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# **Appendix**





## Appendix 3 (a)

Table 3 (A). Biodegradation of *p*-cresol by various microorganisms

| Microorganism                                  | Max. Conc. (mg L <sup>-1</sup> ) of PC | Operating condition             | Removal time            | Author                       |
|--|--|---------------------------------|-------------------------|------------------------------|
| <i>Stenotrophomonas sp.</i> (MF004205)         | 500 mg L <sup>-1</sup>                 | 27 °C, pH 7                     | 100% in 48 h            | Bera et al., (2019)          |
| <i>Serratia marcescens</i> ABHI001             | 85 mg L <sup>-1</sup>                  | Not available                   | 85% in 18 h             | Singh et al., (2017)         |
| <i>Pseudomonas putida</i>                      | 200 mg L <sup>-1</sup>                 | 35 °C, pH 8, Immobilized on PVA | 85% in 4 h              | Surkatti et al., (2014)      |
| <i>Pseudomonas spp.</i>                        | 100 mg L <sup>-1</sup>                 | 30 °C, pH 8                     | 100% in 36.9 h          | Hamitouche et al., (2014)    |
| <i>Bacillus sp.</i> PHN 1                      | 4326 mg L <sup>-1</sup>                | 30 °C, pH 7                     | 96% in 120 h            | Tallur et al., (2009)        |
| <i>Pseudomonas citronellolis</i> NS1           | 1200 mg L <sup>-1</sup>                | Not available                   | 100% in 40 h            | Panigrahy et al., (2020)     |
| <i>Advenella sp.</i>                           | 750 mg L <sup>-1</sup>                 | 30 °C, pH 7                     | 100% in less than 100 h | Xenofontos et al., (2016)    |
| <i>Bacillus sp.</i> strain PHN 1               | 200 mg L <sup>-1</sup>                 | room temp., pH 7.5, 150 rpm     | 100% removal in 168 h   | Tallur et al., (2006)        |
| <i>Stenotrophomonas maltophilia</i> strain KB2 | 540 mg L <sup>-1</sup>                 | 30 °C, pH 7.1, 130 rpm          | 100% removal in 24 h    | Wojcieszynska et al., (2011) |
| <i>Arthrobacter sp.</i> W1                     | 500 mg L <sup>-1</sup>                 | 30 °C, pH 7.0, 150 rpm          | 100% removal in 88 h    | Wang et al., (2009)          |

## Appendix 3 (b)

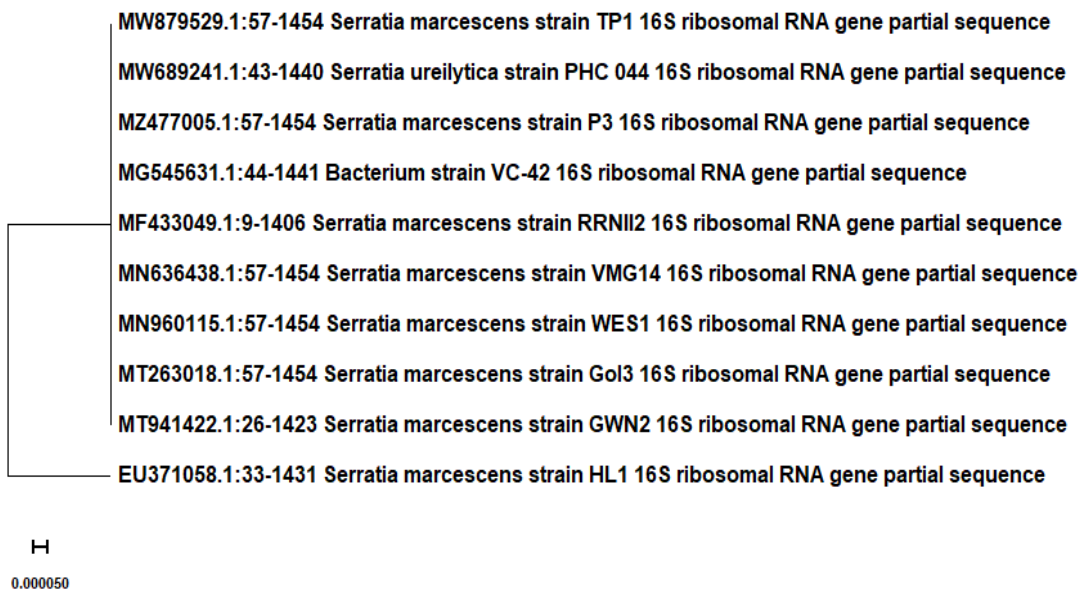


Fig. (A-3.1) Phylogenetic tree of potential bacterial species *Serratia marcescens* strain HL 1 after the characterization the 16S rRNA technique

## Appendix 3 (c)

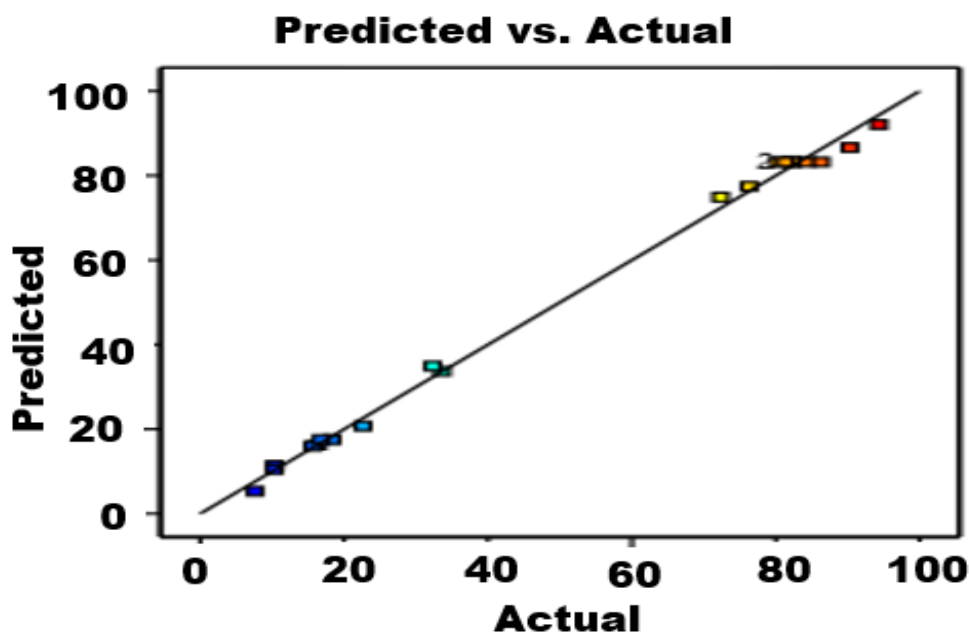
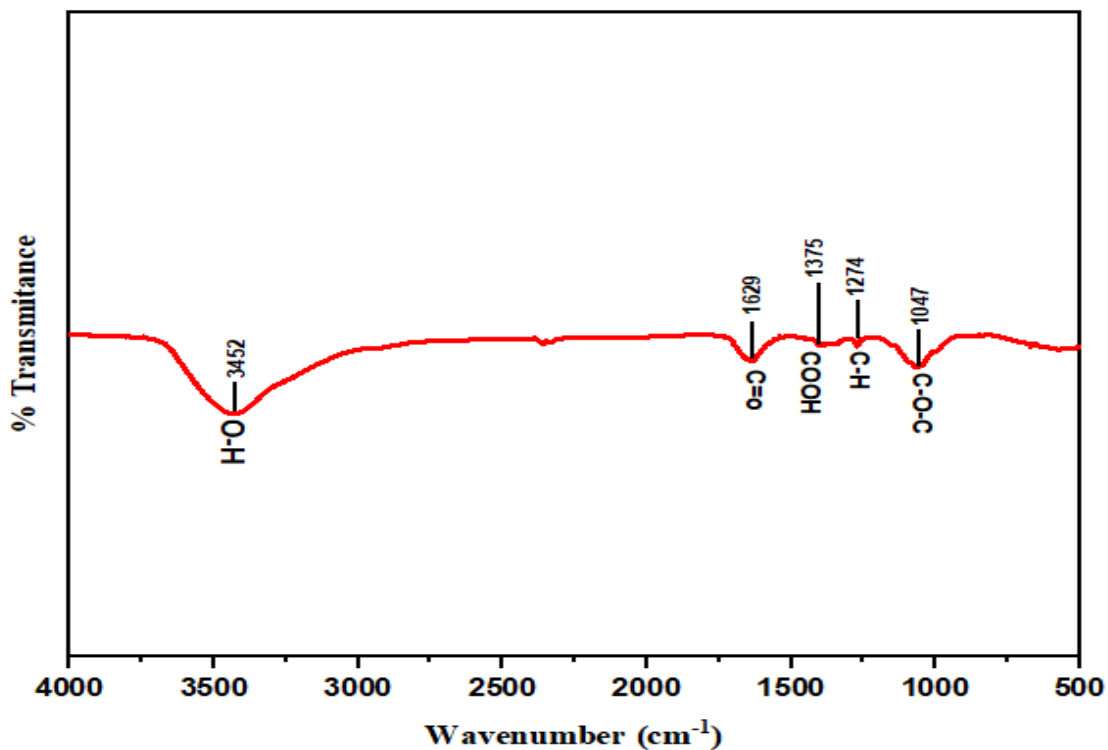


Fig. (A-3.2) Relationship between actual and predicted value

## Appendix 3 (d)

Fig. (A-3.3) FTIR spectra of *p*-cresol biodegradation

## Appendix 3 (e)

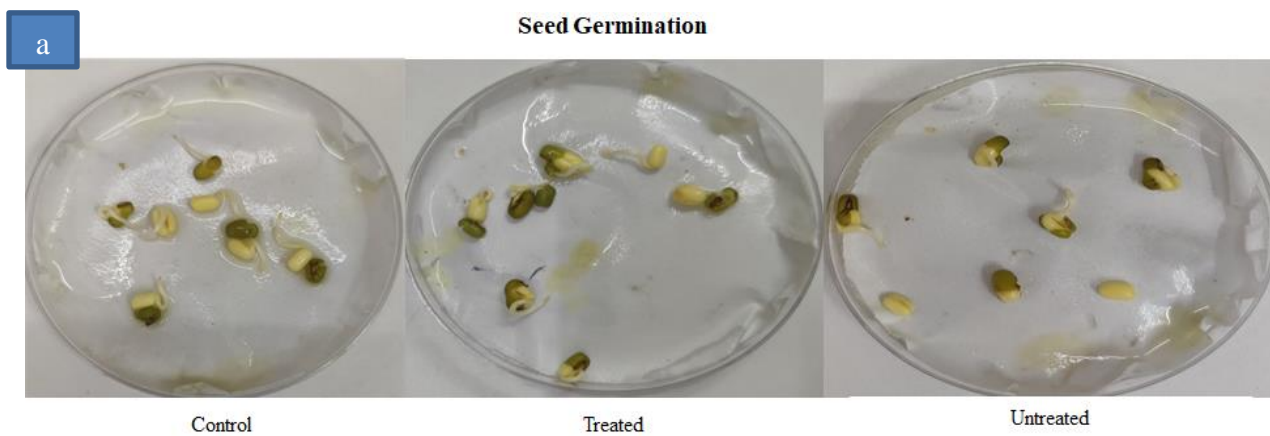
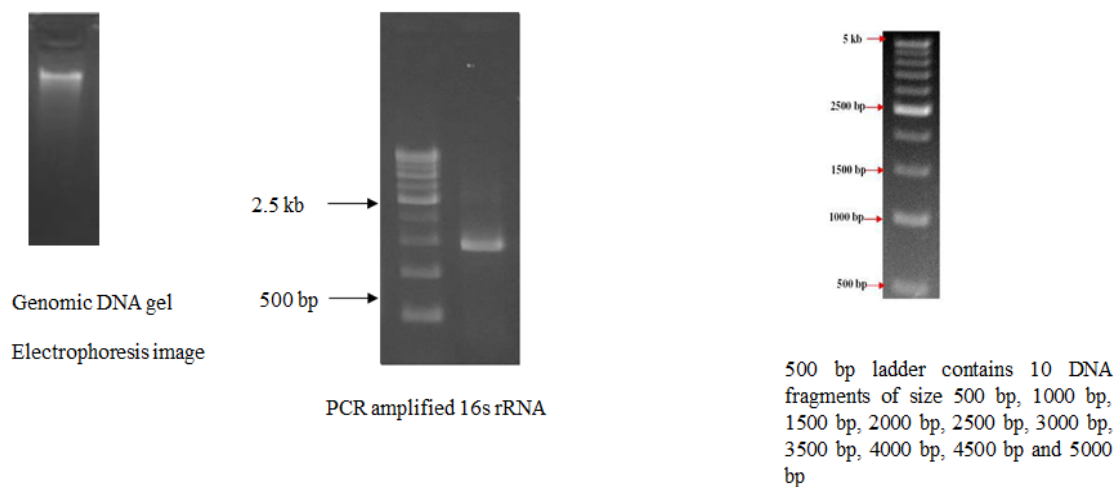


Fig. (A-3.4) Seed germination



**Fig. (A-3.5) Seed growth**

**Appendix 4 (a)**

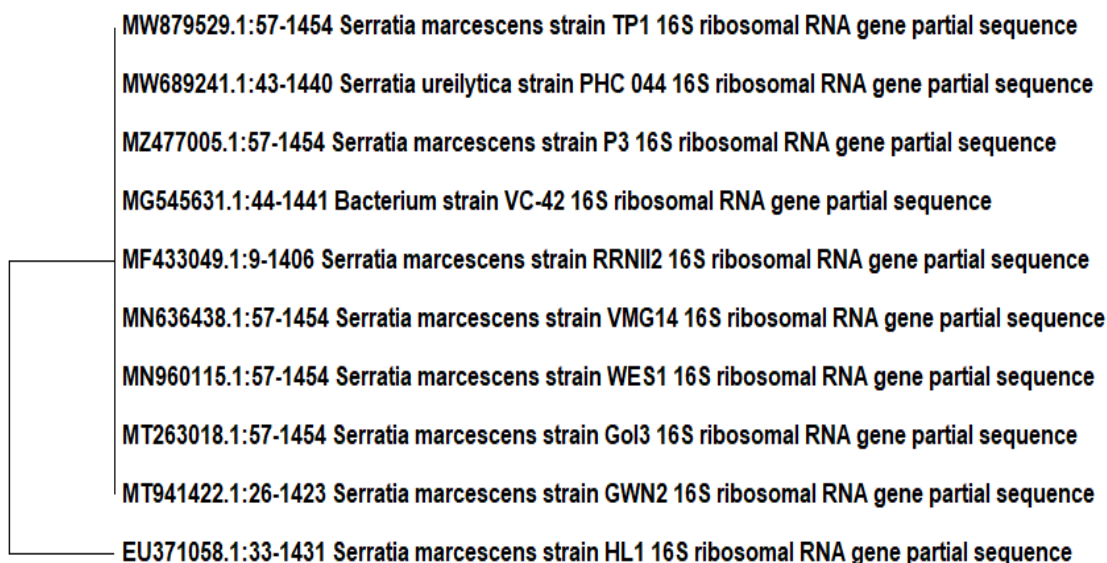


**Fig. (A-4.1) Aligned Sequence (Data of Sample PCR amplification images)**

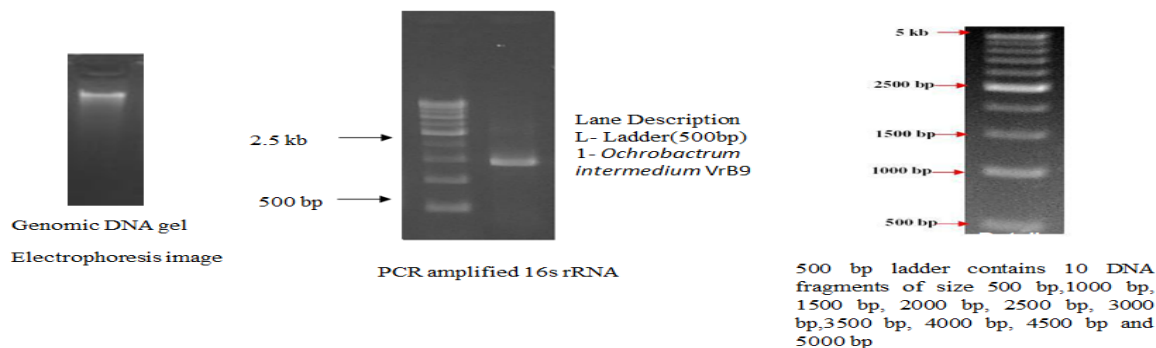
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TCATGGAGTCGAGTTGCAGACTCCAATCCGGACTACGACGTA CTTTATGAGGTCCGCTTGCTCT
CGCGAGGTCGCTTCTCTTTGTATACGCCATTGTAGCACGTGTGTAGCCCTACTCGTAAGGGCCA
TGATGACTTGACGTCATCCCCACCTTCCCTCCAGTTTATCACTGGCAGTCTCCTTTGAGTTCCCG
GCCGAACCGCTGGCAACAAAGGATAAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATTTTAC
AACACGAGCTGACGACAGCCATGCAGCACCTGTCTCAGAGTTCCCGAAGGCACCAATCCATCTC
TGAAAGTTCTCTGGATGTCAAGAGTAGGTAAGGTTCTTCGCGTTGCATCGAATTAACCACAT
GCTCCACCGCTTGTGCGGGCCCCCGTCAATTCATTTGAGTTTTAACCTTGC GGCCGTACTCCCC
AGGCGGTGATTTAACGCGTTAGCTCCGGAAGCCACGCCTCAAGGGCACAACCTCCAAATCGAC
ATCGTTTACAGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGCACCTGA
GCGTCAGTCTTCGTCCAGGGGGCCGCTTTCGCCACCGGTATTCTCCAGATCTCTACGCATTTCC
ACCGCTACACCTGGAATTCTACCCCCCTCTACGAGACTCTAGCTTGCCAGTTTCAAATGCAGTT
```

CCCAGGGTTGAGCCCGGGGATTTACATCTGACTTAACAAACCCCTGCGTGCGCTTTACGCC  
 AGTAATTCCGATTAACGCTTGCACCCTCCGTATTACCGCGGCTGCTGGCACGGAGTTAGCCGGT  
 GCTTCTTCTGCGAGTAACGTCAATTGATGAGCGTATTAAGTTCACCACCTTCCTCCTCGCTGAA  
 AGTGCTTTACAACCCGAAGGCCTTCTTACACACACGCGGCATGGCTGCATCAGGCTTGCGCCCAT  
 TGTGCAATATTCCCCTGCTGCCTCCCGTAGGAGTCTGGACCGTGTCTCAGTTCCAGTGTGGC  
 TGGTCATCCTCTCAGACCAGCTAGGGATCGTTCGCTAGGTGAGCCATTACCCACCTACTAGCT  
 AATCCCATCTGGGCACATCTGATGGCAAGAGGCCCGAAGGTCCCCCTCTTTGGTCTTGCGACGT  
 TATGCGGTATTAGCTACCGTTTCCAGTAGTTATCCCCCTCCATCAGGCAGTTTCCAGACATTA  
 CTCACCCGTCCGCCGCTCGTCACCCAGGGAGCAAGCTCCCCTGTGCTACCGCTC

**Fig. (A-4.2) Sequence of isolated bacterial species (*Serratia marcescens* strain HL1)**



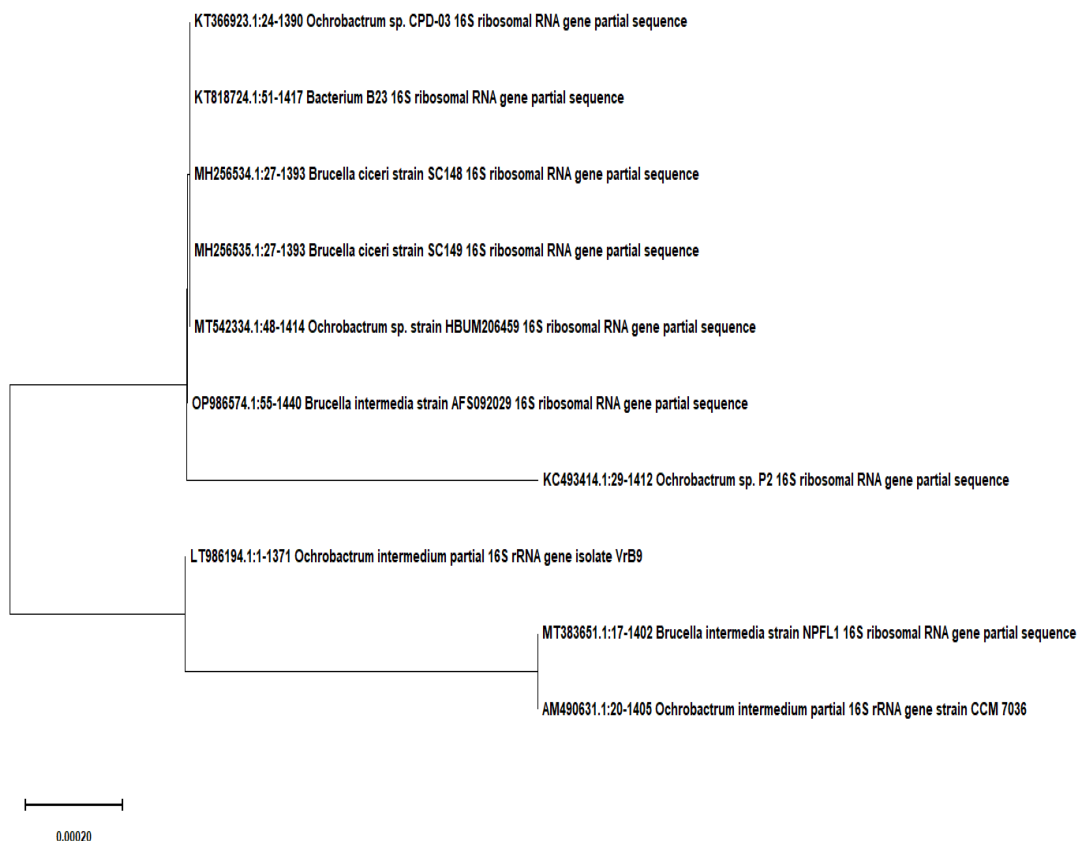
**Fig. (A-4.3) Phylogenetic tree of potential bacterial species *Serratia marcescens* strain HL 1 after the characterization the 16S rRNA technique**



**Fig. (A-4.4) Aligned Sequence (Data of Sample PCR amplification images)**

CCTGCCTCCTTGCGGTTAGCACAGCGCCTTCGGGTAAAACCAACTCCCATGGTGTGACGGGCGG  
TGTGTACAAGGCCCGGGAACGTATTCACCGCGGCATGCTGATCCGCGATTACTAGCGATTCCAA  
CTTCATGCACTCGAGTTGCAGAGTGCAATCCGAAGTGAAGATGGCTTTTGGAGATTAGCTCACAC  
TCGCGTGCTCGCTGCCCACTGTCACCACCATTTGTAGCACGTGTGTAGCCCAGCCCCGTAAGGGCC  
ATGAGGACTTGACGTCATCCCCACCTTCCTCTCGGCTTATCACCGGCAGTCCCCTTAGAGTGCC  
CAACTGAATGCTGGCAACTAAGGGCGAGGGTTGCGCTCGTTGCGGGACTTAACCCAACATCTCA  
CGACACGAGCTGACGACAGCCATGCAGCACCTGTCTCCGATCCAGCCGAAGTGAAGGATAGTGT  
CTCCACTAACCGCGATCGGGATGTCAAGGGCTGGTAAGGTTCTGCGCGTTGCTTCGAATTAAC  
CACATGCTCCACCGCTTGTGCGGGCCCCCGTCAATTCCTTTGAGTTTTAATCTTGCGACCGTAC  
TCCCCAGGCGGAATGTTAATGCGTTAGCTGCGCCACCGAAGAGTAAACTCCCCAACGGCTAAC  
ATTCATCGTTTTACGGCGTGGACTACCAGGGTATCTAATCCTGTTTGCTCCCCACGCTTTCGCAC  
CTCAGCGTCAGTAATGGTCCAGTGAGCCGCCTTCGCCACTGGTGTTCCTCCGAATATCTACGAA  
TTTCACCTCTACACTCGGAATTCACCTCACCTCTACCATACTCAAGACTAACAGTATCAAAGGC  
AGTTCGGGGTTGAGCCCCGGGATTTACCCCTGACTTATTAGCCCGCCTACGTGCGCTTTACG  
CCCAGTAAATCCGAACAACGCTAGCCCCCTTCGTATTACCGCGGCTGCTGGCACGAAGTTAGCC  
GGGGCTTCTTCTCCGGTTACCGTCATTATCTTCACCGGTGAAAGAGCTTTACAACCCTAGGGCC  
TTCATCACTCACGCGGCATGGCTGGATCAGGCTTGCGCCCATTGTCCAATATTCCCCCTGCTG  
CCTCCCGTAGGAGTCTGGGCCGTGTCTCAGTCCCAGTGTGGCTGATCATCCTCTCAGACCAGCT  
ATGGATCGTCGCCTTGGTAGGCCTTTACCCACCAACTAGCTAATCCAACGCGGGCCGATCATT  
TGCCGATAAATCTTTCCCCCCCATAAAATGCCAGGGCATTTTATGATTCCTGAAATTTTAAAGG  
GGGGCACATACGGTATTAGCACAAGTTTCCCTGAGTTATTCCGTAGCAAATGGTACGTTCCCAC  
GCGTACTCACCCGTCTGCCGCTCCCCTTGCGGGGCGCTCGA

**Fig. (A-4.5) Sequence of isolated bacterial species (*Ochrobactrum intermedium* VrB9)**



**Fig. (A-4.6) Phylogenetic tree of potential bacterial species *Ochrobactrum intermedium* VrB9 after the characterization the 16S rRNA technique**

#### Appendix 4 (b)

**Table 4 (A)**

| Bacterial strain | % removal |
|------------------|-----------|
| S1               | 56±1.23   |
| S2               | 45±2.32   |
| S1+S2            | 72±2.85   |

**S1: *Serratia marcescens* strain HL 1**

**S2: *Ochrobactrum intermedium* VrB9**

**S1+S2: *Serratia marcescens* strain HL 1 + *Ochrobactrum intermedium* VrB9**

## Appendix 4 (c)

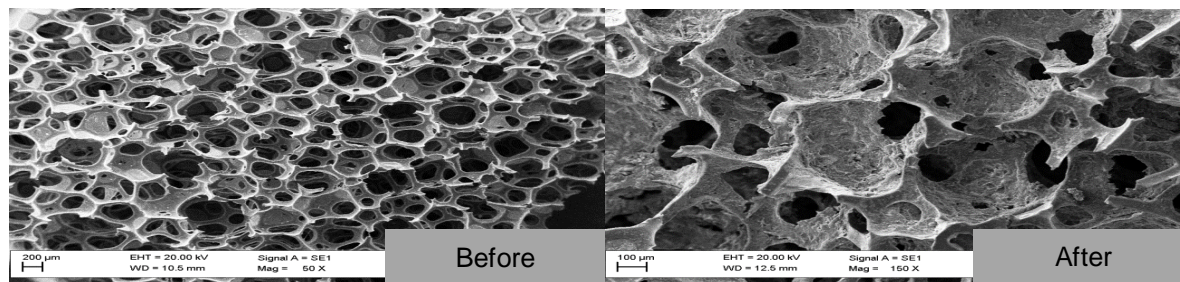


Fig. (A-4.7) SEM analysis of carrier before (0<sup>th</sup> day) and after formation (15<sup>th</sup> day) of biofilm

## Appendix 4 (d)

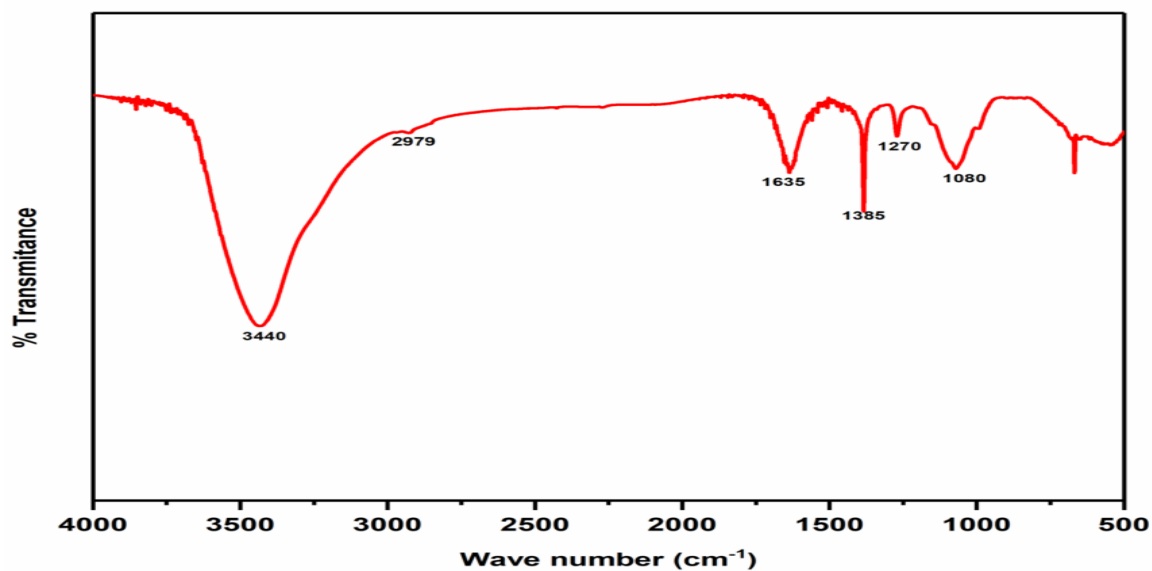
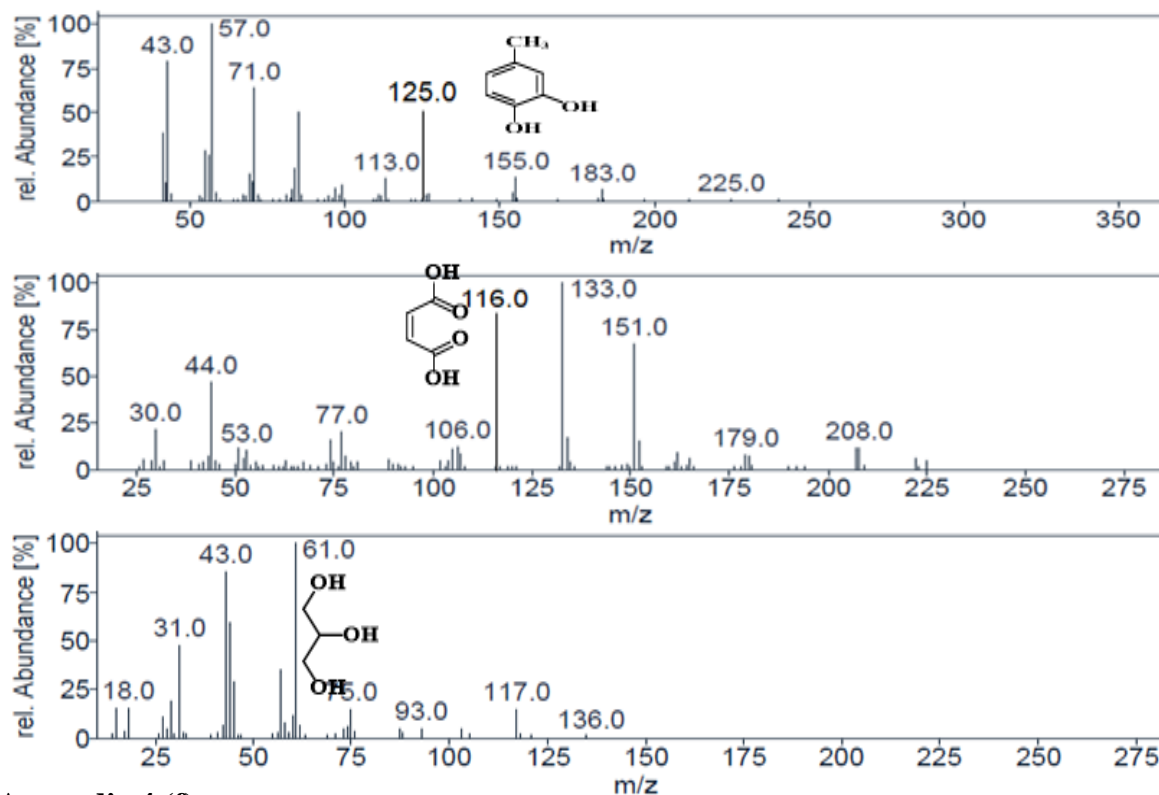


Fig. (A-4.8) FTIR spectra of biodegraded *p*-cresol

## Appendix 4 (e)



## Appendix 4 (f)

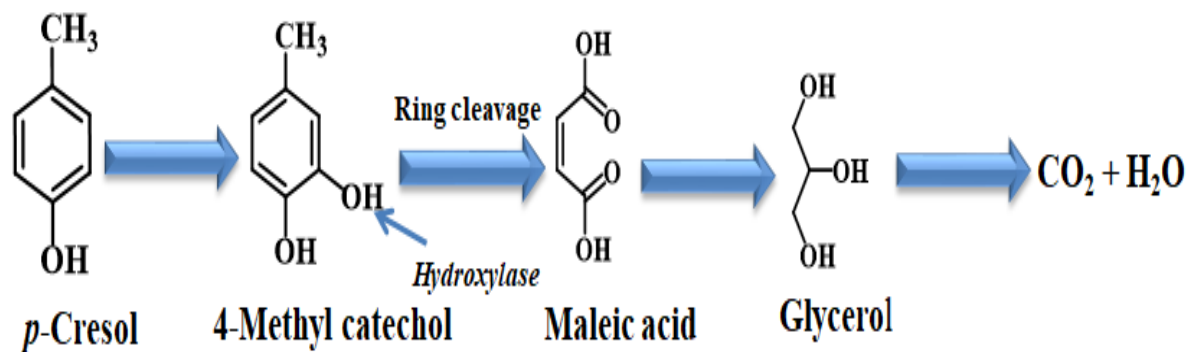


Fig. (A-4.9) Proposed metabolic path for the bacterial consortia to biodegrade *p*-cresol

## Appendix 4 (g)

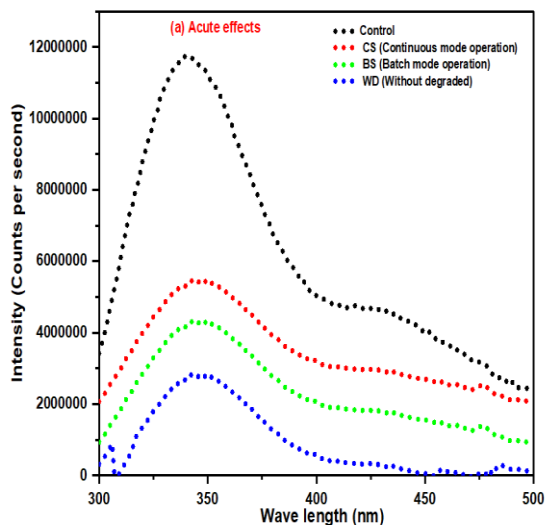


Fig. (A-4.10) Acute bioluminescence intensity of *p*-cresol (initial concentration  $500 \text{ mgL}^{-1}$ ) control (distilled water), CS, BS and WD *p*-

## Appendix 4 (h)

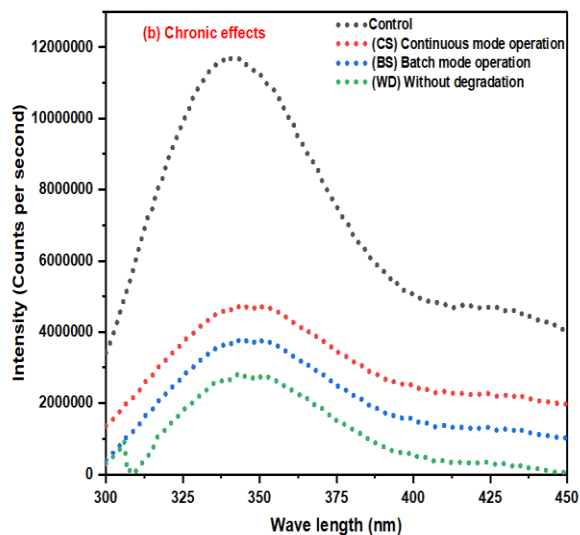


Fig. (A-4.11) bioluminescence intensity of *p*-cresol (initial concentration  $500 \text{ mgL}^{-1}$ ) control (distilled water), CS, BS and WD *p*-cresol

## Appendix 4 (i)

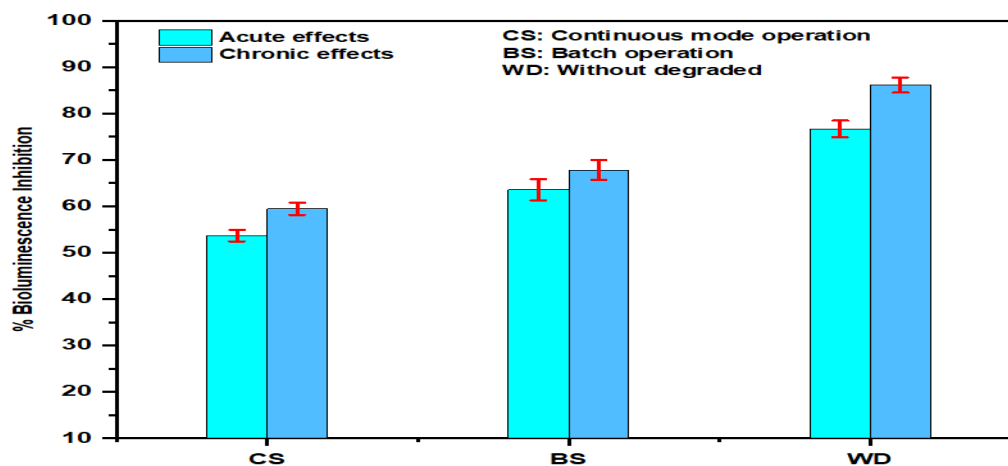
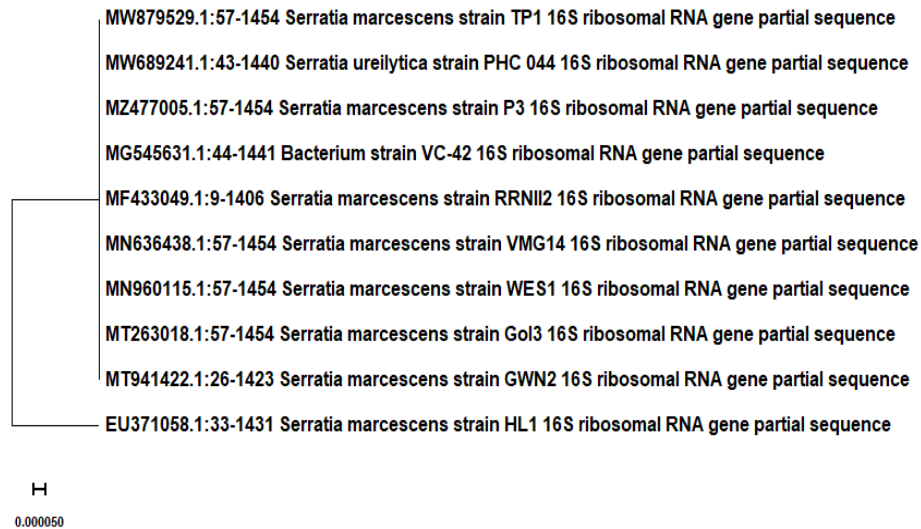


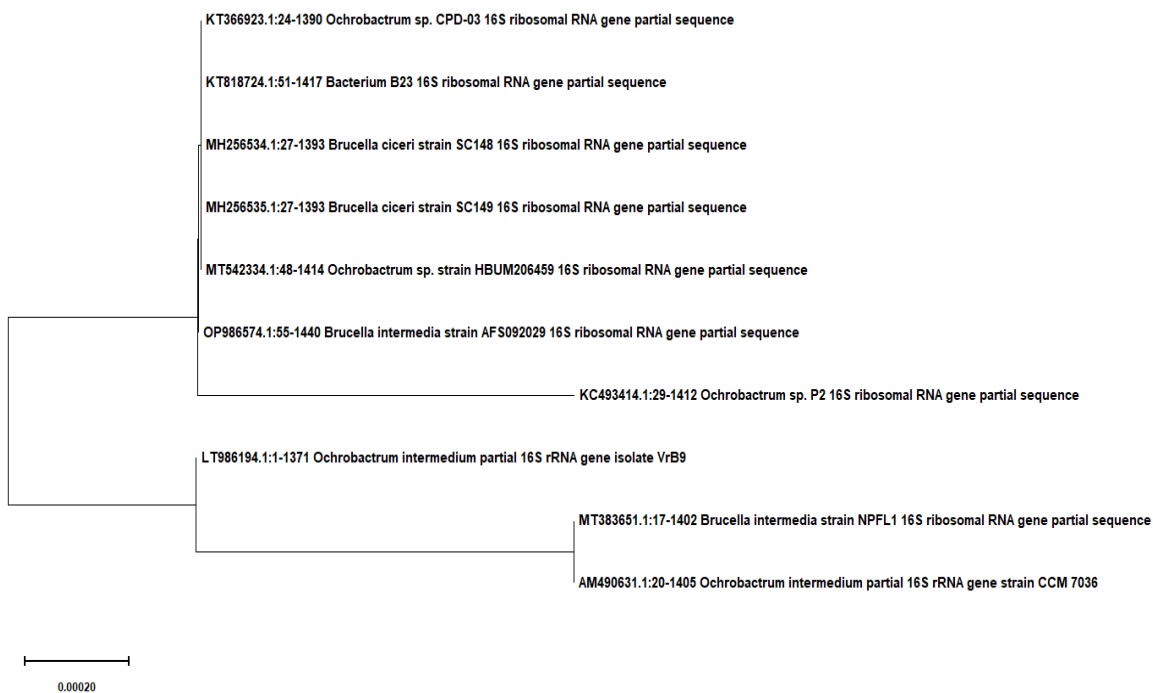
Fig. (A-4.12) Acute and chronic % bioluminescence inhibition of *p*-cresol (initial concentration  $500 \text{ mgL}^{-1}$ ) CS, BS and WD *p*-cresol

## Appendix 5 (a)



**Fig. (A-5.1) Phylogenetic tree of potential bacterial species *Serratia marcescens* strain HL 1 after the characterization of the 16S rRNA technique**

## Appendix 5 (b)



**Fig. (A-5.2) Phylogenetic tree of potential bacterial species *Ochrobactrum intermedium* VrB9 after the characterization the 16S rRNA technique**

## Appendix 5 (c)

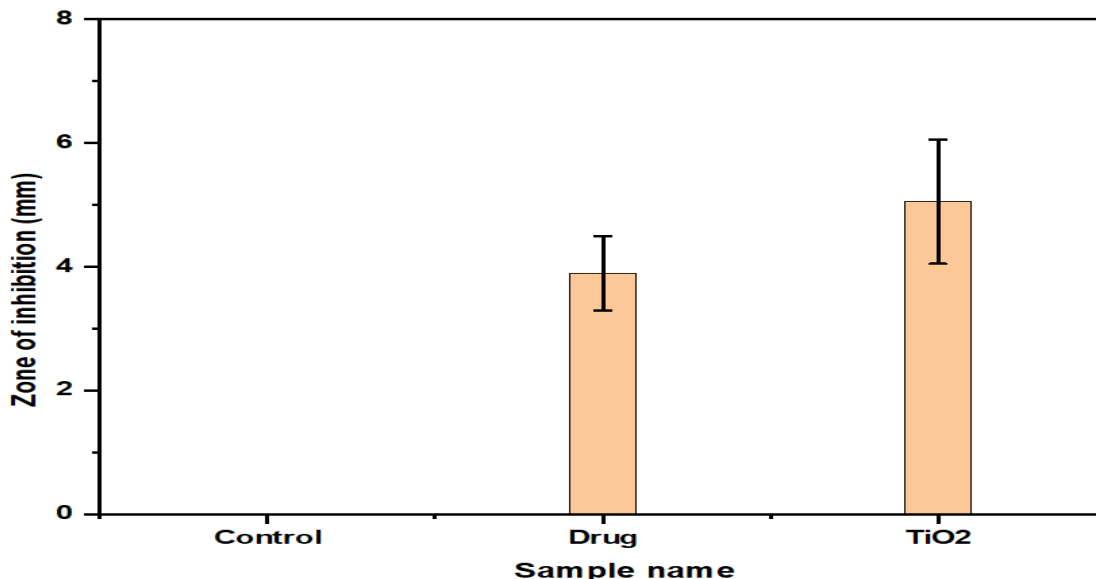


Fig. (A-5.3) Comparative study of inhibitory action of TiO<sub>2</sub> nanoparticles corresponding to gram positive (*S. aureus*) and gram-negative bacteria (*E. coli*)

## Appendix 5 (d)

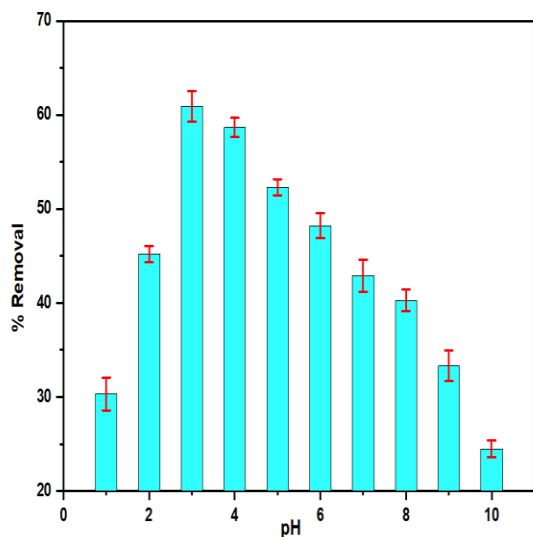


Fig. (A-5.4) Effect of pH on photocatalytic degradation of *p*-cresol (concentration: 100 mg L<sup>-1</sup>; Catalyst dose: 0.5g L<sup>-1</sup>, irradiation time 300 min)

## Appendix 5 (e)

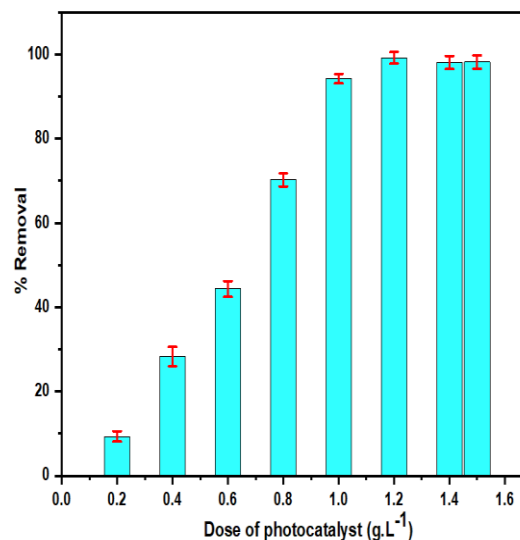


Fig. (A-5.5) Effect of dose of photocatalytic degradation of *p*-cresol (concentration: 100 mg L<sup>-1</sup>; pH: 7, irradiation time 300 min)

## Appendix 5 (f)

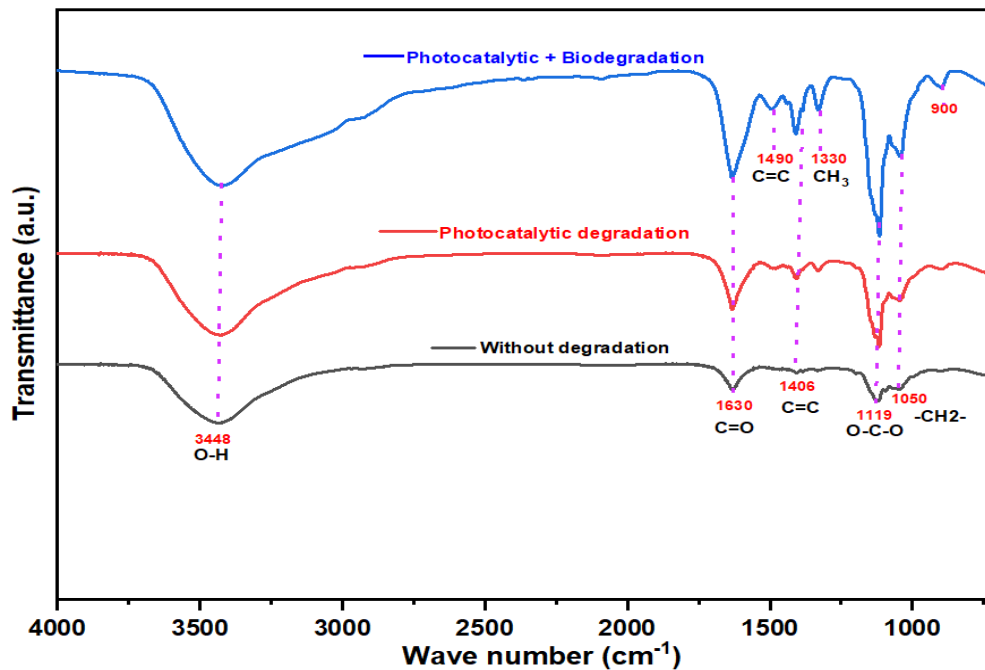


Fig. (A-5.6) FTIR spectra of *p*-cresol solution

### Appendix 5 (g)

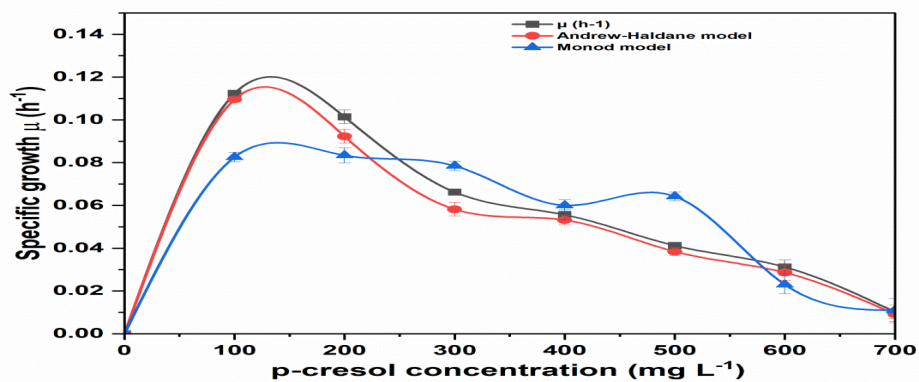
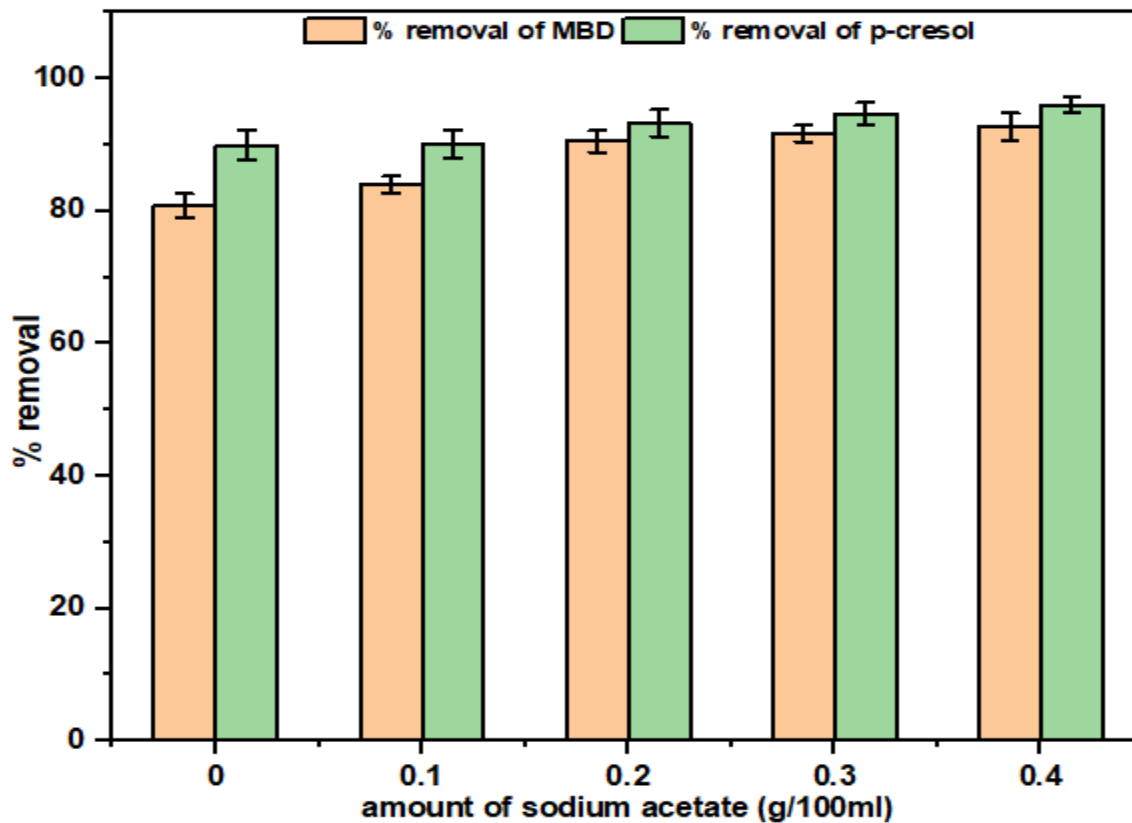


Fig. (A-5.7) Comparative study of experimental value with different Growth inhibition kinetic model

## Appendix 6 (a)



**Fig. (A-6.1) Effect of Sodium acetate on simultaneous % removal of *p*-cresol and methylene blue dye**



**List of Publications &  
Conferences/Workshops Attended**





1. **Jaiswal, V.K.**, Maurya, K.L., Sonwani, R.K., Singh, R.S., 2023. Biodegradation of p-cresol by *Serratia marcescens* strain HL 1 in batch system: Process optimization, growth kinetic study, phytotoxicity and chlorophyll assessment. **Bioresour. Technol. Reports** **22**, 101426. <https://doi.org/10.1016/j.biteb.2023.101426>
2. **Jaiswal, V.K.**, Sonwani, R.K., Singh, R.S., 2023. Construction and performance assessment of Recirculating packed bed biofilm reactor (RPBBR) for effective biodegradation of p-cresol from wastewater. **Bioresour. Technol.** 384, 129372. <https://doi.org/10.1016/j.biortech.2023.129372>
3. **Jaiswal, V.K.**, Dutta, A., Verma, V., Singh, R.S., 2023. Bioresource Technology Degradation of p-cresol in the presence of UV light driven in an integrated system containing photocatalytic and packed bed biofilm reactor. **Bioresour. Technol.** 387, 129706. <https://doi.org/10.1016/j.biortech.2023.129706>
4. **Jaiswal, V.K.**, Sonwani, R.K., Singh, R.S., 2023. Assessment of enhanced p-cresol biodegradation by encapsulating pre-immobilised *Serratia marcescens* strain HL 1 on tea waste biochar into polyvinyl alcohol / sodium alginate matrix. **Biochem. Eng. J.** 199, 109046. <https://doi.org/10.1016/j.bej.2023.109046>
5. **Jaiswal, V.K.**, Sonwani, R.K., Singh, R.S., 2024. Journal of Water Process Engineering Simultaneous removal of p -cresol and methylene blue dye through upward-flow packed bed biofilm reactor (UFPBBR): Kinetics , phytotoxicity and bacterial toxicity assessment. *J. Water Process Eng.* 58, 104868. <https://doi.org/10.1016/j.jwpe.2024.104868>
6. **Jaiswal, V.K.**, Gupta, A.D., Kushwaha, R., Kumar, R., Singh, K., Singh, H., Mohan, D., Singh, R.S., 2025. Arsenic removal from water using an acid-modified biochar. *J Mol Struct* 1324, 140904. <https://doi.org/10.1016/j.molstruc.2024.140904>
7. **Jaiswal, V.K.**, Gupta, A.D., Sonwani, R.K., Giri, B.S., Singh, R.S., 2024. Enhanced biodegradation of 2, 4-dichlorophenol in packed bed biofilm reactor by impregnation of polyurethane foam with Fe<sub>3</sub>O<sub>4</sub> nanoparticles: Bio-kinetics, process optimization, performance evaluation and toxicity assessment. **Bioresour Technol** 406. <https://doi.org/10.1016/j.biortech.2024.131085>

- ❑ International Conference on Biotechnology for Sustainable Bioresources and Bioeconomy (BSBB-2022).
- ❑ National conference on Environmental and Industrial Biotechnology (NCEIB-2022)
- ❑ Training Programme on Wastewater Characterization, Treatment and Reuse (WCTR - 2023)
- ❑ International Conference on Technologies and Innovations for Sustainable Development (TISD-2023)
- ❑ International Sustainability Conference On Health, Safety, Fire And Environmental Advances (HSFEA-2023)
- ❑ International conference on Advance and Innovation in Biotechnology for Sustainable Bioresource and Bioeconomy (AI-BSBB -2023)
- ❑ An International Conference on Energy Transition: Challenges and Opportunity (IICHE-CHEMCON-2023)