-gated signaling pathways and thereby, improve the osteogenic response however, the actual quantitative study have not been performed yet.

By considering the above backdrops, the present study reveals the influence of the combined action of electrostatic surface polarization charge and dynamic pulsed electrical stimulation alongwith compositional modification towards improving the osteogenic response of emerging piezo-bioceramics, sodium potassium niobate [ $\text{Na}_x\text{K}_{1-x}\text{NbO}_3$  ( $x = 0.2 - 0.8$ ), NKN] prepared by cold isostatic pressing (Pressure: 330 MPa). Thereafter, the synergistic effect of surface polarization and compositional variation of Na and K contents in biocompatible  $\text{Na}_x\text{K}_{1-x}\text{NbO}_3$  ( $x = 0.2$  to  $0.8$ ) was investigated in improving their antibacterial performance. An *in vivo* study to examine the local and systemic toxicity of NKN nanoparticulates in rat's model has also been performed.

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